Analytical and microscopical studies on the protective effect of ascorbic acid (vitamin C) and beta-carotene against the toxicity induced by fenitrothion on the liver of female albino rats

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Abstract

The organophosphate insecticide fenitrothion is a contact insecticide and selective acaricide. It is used as a fly, mosquito and cockroach, residual contact spray for farms and public health programs.

The objectives of the present study were to evaluate the toxicity of fenitrothion on the female rat and the possible protective effects of ascorbic acid (vitamin C) and beta–carotene as antioxidant agents against the toxicity induced by fenitrothion. Sixty of adult female albino rats were randomly assigned to six equal groups including control group and groups treated orally with ascorbic acid (10mg/kg b.w.), beta–carotene (10mg/kg b.w.) with or without fenitrothion (15.65 mg/kg b.w. i.e 1/16 LD₅₀) for 28 successive days.

Ingestion of fenitrothion caused a significant increase in ALT (alanine transferase), AST (aspartate transferase), and AP (alkaline phosphatase). It decreased significantly GL (glucose) level, AChE (acetyl cholinesterase) and GSH (glutathion reductase) activities, while, it had insignificant effects on TB (total bilirubine) and a slight decrease in TP (total protein). The histological study of female rat liver tissues by Hx & Eosin, P.A.S, and Methyl Green Pyronine revealed that, fenitrothion showed vascular and degenerative changes in the hepatic cells, Also, it caused a significant decrease in glycogen contents and depletion in of nucleic acids in hepatic cells.

Treatments with ascorbic acid and beta–carotene plus fenitrothion hasn’t been caused any significant changes in all parameters in serum of female rats. Treatment with ascorbic acid plus fenitrothion resulted a significant improvement in all parameters tested regarding to the histological study, while, beta-carotene plus fenitrothion showed the same improvement except in glycogen content in hepatic cells.


Refree : Prof ; Moustaf Ismail
Refree : Prof ; Hasan. Sabry
Introduction

The immense increase in agricultural productivity which has occurred since World War II owes much to the introduction of synthetic pesticides. The powerful insecticidal properties of the organophosphorous compounds promoted their widespread use for the control of agricultural pests and invertebrate vector of animals and human disease. Unfortunately, the problems of residual toxicity has led to significant damage to many ecosystems and concern that their presence in all types of food might present a carcinogenic hazard to man (Concon, 1988).

The organophosphate insecticide fenitrothion is a contact insecticide and selective acaricide of low ovicidal properties. It may also be used as a fly, mosquito, and cockroach residual contact spray for farms and public health programs [Worthing, 1983]. In the last years, the using of organophosphate insecticide in Egypt was increased; humans and non-target organisms are exposed to these pesticides, via food, water, and air. There is much toxicological effect due to the chemical. Fenitrothion is greater acetylcholinesterase (AChE) inhibitor than Malathion (Berber et al 1999).

Manisha et al (1999), reported that organophosphates had been shown to produce oxidative stress through the generation of free oxygen radicals. Dwivedi et al (1998), found that the hepatic antioxidant defense system, glutathion reductase (GSH). Comprising catalase and GSH peroxidase significantly increased in OP treated animals.

Hai et al (1995), reported that dichlorvos exposure significantly decreased glutathion peroxidase activity and has an inhibitory effect on acetylcholinesterase and induced changes characteristic of oxidative stress.

Łukaszewicz et al (1999), concluded that the changes in antioxidant enzyme activities and malondialdehyde concentration in the blood indicate hypoxia in the first period of intoxication (within 24 hours) and reoxidation in the later period. Early studies have been shown that an antioxidant has a direct effect on oxygen free radicals, which caused lipid peroxidant in the liver and also glutathion (GSH) precursor (Sen, 1999).

L-Ascorbic acid (Vitamin C) occurs widely in nature. Its use as oxygen scavengers and also as synergists in a wide variety of food products.

Ascorbic acid had been allocated an ADI “not limited” by the JECFA (FAO / WHO, 1981).

Stevenson et al (1995) concluded that, the ability of antioxidant (ascorbic acid) vitamin C and vitamin E to inhibit dieldrin – induced DNA synthesis suggested oxidative stress involved in the toxicity of this insecticides, while, the inability of these vitamins to prevent all hepatotoxic changes indicate other factors are also involved. Geetanjali et al, (1993) supported the protective role of ascorbic acid against the organophosphate dimethoate.

Beta-carotene (provitamin A) is mainly used as food colorant. It is effective quenchers of singlet oxygen and can act as antioxidants by preventing the formation of hydro – peroxides. It is not toxic to rats, dogs and humans. An ADI of 0.5 mg / kg b. w. has been allocated by the GECFA (FAO / WHO, 1974).

Salem et al. (1995), found that organophosphorus compounds toxicity
caused many degenerative and necrotic changes in the blood vessels as well as in the liver, lungs, kidney, intestines testicles, and siatic nerve. Also, they found that, the use of antioxidant vitamins A., D, E prove to be beneficial in over coming and minizing the pathological action of phospholan toxicity.

Minakata, et. al. (1995), found that, plasma component revealed appreciable changes in the concentrations of an acute – phase reactant protein and some vitamins that act as antioxidant , vitamin C and its radical increased by 1-5 and 1-7 fold of the control, restively after treatment of paraquat .

Do et al (1997), reported that, the inhibitory effect of organophosphate insecticide -dichlorvos- on acetylcholine esterase, induced changes Characteristic of oxidative stress .

Nabila et al (2001), reported that, treatment with antioxidant cystein or green tea plus pyriproxyfen showed a significant protective effect in the parameters of liver kidney functions regarding the biochemical, histolgical and histochemical studies.

The objectives of the present study were to evaluate the protective effect of the antioxidants ascorbic acid (vitamin C) and beta-carotene against toxicity induced by the insecticide fenitrothion on female rats.

Kadry et al (2001) reported that, fenitrothion showed significant inhibition in the ACHE and glutatione-S-transferase (GST) activities after exposure while AST, ALT, TB, creatinine and urea exhibited general significant elevation compared with the control values .

Materials and Methods

Materials:

Fenitrothion,[o,o-dimethylo-(3 –methyl 1 – 4 - nitrophenyl) phosphorothioate] (Fig. 1) was purchased in the form of emulisifiable concentrate formulation (50 EC) from Sumitomo Chemical Co. (Osaka)-Japan. The orally LD$_{50}$ of fenitrothion was determined according to Finney (1952). The selected dose was 15.65 mg / kg of body weight (equivalent of 1/16 of LD$_{50}$).

![Fenitrothion](image)

Fig (1): Fenitrothion.

L-Ascorbic acid (vitamin C), 3-keto-1-glucofur- anolactone (Fig. 2) was obtained in from of tablets from Memphis Co. Pharm. & Chemical Ind . (Cairo – A.R.E.).
Analytical and microscopical studies

![L-Ascorbic acid diagram]

Fig (2) : L-Ascorbic acid
Beta-carotene (provitamin A) (Fig. 3) was purchased from Memphis Co. Pharm. & Chemical Ind. (Cairo – A. R. E.)

![Beta-carotene diagram]

Fig (3): Beta-carotene

**a-Animals:**

Three months old, sexually mature female albino rats (derived from spragu – Dawley) weighing 150 ±10 g were purchased from animal house colony, Giza, Egypt. The animals were maintained on standard laboratory diet (protein : 16-04%; Fat : 3,63; Fiber : 401% and metabolic energy : 2887 kcal, kg) and water ad libitum for one week to aclimatize under the laboratory conditions, temp. (27±3°C–60±5RH). After acclimatization period, animals were divided into six groups (10 rats / group) and housed in glass cages (65 x 70 x 20 cm).

Experimental animals were given orally daily for 28 successive days; group 1, untreated control; group 2, were given orally fenitrothion (15.65 mg / kg b.w) equivalent 1/16 of LD$_{50}$; group 3, were given orally fenitrothion plus beta-carotene (10 mg / kg b.w) group 4, were given fenitrothion plus ascorbic acid with the same doses, group 5, were given ascorbic acid (vitamin C) (10 mg / kg b.w); group 6, were given orally beta-carotene (10 mg / kg b.w).

The doses were given at similar times each day, and adjusted as necessary to maintain its constant serum level in for body weight. During the period of administration , the animals were observed closely, each day for signs of toxicity .

After the end of the treatment period, blood samples were collected from rat tail veins and kept in heparinized tubes for biochemical analysis.
Biochemical analysis:
Acetyl cholinesterase (AChE) activity was determined in serum samples using 0.1 M phosphate buffer, pH 7.2 and acetylthiocholin iodide as substrate, according to the method of Ellman et al., (1961). Alanine transferase (ALT) and aspartate transferase (AST) were determined in serum according to the method described by Reitman and Frankel (1957). Total bilirubin (TB) was determined according to the method of Pearlman and Lee (1971).
Alkaline phosphatase (AP) and glutathion reductase (GSH) were determined in serum samples using 0.1 M carbonate buffer, pH 10.2 and diphenyl phosphate as substrate for (AP) and 0.2 M sodium phosphate buffer, pH 7.2 and 1-chloro-2,4-dinitrobenzene (CDNB) as a substrate for (GSH) according to the method of Powell and smith, (1954) and Habig et al. (1974), respectively. Also, total protein (TP) and serum glucose (GL) were determined according to the method of Bradford (1976) and Trinder (1969), respectively.

Histological studies:
All animals were sacrificed and dissected on day 28, livers, were exised and fixed in 10% neutral formalin. Paraffin sections of 7 micrometer thickness were prepared and stained by Hx and Eosin for morphological study (Drury and Wallington, 1980). P.A.S. (Pearse, 1968) to demonstrate glycogen content and Methyl Green Pyronin (Clayden, 1971) for nucleic acid content. Sections stained with PAS were subjected to analysis by using the image analyzer (Lecica image analyzer) and the obtained data were tabulated and statistically analyzed.

Statistical analysis:
All data were subjected to statistical analysis under standing Medical statistical methods by Leonard (1983).

Results

Biochemical analysis:
Effects of antioxidants [i.e. ascorbic acid (10 mg / kg b.w)] and [beta-carotene (10 mg / kg b.w)] as protectors and insecticide fenitrothion (15.65 mg/kg b.w) on serum biochemical analysis of female rats were studied.
Data in Table (1) and Fig. (4), showed that, oral administration with fenitrothion alone (15.65 mg / kg b.w) resulted a significant increase in aspartate transaminase (AST) activity compared with control (P < 0.05).
Treatment with fenitrothion plus ascorbic acid compared with fenitrothion alone resulted in a significant decrease in the level of AST in blood (P. value < 0.05), however, treatment with fenitrothion plus beta-carotene compared with fenitrothion alone resulted in a significant decrease in level of AST in blood (P < 0.05).
Treatment with ascorbic acid alone compared with control specimens resulted in significant increase in the level of AST in blood (P value < 0.05).
Treatment with beta-carotene alone compared with control specimens resulted in a significant increase in the level of AST in blood (P < 0.05).
Data in Table (2) and Fig. (5), showed that, oral administration with fenitrothion alone resulted in a significant increase in alanine transaminase (ALT) activity in blood when compared with control group (P < 0.05).
Treatment with fenitrothion plus ascorbic acid compared with fenitrothion alone resulted in a significant decrease in the level of ALT in blood (P < 0.05).

Table (1): Effect of fenitrothion alone and in combination with ascorbic acid and beta-carotene on AST activity in female rats.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control</th>
<th>Fenitrothion</th>
<th>Fenitro+ascorbic acid</th>
<th>Fenitro.+beta carotene</th>
<th>ascorbic acid</th>
<th>beta carotene</th>
</tr>
</thead>
<tbody>
<tr>
<td>AST (U/L)</td>
<td>26</td>
<td>43.33</td>
<td>11.67</td>
<td>26</td>
<td>22.33</td>
<td>29</td>
</tr>
<tr>
<td></td>
<td>17</td>
<td>30</td>
<td>17</td>
<td>36.33</td>
<td>25.33</td>
<td>30</td>
</tr>
<tr>
<td></td>
<td>17.33</td>
<td>26.67</td>
<td>12</td>
<td>18.33</td>
<td>26</td>
<td>27</td>
</tr>
<tr>
<td>Mean</td>
<td>20.11</td>
<td>33.33</td>
<td>13.56</td>
<td>26.89</td>
<td>24.55</td>
<td>28.67</td>
</tr>
<tr>
<td>S.D.</td>
<td>5.10</td>
<td>8.82</td>
<td>2.99</td>
<td>9.03</td>
<td>1.95</td>
<td>1.53</td>
</tr>
</tbody>
</table>

Gr.comp. C:Fenitro C:Fenitro+F+ascorbic acid C:Fenitro+F+beta carotene C:ascorbic acid C:beta carotene

T.Value 0.0146 0.1467 0.2180 0.1939 0.0503

P value <0.05 <0.05 <0.05 <0.05 <0.05

Signif. Sig. inc Sig. dec Sig. dec Sig. inc Sig. inc

Fig. (4): Effect of fenitrothion alone and in combination with ascorbic acid and beta-carotene on AST activity in female rats.
Table (2): Effect of fenitrothion alone and in combination with ascorbic acid and beta-carotene on ALT activity in female rats.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control</th>
<th>Fenitrothion</th>
<th>Fenitro+ascorbic acid</th>
<th>Fenitro.+beta carotene</th>
<th>ascorbic acid</th>
<th>beta carotene</th>
</tr>
</thead>
<tbody>
<tr>
<td>ALT (U/L)</td>
<td>20.67</td>
<td>34.6</td>
<td>22.66</td>
<td>19.67</td>
<td>12.83</td>
<td>24.5</td>
</tr>
<tr>
<td></td>
<td>13.67</td>
<td>21.67</td>
<td>13.66</td>
<td>27.5</td>
<td>21.5</td>
<td>17.67</td>
</tr>
<tr>
<td></td>
<td>15.5</td>
<td>24.66</td>
<td>21.67</td>
<td>24.67</td>
<td>17</td>
<td>24.5</td>
</tr>
<tr>
<td>Mean</td>
<td>16.61</td>
<td>26.98</td>
<td>19.33</td>
<td>23.95</td>
<td>17.11</td>
<td>22.22</td>
</tr>
<tr>
<td>S.D.</td>
<td>3.63</td>
<td>6.77</td>
<td>4.94</td>
<td>3.96</td>
<td>4.34</td>
<td>3.94</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Gr.comp.</th>
<th>C:Fenitro</th>
<th>Fenitro:F.+ascorbic acid</th>
<th>Fenitro:F.+beta carotene</th>
<th>C:ascorbic acid</th>
<th>C:beta carotene</th>
</tr>
</thead>
<tbody>
<tr>
<td>T.Value</td>
<td>0.0147</td>
<td>0.1298</td>
<td>0.1211</td>
<td>0.4615</td>
<td>0.0402</td>
</tr>
<tr>
<td></td>
<td>&lt;0.05</td>
<td>&lt;0.05</td>
<td>&lt;0.05</td>
<td>&lt;0.05</td>
<td>&lt;0.06</td>
</tr>
<tr>
<td>Sig. inc</td>
<td>Sig.dec</td>
<td>Sig.dec</td>
<td>Sig. inc</td>
<td>Sig. inc</td>
<td>Sig. inc</td>
</tr>
</tbody>
</table>

Fig. (5): Effect of fenitrothion alone and in combination with ascorbic acid and beta-carotene on ALT activity in female rats.

Treatment with fenitrothion plus beta-carotene compared with fenitrothion alone caused a significant decrease in level of ALT in blood (P < 0.05).

Treatment with ascorbic acid alone compared with control caused a significant increase in the level of ALT in blood (P < 0.05).

Treatment with beta-carotene alone compared with control specimens resulted in a significant increase in the level of ALT in blood (P < 0.05).
Also, the data in Table (3) and Fig. (6), showed a significant decrease in serum glucose (GL) content with control (P < 0.05).

Treatment with fenitrothion plus ascorbic acid compared with fenitrothion alone resulted in a significant increase in glucose level in blood when compared with control group (P < 0.05).

Treatment with fenitrothion plus beta-carotene compared with fenitrothion alone resulted in a significant increase in level of glucose in blood (P < 0.05).

Treatment with ascorbic acid alone compared with control caused a significant decrease in the level of blood glucose (P value < 0.05).

Treatment with beta-carotene alone compared with control specimens resulted in a significant decrease in the level of blood glucose (P < 0.05).

In Table (4) and Fig. (7), the date showed a significant increase in Total bilirubin (TB) content with fenitrothion alone compared with control (P. < 0.05)

Treatment with fenitrothion plus ascorbic acid compared with fenitrothion alone resulted in a significant decrease in total bilirubine (TB) level in blood when compared with control (P< 0.05).

Table (3): Effect of fenitrothion alone and in combination with ascorbic acid and beta-carotene on glucose (GL) level in female rats.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control</th>
<th>Fenitrothion</th>
<th>Fenitro+ascorbic acid</th>
<th>Fenitro.+beta carotene.</th>
<th>ascorbic acid</th>
<th>beta carotene</th>
</tr>
</thead>
<tbody>
<tr>
<td>Glucose (mg/dl)</td>
<td>104.58</td>
<td>60.17</td>
<td>97.58</td>
<td>101.55</td>
<td>97.46</td>
<td>96.27</td>
</tr>
<tr>
<td></td>
<td>121.7</td>
<td>51.36</td>
<td>104.31</td>
<td>95.95</td>
<td>72.04</td>
<td>90.68</td>
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<tr>
<td></td>
<td>115.33</td>
<td>55.09</td>
<td>94.59</td>
<td>93.58</td>
<td>89.75</td>
<td>85.92</td>
</tr>
<tr>
<td>Mean</td>
<td>113.87</td>
<td>55.54</td>
<td>98.83</td>
<td>97.03</td>
<td>86.42</td>
<td>90.96</td>
</tr>
<tr>
<td>S.D.</td>
<td>8.65</td>
<td>4.42</td>
<td>4.98</td>
<td>4.09</td>
<td>13.03</td>
<td>5.18</td>
</tr>
<tr>
<td>T.Value</td>
<td>0.0082</td>
<td>0.0134</td>
<td>0.0024</td>
<td>0.0778</td>
<td>0.0443</td>
<td></td>
</tr>
<tr>
<td>P value</td>
<td>&lt;0.05</td>
<td>&lt;0.05</td>
<td>&lt;0.05</td>
<td>&lt;0.05</td>
<td>&lt;0.06</td>
<td></td>
</tr>
<tr>
<td>Signif.</td>
<td>Sig.dec</td>
<td>Sig. inc</td>
<td>Sig. inc</td>
<td>Sig.dec</td>
<td>Sig.dec</td>
<td></td>
</tr>
</tbody>
</table>
Fig. (6): Effect of fenitrothion alone and in combination with ascorbic acid and beta-carotene on glucose (GL) level in female rats.

Table (4): Effect of fenitrothion alone and in combination with ascorbic acid and beta-carotene on total bilirubine (TB) in female rats.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control</th>
<th>Fenitrothion</th>
<th>Fenitro+ascorbic acid</th>
<th>Fenitro+beta carotene</th>
<th>ascorbic acid</th>
<th>beta carotene</th>
</tr>
</thead>
<tbody>
<tr>
<td>T.B(mg/dl)</td>
<td>1.5</td>
<td>1.51</td>
<td>1.29</td>
<td>1.4</td>
<td>0.69</td>
<td>1.32</td>
</tr>
<tr>
<td></td>
<td>1.18</td>
<td>1.33</td>
<td>1.03</td>
<td>1.58</td>
<td>1.32</td>
<td>1.03</td>
</tr>
<tr>
<td></td>
<td>1.33</td>
<td>1.26</td>
<td>1.57</td>
<td>1.65</td>
<td>1.1</td>
<td>1.57</td>
</tr>
<tr>
<td>Mean</td>
<td>1.34</td>
<td>1.37</td>
<td>1.30</td>
<td>1.61</td>
<td>1.04</td>
<td>1.31</td>
</tr>
<tr>
<td>S.D.</td>
<td>0.16</td>
<td>0.13</td>
<td>0.27</td>
<td>0.23</td>
<td>0.32</td>
<td>0.27</td>
</tr>
<tr>
<td>T.Value</td>
<td>0.3433</td>
<td>0.1580</td>
<td>0.3898</td>
<td>0.1956</td>
<td>0.4225</td>
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<tr>
<td>P value</td>
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<td>&lt;0.05</td>
<td>&lt;0.05</td>
<td>&lt;0.06</td>
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</tr>
<tr>
<td>Signif.</td>
<td>Sig. inc</td>
<td>Sig.dec</td>
<td>Sig. inc</td>
<td>Sig.dec</td>
<td>Sig.dec</td>
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</tr>
</tbody>
</table>

Fig. (7): Effect of fenitrothion alone and in combination with ascorbic acid and beta-carotene on total bilirubine (TB) in female rats.
Analytical and microscopical studies

Treatment with fenitrothion plus beta-carotene compared with fenitrothion alone caused a significant decrease in the activity of AP in blood (P < 0.05).

Treatment with ascorbic acid alone compared with control caused a significant increase in the activity of AP in blood (P value < 0.05).

Treatment with beta-carotene alone compared with control specimens results in a significant increase in the activity of AP in blood (P value < 0.05).

Data in Table (6) and Fig. (9), showed that, oral administration with fenitrothion alone resulted in a significant decrease in total protein (TP) content in blood when compared with control group (P value < 0.05).

Treatment with fenitrothion plus ascorbic acid compared with fenitrothion alone resulted in a significant increase in the activity of TP in blood (P < 0.05).

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control</th>
<th>Fenitrothion</th>
<th>Fenitro+ascorbic acid</th>
<th>Fenitro.+beta carotene.</th>
<th>ascorbic acid</th>
<th>beta carotene</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean</td>
<td>28.32</td>
<td>145.4</td>
<td>27.3</td>
<td>32.7</td>
<td>49.16</td>
<td>43.18</td>
</tr>
<tr>
<td></td>
<td>27.27</td>
<td>147.2</td>
<td>25</td>
<td>41.3</td>
<td>38.01</td>
<td>44.8</td>
</tr>
<tr>
<td></td>
<td>31.1</td>
<td>120.5</td>
<td>26.9</td>
<td>59</td>
<td>41.1</td>
<td>58.2</td>
</tr>
<tr>
<td>S.D.</td>
<td>1.98</td>
<td>14.92</td>
<td>1.23</td>
<td>13.41</td>
<td>5.76</td>
<td>8.24</td>
</tr>
<tr>
<td>Gr.comp.</td>
<td>C:Fenitro</td>
<td>Fenitro:F.+ascorbic acid</td>
<td>Fenitro:F.+beta carotene.</td>
<td>ascorbic acid</td>
<td>beta carotene</td>
<td></td>
</tr>
<tr>
<td>T.Value</td>
<td>0.0040</td>
<td>0.0041</td>
<td>0.0109</td>
<td>0.0291</td>
<td>0.0167</td>
<td></td>
</tr>
<tr>
<td>P value</td>
<td>&lt;0.05</td>
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<td>&lt;0.05</td>
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<tr>
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<td>Sig.dec</td>
<td>Sig.dec</td>
<td>Sig. inc</td>
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<td></td>
</tr>
</tbody>
</table>

Table (5): Effect of fenitrothion alone and in combination with ascorbic acid and beta-carotene on alkaline phosphatase (AP) activity in female rats.
Fig. (8): Effect of fenitrothion alone and in combination with ascorbic acid and beta-carotene on alkaline phosphatase (AP) activity in female rats.

Table (6): Effect of fenitrothion alone and in combination with ascorbic acid and beta-carotene on total protein (TP) in female rats.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control</th>
<th>Fenitrothion</th>
<th>Fenitro+ascorbic acid</th>
<th>Fenitro.+beta carotene.</th>
<th>ascorbic acid</th>
<th>beta carotene</th>
</tr>
</thead>
<tbody>
<tr>
<td>TP (g/dl)</td>
<td>5.47</td>
<td>4.57</td>
<td>6.31</td>
<td>5.06</td>
<td>5.34</td>
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<td></td>
<td>5.85</td>
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<td>6.76</td>
<td>6.76</td>
<td>5.77</td>
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<td></td>
<td>5.37</td>
<td>5.01</td>
<td>5.74</td>
<td>5.74</td>
<td>4.93</td>
<td>5.07</td>
</tr>
<tr>
<td>Mean</td>
<td>5.56</td>
<td>4.76</td>
<td>6.27</td>
<td>5.85</td>
<td>5.35</td>
<td>5.03</td>
</tr>
<tr>
<td>S.D.</td>
<td>0.25</td>
<td>0.23</td>
<td>0.51</td>
<td>0.86</td>
<td>0.42</td>
<td>0.46</td>
</tr>
<tr>
<td>T.Value</td>
<td>0.0374</td>
<td>0.1153</td>
<td>0.1935</td>
<td>0.0971</td>
<td>0.0547</td>
<td></td>
</tr>
<tr>
<td>P value</td>
<td>&lt;0.05</td>
<td>&lt;0.05</td>
<td>&lt;0.05</td>
<td>&lt;0.05</td>
<td>&lt;0.06</td>
<td></td>
</tr>
<tr>
<td>Signif.</td>
<td>Sig.dec</td>
<td>Sig. inc</td>
<td>Sig. inc</td>
<td>Sig.dec</td>
<td>Sig.dec</td>
<td></td>
</tr>
</tbody>
</table>
Treatment with fenitrothion plus beta-carotene compared with fenitrothion alone caused a significant increase in the content of TP in blood (P < 0.05).

Treatment with ascorbic acid alone compared with control caused in a significant decrease in the content of TP in blood (P < 0.05).

Treatment with beta-carotene alone compared with control caused in a significant decrease in the level of TP in blood (P < 0.05).

Data in Table (7) and Fig. (10), showed that, oral administration with fenitrothion alone resulted in a significant decrease in specific activity of AChE (S.A) in blood when compared with control group (P value < 0.05).

Treatment with fenitrothion plus ascorbic acid compared with fenitrothion alone resulted in a significant increase in the S.A of AChE in blood (P < 0.05).

Table (7): Effect of fenitrothion alone and in combination with ascorbic acid and beta-carotene on specific activity (S.A) of acetyl cholinesterase (AChE) in female rats.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control</th>
<th>Fenitrothion</th>
<th>Fenitro+ascorbic acid</th>
<th>Fenitro.+beta carotene.</th>
<th>ascorbic acid</th>
<th>beta carotene</th>
</tr>
</thead>
<tbody>
<tr>
<td>AChE (S.A)*</td>
<td>23.85</td>
<td>15.45</td>
<td>25.33</td>
<td>22.19</td>
<td>29.15</td>
<td>24.27</td>
</tr>
<tr>
<td></td>
<td>23.05</td>
<td>16.25</td>
<td>25.65</td>
<td>20.91</td>
<td>29.65</td>
<td>24.15</td>
</tr>
<tr>
<td></td>
<td>22.75</td>
<td>16.95</td>
<td>24.85</td>
<td>21.55</td>
<td>28.95</td>
<td>23.97</td>
</tr>
<tr>
<td>Mean</td>
<td>23.22</td>
<td>16.22</td>
<td>25.28</td>
<td>21.55</td>
<td>29.25</td>
<td>24.13</td>
</tr>
<tr>
<td>S.D.</td>
<td>0.57</td>
<td>0.75</td>
<td>0.40</td>
<td>0.64</td>
<td>0.36</td>
<td>0.15</td>
</tr>
<tr>
<td>T.Value</td>
<td>0.0057</td>
<td>0.0008</td>
<td>0.0021</td>
<td>0.0020</td>
<td>0.0335</td>
<td></td>
</tr>
<tr>
<td>P value</td>
<td>&lt;0.05</td>
<td>&lt;0.05</td>
<td>&lt;0.05</td>
<td>&lt;0.05</td>
<td>&lt;0.06</td>
<td></td>
</tr>
<tr>
<td>Signif.</td>
<td>Sig.dec</td>
<td>Sig. inc</td>
<td>Sig. inc</td>
<td>Sig. inc</td>
<td>Sig. inc</td>
<td></td>
</tr>
</tbody>
</table>

* S.A : Specific activity (n mole/min./mg protein)
Table (8): Effect of fenitrothion alone and in combination with ascorbic acid and beta-carotene on specific activity (S.A) of glutathion reductase (GSH) in female rats.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control</th>
<th>Fenitrothion</th>
<th>Fenitro+ascorbic acid</th>
<th>Fenitro.+beta carotene.</th>
<th>ascorbic acid</th>
<th>beta carotene</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>GSH (S.A)</strong></td>
<td>2.35</td>
<td>1.51</td>
<td>3.25</td>
<td>2.85</td>
<td>3.41</td>
<td>3.11</td>
</tr>
<tr>
<td></td>
<td>2.29</td>
<td>1.8</td>
<td>3.12</td>
<td>2.81</td>
<td>2.92</td>
<td>2.9</td>
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<tr>
<td></td>
<td>2.11</td>
<td>1.65</td>
<td>3.09</td>
<td>2.69</td>
<td>3.45</td>
<td>2.85</td>
</tr>
<tr>
<td><strong>Mean</strong></td>
<td>2.25</td>
<td>1.65</td>
<td>3.15</td>
<td>2.78</td>
<td>3.26</td>
<td>2.95</td>
</tr>
<tr>
<td><strong>S.D.</strong></td>
<td>0.12</td>
<td>0.15</td>
<td>0.09</td>
<td>0.08</td>
<td>0.30</td>
<td>0.14</td>
</tr>
<tr>
<td><strong>Gr.comp.</strong></td>
<td>C:Fenitro</td>
<td>Fenitro:F.+ascorbic acid</td>
<td>Fenitro:F.+beta carotene.</td>
<td>C:ascorbic acid</td>
<td>C:beta carotene</td>
<td></td>
</tr>
<tr>
<td><strong>T.Value</strong></td>
<td>0.0197</td>
<td>0.0112</td>
<td>0.0068</td>
<td>0.0197</td>
<td>0.0022</td>
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</tr>
<tr>
<td><strong>P value</strong></td>
<td>&lt;0.05</td>
<td>&lt;0.05</td>
<td>&lt;0.05</td>
<td>&lt;0.05</td>
<td>&lt;0.06</td>
<td></td>
</tr>
<tr>
<td><strong>Signif.</strong></td>
<td>Sig.dec</td>
<td>Sig. inc</td>
<td>Sig. inc</td>
<td>Sig. inc</td>
<td>Sig. inc</td>
<td></td>
</tr>
</tbody>
</table>

* S.A : Specific activity (n mole/min./mg protein)
Treatment with fenitrothion plus beta-carotene compared with fenitrothion alone caused a significant increase in S.A of GSH in blood (P < 0.05).

Treatment with ascorbic acid alone compared with control caused a significant increase in S.A of GSH in blood (P < 0.05).

Treatment with beta-carotene alone compared with control specimens resulted in a significant increase in S.A of GSH in blood (P < 0.05).

**Histological studies**

Histological section of the liver of control rat stained with HX and Eosin showed the central vein with radiating hepatic cords (Fig. 12 A). However, after administration of the insecticide fenitrothion there was congestion of the blood vessels of the liver as demonstrated by dilated central vein and hepatic sinusoids with Hemorrhagic areas between the hepatic cells (Fig. 12 B). Moreover, there are evidence of degeneration of hepatic cells in some areas of the hepatic cells (Fig. 12 C).

With combination of the insecticide with beta-carotene there is relative decrease or improvement of the vascular dilation without improvement in the degeneration seen in some areas of the hepatic lobules. There are plenty of von kuppfer cells (Fig. 12 D).

On the other hand, combination of the insecticide with ascorbic acid causes relative improvement in the degeneration of the hepatic cells which returns to nearly normal pictures without improvement in the vascular dilatation (Fig. 12 E).

However, treatment with beta-carotene alone causes no vascular or cellular changes in hepatic cells (Fig. 12 F). The same results were obtained with treatment with ascorbic acid alone (Fig. 12 F).

Staining of sections of liver with P.A.S. stain showed normal distribution of glycogen content in the hepatic cells (Fig. 13 A).
Fig. (12): HX / Eosin Stain

A) A photomicrograph of control liver demonstrating a central vein with radiating hepatic cords, (HX & E X200).

B) A Photomicrograph of rat liver treated with fenitrothion showing dilated central vein with hemorrhagic area between hepatic cells (HX & E X200).

C) A photomicrograph of the previous group showing vascular dilatation and degeneration in some areas of hepatic cell (HX & E X200).

D) A Photomicrograph of rat liver treated with fenitrothion plus beta-carotene showing a decrease in vascular dilatation & degeneration of some hepatic cell (HX & E X200).

E) A Photomicrograph of rat liver treated with fenitrothion plus ascorbic acid showing vascular dilatation and improvement of degeneration of hepatic cell (HX & E X200).

F) A Photomicrograph of rat liver treated with ascorbic acid alone showing a central vein with radiating hepatic cords (as the control), the same result as with beta-carotene (HX & E X200).
optical density (O.D.) of glycogen in control groups was 0.524 (S.D ± 0.066 and S.E. 0.038) . Table (9) and Fig. (14).

After administration of fenitrothion there is depletion of the glycogen content of hepatic cells after staining with P.A.S stain (Fig.13B). The optical density (O.D) of glycogen group was 0.319 (S.D. 0.024, S.E, 0.014) Table (9) and Fig. (14). Combination of insecticide with beta-carotene causes no improvement but only decrease in glycogen content of hepatic cells with still relative vascular dilatation (Fig. 13 C). The optical density (O.D) of glycogen was 0.262, S.D = 0.050, S.E = 0.029 Table (9) and Fig. (14).

On the other hand combination of the insecticide with ascorbic acid causes more relative improvement or increase in glycogen content of hepatic cells (Fig. 13 D). The optical density (O.D) of glycogen was 0.476, S.D = 0.073, S.E = 0.042 . Table (9) and Fig. (14). Treatment with beta-carotene alone causes no change in glycogen content of hepatic cells (Fig. 13 E). The optical density (O.D) of glycogen was 0.393, S.D = 0.060, S.E 0.034 . Table (9) and Fig. (14).

The same results were obtained with treatment with ascorbic acid alone (Fig. 13 F). The optical density (O.D) of glycogen was 0.452 S.D = 0.060, S.E ± 0.034 . Table (9) and Fig. (14).

However comparison between the optical density (O.D) in control specimens and specimens received the insecticide revealed a T. value = 0.007 with a P = < 0.05 meaning a significant decrease in glycogen content compared to control group Table (9).

Also, comparison between the optical density (O.D) in specimens received insecticide and specimens received insecticide with beta-carotene revealed a T. value = 0.157 with a P = < 0.05 meaning a significant decrease in glycogen content compared to specimens received insecticide . Comparison between the optical density (O.D) in specimens
Fig. (13): P.A.S. Stain

A) A photomicrograph of control liver demonstrating a normal distribution of glycogen content (P.A.S. Stain X200).

B) A photomicrograph of rat liver treated with fenitrothion showing relative decrease in glycogen content of hepatic cells (P.A.S. Stain X200).

C) A photomicrograph of rat liver treated with fenitrothion plus bete-carotene showing a slight increase in glycogen content and relative vascular dilatation (P.A.S. Stain X200).

D) A photomicrograph of rat liver treated with fenitothion plus ascorbic acid showing a relative increase in amount of glycogen in hepatic cells (P.A.S. Stain X200).

E) A photomicrograph of rat liver treated with beta-carotene alone showing a normal distribution of glycogen content (as control) (P.A.S. Stain X200).

F) A photomicrograph of rat liver treated with ascorbic acid alone showing the same result of control (P.A.S. Stain X200).
Analytical and microscopical studies

Table (9): Changes in the glycogen content in the rat's liver under the effect of treatment with fenitrothion alone and combination with ascorbic acid and beta-carotene

<table>
<thead>
<tr>
<th></th>
<th>Control</th>
<th>Fenitrothion</th>
<th>Insec. + Beta carotene</th>
<th>Insec. + Ascorbic acid</th>
<th>Beta carotene</th>
<th>Ascorbic acid</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sample 1</td>
<td>0.532</td>
<td>0.324</td>
<td>0.253</td>
<td>0.433</td>
<td>0.425</td>
<td>0.430</td>
</tr>
<tr>
<td>2</td>
<td>0.453</td>
<td>0.292</td>
<td>0.315</td>
<td>0.433</td>
<td>0.430</td>
<td>0.443</td>
</tr>
<tr>
<td>3</td>
<td>0.585</td>
<td>0.340</td>
<td>0.217</td>
<td>0.561</td>
<td>0.325</td>
<td>0.483</td>
</tr>
<tr>
<td>Mean</td>
<td>0.524</td>
<td>0.319</td>
<td>0.262</td>
<td>0.476</td>
<td>0.393</td>
<td>0.452</td>
</tr>
<tr>
<td>S.E.</td>
<td>0.066</td>
<td>0.024</td>
<td>0.050</td>
<td>0.073</td>
<td>0.060</td>
<td>0.060</td>
</tr>
<tr>
<td>T.value</td>
<td></td>
<td>0.007</td>
<td>0.157</td>
<td>0.021</td>
<td>0.101</td>
<td>0.073</td>
</tr>
<tr>
<td>P.Value</td>
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<td>Sig.dec. &lt; 0.05</td>
<td>Sig.inc. &lt; 0.05</td>
<td>Non-Sig.</td>
<td>Non-Sig.</td>
<td></td>
</tr>
</tbody>
</table>

![Fig. (14) Changes in the glycogen content under the effect of treatment with fenitrothion alone and combination with ascorbic acid and beta-carotene](image)

Received insecticide and specimens received insecticide with ascorbic acid revealed a T. value = 0.021 and a P value = < 0.05 meaning a significant increase in glycogen content.

Comparison between the optical density (O.D) in control specimens and specimens received beta-carotene revealed a T. value = 0.101 and a P value indicating non significant change in glycogen content.

Comparison between the optical density (O.D) in control specimens and specimens received ascorbic acid revealed a T value = 0.073 and a P value meaning non significant change in glycogen content.

Staining control specimens with methyl green pyronin to detect nucleic acid control shows its normal distribution in hepatic cells (Fig. 15 A). However administration of insecticide fenitrothion cause decrease in the nucleic acid content of hepatic cells (Fig. 15 B).

Combination of the insecticide with beta-carotene causes no improvement and there is still also
depletion of nucleic acid content of the hepatic cells (Fig. 15 C). However, combination of the insecticide with ascorbic acid causes no change in the depleted amount of nucleic acid (Fig. 15 D).

Administration of beta-carotene alone causes no change in nucleic acid content of hepatic cells compared with control specimens (Fig. 15 E). However, the same results were obtained after administration of ascorbic acid alone (Fig. 15 F).

Fig. (15): M.G.P Stain

A) A photomicrograph of control rat liver demonstrating a normal presence of nucleic acid content in hepatic cells (M.G.P. Stain X250).
B) A Photomicrograph of rat liver treated with fenitrothion showing relative depletion of nucleic acid content in hepatic cells (M.G.P. Stain X250).
C) A photomicrograph of rat liver treated with fenitrothion plus beta-carotene showing a decrease in nucleic acid content in hepatic cells (M.G.P. Stain X250).
D) A Photomicrograph of rat liver treated with fenitrothion plus ascorbic acid showing a decrease in nucleic acid content in hepatic cells (M.G.P. Stain X250).
E) A Photomicrograph of rat liver treated with beta-carotene alone showing a normal nucleic acid content in hepatic cells (as control) (M.G.P. Stain X250).
F) A Photomicrograph of rat liver treated with ascorbic acid alone showing a normal content of nucleic acid in hepatic cells (as control) (M.G.P. Stain X250).
Analytical and microscopical studies

Discussion

Several investigators have been reported that the application of fenitrothion as organophosphorus compound has been used extensively as insecticide and acaricide for control of a wide range of pests [Worthing, 1983 and 1987]. Therefore, human and animals may be subjected to such compound via food, water and air and this could affect their health as demonstrated by Yamanaka et al. (1993). In the present study, the toxicity of fenitrothion was evaluated in female rats as a laboratory animal model and the possible protective effects of certain antioxidants (i.e ascorbic acid and beta-carotene) against this toxicity were also evaluated.

The present results of the estimated biochemical parameters indicated that, ingestion of fenitrothion caused a significant increase in ALT, AST, and AP activities, these results may indicate degenerative changes and hypofunction of liver, Justin et. al, 1991, Gomes et al 1999, and Kadry et al, 2001. The increasing of AP activity may be due to the induction of such enzyme as a response to the remained amounts of fenitrothion and its metabolites which needs more hydrolysis of phosphate bond to be able to eliminate. Furthermore, it is well documented that, the elevation in serum alkaline phosphatase (AP) indicates that there is an impairment of liver function and obstruction of flow (Lykasova and Rabinovich; 1988, Stevens and Galla, 1989). The results also showed that treatment with fenitrothion decreased significantly serum level of AChE, GL and GSH, while, it had insignificant effects on TB and it caused a slight decrease in TP. The inhibitory effects on AChE was expected, as the organophosphate compounds are anticholinesterase agents such finding was reported since, 1950’s by many investigators [Barber et al 1999, Gome, et al 1999, Nigg and Knaak 2000].

The decreasing effect of fenitrothion on GSH is supported by the fact that glutathion, S-transferase is used in catalyzing the conjugation process of fenitrothion and / or its metabolites with glutathion as starting point of the limination process.

Furthermore, this explanation was in harmony with other results obtained by Motoyame and Dauterman, (1978); Matsumora, 1985; Datta et al (1997); Ecobicon, 1996, and Kadry et al. 2001. The slight increase in TB may be due to the effect of fenitrothion on liver functions [stevens and Gallo 1989; Paulino et al; 1996, and Nabila et al, 2001].

Thus, the present results clearly indicated that fenitrothion has a harmful and stressful influence on liver’s functions. Also, Manisha et al, 1999; reported that organophosphates have been shown to produce oxidative stress through the generation of free oxygen radical.

It is of interest to mention that, the treatment of female rats with fenittrothion plus beta-carotene or ascorbic acid didn’t cause any significant change in AST and ALT activities. The same result, were sighted in AChE, glucose, TB and TP. Ascorbic acid plus fenitrothion was more efficient in formation of AP than beta-carotene plus fenitrothion. It is important to notice that GSH concentration was increased in female rats after treatment with fenitrothion combined with ascorbic acid and beta-carotin, but this was more pronounced
in the former. This indicated that ascorbic acid was more efficient than beta-carotin in the formation of GSH in liver of female rats. It was suggested that ascorbic acid exert its protective effects by acting as a precurs for GSH, the main low molecular antioxidant and as a free radical scavenger [Lin-Chun et al 1998, Kang-Soon et al 1998, Skrzydlewska and Farbiszeweski, 1999]. These authors reported an increase in glutathione reductase in the liver and GSH, related enzymes in serum and liver. Also, they reported that, ascorbic acid may alleviate oxidative tissue damage by quenching radical as they are produced or aggravate damage by interacting with free meta thions released from damaged cells to accelerate hydroxyl radical production.

Present results also showed that both of beta-carotene and ascorbic acid can increase of GSH level in the liver when given alone without insecticide. The protective effects of beta-carotene and ascorbic acid against fenitrothion may be due to the antioxidant actions of GSH [Lawlor and O, Brien 1997, Newholm 1999, and Nabila et al 2001).

Fenitrothion, however, is a greater acetyl choline estrase inhibitor than matathon (Barber et al, 1999). The toxic effect of such insecticide included, Hemorrhage, congestion which can most probably explained by the toxic effect of the drug on the endothelium of sinusoids and intercellular substance between such cells.

The degenerative effect on the hepatic cells can also explained by the same way with the decrease in the nucleic acid content. Such results are in agreement with those of Nabila et al, 2001, who studied the effect of pyriproxyfen on the liver of female rats and concluded that there was disorganization of hepatic cords with debris from necrotic cells, massive cellular infiltration around the congested vascular and scattered hepatocellular degeneration with individual cell necrosis. Also, Salem et al 1995, on their study on the pathological action of chronic organo phosphorous toxicity concluded, that there was degenerative and necrotic changes in the wall of blood vessels as well as in the liver, lungs, kidneys, intestines and testicles.

Antioxidants such as beta-carotene, however improves some toxic changes specially vascular changes most probably through their action on the metabolic processes of the endothelial cells and inter cellular substance this was prooved by the work of Junqueira et al 1988, on lindane toxicity with morphologilical evidence and hepatic cell damage. On the other hand, there was no improvement of the nucleic acid content with compination with beta-carotene and insecticide.

However, the effect of ascorbic acid as a reducing agent can ameliorate the degenerative effect of the insecticide on the metabolic processes of hepatic cells. The work of Kang – Soon et al. 1998, prooved that pretretment with vit.c prevented the pathological changes in the lungs of rats treated with 50 mg/kg paraquat, also the work of Nabila et al (2001) on the effect of cysteine and green tea on the liver of albina rats treated with pyriproxyfen insecticide may have similar explana -tion. She attributed the protective effect of cysteine may be through its action as a precursor for GSH the main low molecular antioxidant and as a free radical scavenger.

On the other hand, there was no improvement of the nucleic acid content
with complexion of vit. E. and insecticide.

As regards glycogen content in hepatic cells the toxic effect of the insecticide fenitrothion will affect glycogen storage by such cells as was concluded by Justine et al. 1991, Sato and Izumi 1989, They stated that the disturbance that occurred in the carbohydrate metabolism may be secondary result of the hepatic damage and tissue, injury which may be responsible for impairment of liver and kidney function due to intoxication with insecticide.

Beta- carotene, however, does not improve the glycogen content when given with fenitrothion on the other hand the content decreased which may indicate a sort, synergistic action between the fenitrahion and beta-carotene or ascorbic acid as reducing agent improves the metabolic activity of the hepatic cells so when given with fenitrothion, it caused marked improvement and increase in glycogen content in hepatic cells.

Acknowledgement

The authers would like to express their hearty appreciation sincere gratitude to professor Dr.Sherif M.Hashim professor of Histology Department of Histology Faculty of Medicine Al-Azhar University for his valuable advise , efforts and unlimited help in preparing this work.

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Analytical and microscopical studies

دراسة تحليلية ومجهزية على التأثير الوقائي لحامض الاسكوريك (فيتامين جـ) والبيتاكاروتين ضد سمية مبيد الفيتيروثيون على كبد إناث الفئران البيضاء

أكرم فائق هاشم* - قديري وشاصي محمود**
قسم رقابة النبات
كلية الزراعة -فرع الفيوم- جامعة القاهرة* - كلية الزراعة- جامعة عين شمس**

تمت هذه الدراسة التحليلية والمجهرية على دم وكبد إناث الفئران البيضاء لتقسيم سمية مبيد الفيتيروثيون الفسفوري العضوي في إناث الفئران البيضاء ودراسة احتمال التأثير الوقائي لحامض الاسكوريك (فيتامين جـ) والبيتاكاروتين كمضادات أكيدة ضد سمية المبيد.

وقد استخدم في هذا البحث ستون من إناث الفئران البيضاء البالغة حيث قسمت عشوائيا إلى 6 مجموعات متساوية (10فئران في كل مجموعة).

المجموعة الأولى: استخدمت كمقارنة.
المجموعة الثانية: جرعت بمبيد الفيتيروثيون (15.65 مجم/كم من وزن الجسم وهي تساوي 1/16 من LD50).
المجموعة الثالثة: جرعت بالمبيد حامض الاسكوريك (15.65 مجم/كم من وزن الجسم و10 مجم/كم من وزن الجسم على التوالي).
المجموعة الرابعة: جرعت بالمبيد والبيتاكاروتين (15.65 مجم/كم من وزن الجسم و10 مجم/كم من وزن الجسم على التوالي).
المجموعة الخامسة: جرعت بحمض الاسكوريك فقط (10 مجم/كم من وزن الجسم).
المجموعة السادسة: جرعت بالبيتاكاروتين (10 مجم/كم من وزن الجسم).

تمت عملية التجربة يوميا لمدة 28 يوم.

وبعد نهاية فترة التجربة أظهرت النتائج كما يلي:

المعاملة الفيتيروثيون أدت إلى زيادة ذات دلالة إحصائية في الأنزيمات الناقلة لمجموعة الأمين (AST,ALT) والأنزيمات الفوسفاتيز القاعدية بينما ادي المبيد لانخفاض ذي دلالة إحصائيه في مستوي الجلوكوز ونشاط انزيم الاستيل كولين استيراز والجلوكازين المنخفض. بينما كان له تأثير غير معنوي على البيليروتين الكلي وانخفاض طيف لحلة البروتين الكلي في الدم وقد أظهرت المعالمة بحمض الاسكوريك والمبيد والبيتاكاروتين والمبيد في الدراسات التحليلية عدم حدوث أي تغيير في دلالة إحصائيه في نشاط كل من الأنزيمات المتحدة والمحتوي البروتين الكلي و bèlie وبيتيروبين وكذلك الجلوكوز وذلك مقارنة بالمقارنة.

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وقد بُسنت الدراسات الهستولوجية على عينات من كبد إناث الفئران باستخدام صبغة هيماتوكسيلين والأيوسين ومادة البيروآيد شيف الحامض وصبغة الميثيل بيرونين الاخضر على وجود اتساع في الأوعية الدموية بين الخلايا الكبدية وكذلك نزيف دموي بين الخلايا وظهور نحلال في بعض الخلايا الكبدية ونقص في محتوي الاحتياض النووي والجلوكوجين تحت تأثير المبيد.

وادت المعاملة بالمبيد مع حامض الإسكوريك والملبيد مع الببتاكاروتين إلى تحسين هذه النتائج على الأوعية الدموية تحت تأثير المعاملة بمضادات الأكسدة وكذلك التحلل الخلوي بالكبد تحت تأثير المعاملة بحمض الإسكوريك وتحسين محتوي الأحماض النووية في الخلايا الكبدية وقد نوقشت النتائج في ضوء الأبحاث العلمية المرتبطة في هذا المجال واستنتج من البحث ضرورة أخذ الحيطة والحذر عند استخدام مبيد الفينيتروثيون حصوصا عند استعمال جرعات عالية منه ويمكن باستخدام مضادات الأكسدة التقليل من مظاهر التسمم لهذا المبيد.