Serum Zonulin as a Marker of Intestinal Mucosal Barrier Function in Chronic Liver Diseases in Pediatrics

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ABSTRACT

Background: The term "chronic liver disease" (CLD) refers to a long-term, permanent alteration in the structure of the liver that may result in consequences including cirrhosis and early mortality. A potential contributing element to the pathophysiology of chronic liver disorders and the development of complications in cirrhosis is the dysfunction of the intestinal epithelial barrier. One indicator of intestinal permeability is zonulin.

Objective: This study aimed to assess serum zonulin levels in CLD with and without cirrhosis in pediatrics.

Patients and methods: The study population consisted of 40 children with CLD with cirrhosis, 40 children with CLD without cirrhosis and 20 apparently healthy children serving as a control group. Serum levels of zonulin was determined.

Results: The main finding of the current study was that serum zonulin levels were significantly higher in CLD with cirrhosis patients than in CLD without cirrhosis patients or healthy controls (p < 0.001). The median of serum zonulin levels in CLD with cirrhosis patients, CLD without cirrhosis patients, and healthy controls were 264.24, 8.24 and 39.69 ng/mL respectively.

Conclusion: Serum zonulin levels were significantly increased in liver cirrhosis patients.

Keywords: Serum zonulin, Intestinal Mucosal Barrier Function, CLD, Pediatrics.

INTRODUCTION

CLD is defined as the gradual destruction and regeneration of liver parenchyma over at least six months, resulting in fibrosis and cirrhosis. CLD is one of the leading causes of morbidity and mortality in pediatric^[1].

The integrity of the intestinal epithelium and the tight connection that closes the paracellular gap play a major part in the intestinal epithelium's vital function of separating luminal contents from the interstitium. The intestine tight junctions exhibit selectivity in permeability, which can be either triggered by luminal nutrients healthilv pathologically induced by pathogens, mucosal immune cells and cytokines, and the enteric nervous system. Numerous clinical disorders, both intestinal and systemic, are linked to compromised intestinal barrier function^[2].

called zonulin A protein affects the permeability of the small intestinal wall's tight junction in a reversible manner. Intestinal wall permeability may be accurately and conveniently measured using the serum levels of zonulin ^[3]. Research points to the migration of bacteria and their byproducts, including endotoxin, from the intestinal lumen into the bloodstream as a potential player in the aetiology of long-term liver disorders and the emergence of problems in cirrhosis. The intestinal epithelial barrier's malfunction may play a significant role in enabling bacterial translocation, in addition to changes in the immune system and gut microbiota^[4]. Therefore, The purpose of the research was to evaluate serum zonulin family peptides levels, as a marker of intestinal

mucosal barrier function, in CLD with and without cirrhosis in pediatrics.

PATIENTS AND METHODS Study population:

This was a case control study. The study population consisted of 40 children with CLD with cirrhosis, 40 children with CLD without cirrhosis and 20 apparently healthy children serving as a control group. All patients were recruited during a two-year period (2021-2023) from the Outpatient Clinic and Inpatient Ward of the Paediatric Hepatology, Gastroenterology, and Nutrition Department, National Liver Institute, Menoufia University.

Inclusion criteria: Subjects were matched for age and sex. Cirrhosis was identified by a biopsy or by combining clinical, endoscopic, and radiographic evidence of portal hypertension or cirrhosis.

Exclusion criteria: Children with allergy or sensitivity to gluten or adhere to a gluten-free diet, children with indications of gastrointestinal illness, children with current infection or inflammation, history of immunological abnormalities, and children with diabetes.

History, clinical examination, and investigations:

• Full history taking with stress on family history of similar conditions, consanguinity, presenting symptoms (onset, course, duration and associated symptoms), symptoms of hepatocellular decompensation e.g. ascites and hepatic

encephalopathy. Symptoms of vascular decompensation e.g. hematemesis and melena.

- Clinical examination with stress on jaundice, pallor, hepatomegaly, splenomegaly, ascites.
- Investigations were recruited from patients' files: Routine laboratory parameters, investigations according to suspected etiology, imaging studies, liver biopsy and esophagogastroduodenoscopy.

Blood Sampling: Initially, each subject's blood samples were centrifuged for 10 minutes at 1500 g. After that, the serum was separated and centrifuged again for three minutes at 4°C at 2000 g. After that, the supernatant was kept at -80 °C.

ELISA Measurements: A double-antibody sandwich ELISA Kit (Shanghai Korain Biotech Co., Ltd.) was used to measure the levels of serum zonulin. Serum zonulin concentrations were measured spectrophotometrically using an ELISA microtiter plate reader (ELX800, Biotech Instruments, Inc., USA, SN: 1502175), and the assay was carried out in accordance with the kit manufacturer's instructions.

Ethical approval: The work has been submitted and approved by Menoufia University's National Liver Institute's Ethics Committee. Each participant's parent gave written consent for the

study. Every phase of the study was conducted in accordance with the Helsinki Declaration.

Statistical Analysis

SPSS statistics version 25.0 was used for the statistical analysis. When presenting data for continuous variables, the means \pm SD or median values were used. Frequencies and percentages were used to display data for categorical variables. Depending on the kind of data, the ANOVA, Kruskall-Wallis, or Chi-square test were used to identify significant variations in the clinicopathological characteristics across groups. For normally distributed data, post hoc analyses were used to compare the group differences. P-values ≤ 0.05 were deemed statistically significant.

RESULTS

The study consisted of 40 children with CLD with cirrhosis (15 males and 25 females), 40 children with CLD without cirrhosis (18 males and 22 females) and 20 apparently healthy children (10 males and 10 females). The mean ages were 11.58 ± 3.31 , 11.25 ± 3.39 and 10.90 ± 3.14 years old respectively. There was no difference in age and sex among the three groups. However, hepatomegaly, splenomegaly, jaundice and pallor were significantly different among the studied groups (Table 1).

Table (1): Demographic data and clinical data								
Parameter	CLD	CLD Healthy		P-value				
	with cirrhosis	without cirrhosis	Controls					
	(GI, n= 40)	(GII, n= 40)	(GIII, n= 20)					
Age (year)								
Mean \pm SD	11.58 ± 3.31	11.25 ± 3.39	10.90 ± 3.14	0.751 ^{a, NS}				
Range (min-max)	5.00 - 17.00	5.00 - 17.00	6.0 - 17.0					
Sex [n (%)]								
Male	15 (37.5)	18 (45.0)	10 (50.0)	0.619 ^{b, NS}				
Female	25 (62.5)	22 (55.0)	10 (50.0)					
Splenomegaly [n (%)]								
No	6 (15.0)	35 (87.5)		<0.001 ^{b, HS}				
Yes	34 (85.0)	5 (12.5)						
Hepatomegaly [n (%)]								
No	22 (55.0)	33 (82.5)		0.008 ^{b, HS}				
Yes	18 (45.0)	7 (17.5)						
Jaundice [n (%)]								
No	15 (37.5)	32 (80.0)		<0.001 ^{b, HS}				
Yes	25 (62.5)	8 (20.0)						
Pallor [n (%)]								
No	29 (72.5)	39 (97.5)		0.002 ^{b, HS}				
Yes	11 (27.5)	1 (2.5)						
%: Percentage within disease group								
^a : ANOVA test; ^b : Pearson's X ² -test								

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Table (2) is a list of the laboratory features of the individuals under study. The three groups differed considerably in terms of AST, GGT, ALP, total bilirubin, direct bilirubin, albumin haemoglobin percentage, total leucocytic count, platelet count, and INR. AST, ALT, and GGT levels were elevated in CLD with cirrhosis patients relative to CLD without cirrhosis patients and control participants, whereas platelet count and albumin levels were significantly decreased. Patients with CLD and cirrhosis had higher total bilirubin levels and lower haemoglobin levels as compared to other groups. Notably, compared to CLD patients without cirrhosis or healthy controls, blood zonulin levels were considerably greater in patients with CLD who also had cirrhosis (p < 0.001). The range of serum zonulin levels in CLD with cirrhosis patients, CLD without cirrhosis patients, and healthy controls were 155.18 -499.39. 30.12 50.20 1.70 145.68 and ng/mL respectively (Figure _ _ 1).

 Table (2): Laboratory characteristics in studied groups

Biochemical parameters	CLD with	CLD without	Healthy Controls	P-value	Pairwise
	cirrhosis	cirrhosis	(GIII, n= 20)		comparisons
	(GI, n= 40)	(GII, n= 40)			
Zonulin (ng/mL)					p1<0.001 ^{HS}
Median (IQR)	264.24 (96.87)	8.24 (111.33)	39.69 (8.79)	<0.001 ^{a, HS}	p2<0.001 ^{HS}
Range (min-max)	155.18 - 499.39	1.70 - 145.68	30.12 - 50.20		p3=0.896 ^{NS}
ALT (U/L)					$p1=0.059^{NS}$
Median (IQR)	103.00 (291.25)	43.50 (98.75)	17.00 (6.00)	<0.001 ^{a, HS}	p2<0.001 ^{HS}
Range (min-max)	7.00 - 779.00	7.00 - 1920.00	9.00 - 23.00		p3<0.001 ^{HS}
AST (U/L)					p1<0.001 ^{HS}
Median (IQR)	168.50 (311.00)	39.00 (87.75)	21.00 (7.00)	<0.001 ^{a, HS}	p2<0.001 ^{HS}
Range (min-max)	18.00 - 1393.00	17.00 - 1755.00	9.00 - 28.00		p3<0.001 ^{HS}
ALP (U/L)					p1=0.018 ^s
Median (IQR)	233.50 (188.50)	135.50 (122.25)	93.50 (86.25)	<0.001 ^{a, HS}	p2<0.001 ^{HS}
Range (min-max)	21.00 - 966.00	90.00 - 596.00	48.00 - 240.00		p3<0.001 ^{HS}
GGT (U/L)					p1=0.015 ^s
Median (IQR)	70.00 (72.00)	43.50 (29.25)	19.50 (9.50)	<0.001 ^{a, HS}	p2<0.001 ^{HS}
Range (min-max)	15.00 - 328.00	13.00 - 563.00	12.00 - 31.00		p3<0.001 ^{HS}
Total bilirubin (mg/dL)					p1<0.001 ^{HS}
Median (IQR)	3.90 (4.25)	0.80 (0.30)	0.67 (0.32)	<0.001 ^{a, HS}	p2<0.001 ^{HS}
Range (min-max)	0.40 - 44.00	0.20 - 17.00	0.33 - 1.08		p3=0.335 ^{NS}
Direct bilirubin (mg/dL)					p1<0.001 ^{HS}
Median (IQR)	2.45 (3.47)	0.30 (0.28)	0.15 (0.07)	<0.001 ^{a, HS}	p2<0.001 ^{HS}
Range (min-max)	0.09 - 27.00	0.00 - 14.70	0.10 - 0.21		p3<0.001 ^{HS}
Albumin (g/dL)					p1<0.001 ^{HS}
Median (IQR)	3.10 (1.33)	4.10 (0.28)	4.90 (0.30)	<0.001 ^{a, HS}	p2<0.001 ^{HS}
Range (min-max)	1.60 - 4.40	3.70 - 5.00	4.00 - 5.10		p3<0.001 ^{HS}
Hb (g/dL)					р1<0.001 ^{НS}
Median (IQR)	10.00 (2.98)	12.35 (1.38)	13.05 (1.47)	<0.001 ^{a, HS}	p2<0.001 ^{HS}
Range (min-max)	7.00 - 14.00	8.00-15.00	11.90 - 15.50		p3=0.027 ^s
WBCs ($10^3 \text{ cell/}\mu\text{L}$)					p1=0.041 ^s
Median (IQR)	5.90 (3.75)	7.00 (3.08)	6.60 (2.38)	0.032 ^{a, S}	$p2=0.244^{NS}$
Range (min-max)	1.20 - 13.00	4.30 - 13.00	4.90 - 9.30		p3=0.847 ^{NS}
Platelets (10^3 cell/uL)					p1<0.001 ^{HS}
Median (IOR)	102.00 (77.00)	321.50 (145.50)	284.00 (119.50)	<0.001 ^{a, HS}	p2<0.001 ^{HS}
Range (min-max)	29.00 - 393.00	130.00 - 600.00	170.00 - 432.00		p3=0.839 ^{NS}
INR					p1<0.001 ^{HS}
Median (IOR)	1.50 (0.50)	1.00(0.05)	1.00 (0.07)	<0 001 ^{a, HS}	p2<0.001 ^{HS}
Range (min-max)	0.98 - 3.40	0.80 - 1.20	0.95 - 1.10	~0.001	$p_{3}=0.590^{NS}$
				1	1

IQR: Interquartile range (difference between 1st and 3rd quartile).

a: Kruskal-Wallis test; if significant multiple pairwise comparisons were adjusted by Dunn-Sidak post hoc test.

p1: p-value for the difference between group I and group II

p2: p-value for the difference between group I and group III

p3: p-value for the difference between group II and group III $% \mathcal{A}$

NS: Non significant at p-value ≥ 0.05 S: significant at p-value < 0.05

HS: Highly significant at p-value < 0.01



Figure (1): Serum zonulin levels in the studied groups.

DISCUSSION

The key results of the current study is serum zonulin levels were significantly higher in CLD with cirrhosis patients than in CLD without cirrhosis patients or healthy controls (p < 0~001). The range of serum zonulin levels in CLD with cirrhosis patients, CLD without cirrhosis patients, and healthy controls were 155.18 - 499.39, 1.70 - 145.68 and 30.12 - 50.20 ng/mL respectively.

Chronic liver illnesses have an impact on the growth and incidence of the gut microbiota. Through the "gut-liver axis," changes in the composition and quantity of gut microbiota have a direct impact on the immunological milieu of the liver ^[5, 6], which in turn affects the body's overall immune condition.

According to the current study, individuals with CLD and cirrhosis had considerably higher intestinal permeability. The breakdown of the intestinal barrier directly led to intestinal bacterial translocation. When pathogenic bacteria and their toxic byproducts, like lipopolysaccharide, enter the portal vein, the liver is immediately exposed to bacterial components that damage the liver ^[7, 8]. This causes the liver to produce proinflammatory factors like TNF- α , IL-6, and other immune mediators. The endotoxin that the unbalanced bacteria then continually produce causes systemic and intrahepatic inflammation, which in turn aggravates liver damage and dysbacteriosis, creating a vicious cycle that affects both the gut and the liver ^[9, 10]. Additionally, it was shown that the most severe dysbacteriosis happens during the decompensated phase and that the organisation of the gut microbiota gradually alters as liver cirrhosis progresses ^[11].

Intestinal permeability is indicated by zonulin. By actively controlling the intercellular tight connection, it aids the intestinal barrier in adapting to different physiological and pathological stimuli ^[12, 13]. In the presence of bacteria or gluten, the small intestine may exhibit an increase in the pathological secretion of

zonulin, which in turn may lead to an increase in intestinal permeability and a disruption of intestinal barrier function. This, in turn, may result in bacterial translocation and aberrant presentation of antigens to the intestinal submucosa, as well as an induction of inflammation and an adaptive immune response ^[14, 15].

Numerous autoimmune illnesses, malignancies, and neurological disorders, such as inflammatory bowel disease ^[16], type 1 diabetes ^[17], and multiple sclerosis ^[18], are caused by this pathogenic process mediated by zonulin $^{[12, 14]}$. We found that patients with cirrhosis had significantly higher serum zonulin levels SLD, which may indicate that the breakdown of the intestinal barrier brought on by the zonulin pathway dysfunction is a role in the progression of chronic liver disease. Wang et al. ^[19] findings indicated that zonulin levels were greater in liver cirrhosis patients, which are in line with our findings. As well, our results are consistent with Pietrukaniec et al. [3] who found a statistically significant difference in serum zonulin levels between the group of cirrhotic patients and the healthy controls. Furthermore, our findings are supported by the findings of Voulgaris et al. ^[20] who found that individuals with more severe chronic liver disease had greater serum zonulin levels proving the link between increasing intestinal permeability and liver cirrhosis.

In order to track the association between zonulin and chronic liver disease consequences, the current study can be extended by recruiting patients who satisfy the criteria to increase the sample size and by prolonging the follow-up observation period.

In conclusion: Serum zonulin levels were markedly elevated in patients with liver cirrhosis.

Conflict of interest: None.

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REFERENCES

- 1. Sharma A, Nagalli S (2022): Chronic liver disease: StatPearls. StatPearls Publishing. https://www.ncbi.nlm.nih.gov/books/NBK554597/
- Meoli L, Günzel D (2020): Channel functions of claudins in the organization of biological systems. Biomembranes, 29: 183344. https://doi.org/10.1016/j.bbamem.2020.183344.
- 3. Pietrukaniec M, Migacz M, Żak-Goląb A *et al.* (2019): Zonulin Family Peptide Levels in Ascites and Serum in Patients with Liver Cirrhosis: A Preliminary Study. Disease Markers, 19: 2804091. doi: 10.1155/2019/2804091.
- 4. Woodhouse C, Patel V, Singanayagam A *et al.* (2018): The gut microbiome as a therapeutic target in the pathogenesis and treatment of chronic liver disease. Alimentary Pharmacology & Therapeutics, 47: 192-202.
- 5. Compare D, Coccoli P, Rocco A *et al.* (2012): Gutliver axis: the impact of gut microbiota on nonalcoholic fatty liver disease. Nutrition, Metabolism and Cardiovascular Diseases, 22 (6): 471–476.
- 6. Ohtani N (2015): Microbiome and cancer. Sem Immunopathology, 37 (1): 65–72.
- 7. Brun P, Castagliuolo I, Leo V *et al.* (2007): Increased intestinal permeability in obese mice: new evidence in the pathogenesis of nonalcoholic steatohepatitis. American Journal of Physiology-Gastrointestinal and Liver Physiology, 292 (2): 518–525.
- 8. Frazier T, DiBaise J, McClain C (2011): Gut microbiota, intestinal permeability, obesity-induced inflammation, and liver injury. Journal of Parenteral and Enteral Nutrition, 35 (5): 14–20.
- **9.** Cirera I, Martin Bauer T, Navasa M *et al.* (2001): Bacterial translocation of enteric organisms in patients with cirrhosis. Journal of Hepatology, 34 (1): 32–37.
- 10. Zhou Y, Zheng T, Chen H et al. (2018): Microbial intervention as a novel target in treatment of non-

alcoholic fatty liver disease progression. Cellular Physiology and Biochemistry, 51 (5): 2123–2135.

- **11. Bajaj J, Betrapally N, Hylemon P** *et al.* (2015): Salivary microbiota reflects changes in gut microbiota in cirrhosis with hepatic encephalopathy. Hepatology, 62 (4): 1260–1271.
- **12.** Fasano A (2012): Intestinal permeability and its regulation by zonulin: diagnostic and therapeutic implications. Clinical Gastroenterology and Hepatology, 10 (10): 1096–1100.
- **13.** Queiroz D, Rocha A, Rocha G *et al.* (2006): Association between Helicobacter pylori infection and cirrhosis in patients with chronic hepatitis C virus. Digestive Diseases and Sciences, 51 (2): 370–373.
- 14. Fasano A (2011): Zonulin and its regulation of intestinal barrier function: the biological door to inflammation, autoimmunity, and cancer. Physiological Reviews, 91 (1): 151–175.
- **15.** Asmar R, Panigrahi P, Bamford P *et al.* (2002): Host-dependent zonulin secretion causes the impairment of the small intestine barrier function after bacterial exposure. Gastroenterology, 123 (5): 1607– 1615.
- **16. Turner J (2009):** Intestinal mucosal barrier function in health and disease. Nature Reviews Immunology, 9 (11): 799–809.
- **17.** Mojibian M, Chakir H, Lefebvre D *et al.* (2009): Diabetes specific HLA-DR-restricted proinflammatory T-cell response to wheat polypeptides in tissue transglutaminase antibody negative patients with type 1 diabetes. Diabetes, 58 (8): 1789–1796.
- **18.** Ochoa-Reparaz J, Mielcarz D, Ditrio L *et al.* (2009): Role of gut commensal microflora in the development of experimental autoimmune encephalomyelitis. Journal of Immunology, 183 (10): 6041–6050.
- **19.** Wang X, Li M, Niu Y *et al.* (2019): Serum zonulin in HBV-associated chronic hepatitis, liver cirrhosis, and hepatocellular carcinoma. Disease Markers, 19: 5945721. doi: 10.1155/2019/5945721.
- 20. Voulgaris T, Karagiannakis D, Hadziyannis E *et al.* (2021): Serum zonulin levels in patients with liver cirrhosis: Prognostic implications. World Journal of Hepatology, 13: 1394-1404.