A Six-Year Clinical, Microbiological and Radiological Study Outcome Following Treatment of Peri-implantitis

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Abstract

The present study evaluated the long-term outcome of a combined surgical and composite graft (bovine porous bone mineral BPBM and autogenous bone) in the treatment of peri-implantitis. Six individuals with titanium implants demonstrating a marginal bone loss. Baseline, 1 and 6 years measurements, including introral digital radiographs, gingival index (GI), probing depth (PD) and probing bone level (BL) were performed. In each implant subgingival bacterial samples were obtained and subjected to microbiological analysis by culture. Surgical exposure of the lesions and cleaning of the implants using hydrogen peroxide and 2% aqueous solution of chlorohexidine gluconate were performed. The autogenous bone was harvested from the chin combined with BPBM and packed into the bony defects. The treatment was evaluated clinically, microbiologically and radiographically at 1 and 6 years. Results: revealed a significant reduction in plaque and gingival bleeding. The mean probing depth was 6.94+ 1.16 at the baseline and it was 1.91 + 1.21 at the end of the study period. All treated implants showed bone gain. The mean gain in bone level was 5.12 + 0.5 and radiological evaluation of bone density was measured at crest, mid-implant and lateral apical area of interest AOI 32.0 + 0.87, 72.91 +0.83, 63.0 + 0.94 at baseline and it reached 109.5 + 0.53, 141.63 + 1.19 and 144.0 + 0.47 respectively. The presence of putative periodontal pathogens significantly declined at the end of the study period.

Conclusions:

The results of the study suggest that the use of composite bone grafts as the treatment strategy for peri-implantitis lesions with maintenance of good oral hygiene appears to be an efficacious treatment approach for restoring hard tissue support of dental implants during the 6 year follow-up period.

Introduction

Implant dentistry is a relatively new and fast growing area of oral health services (18). Successful treatment results and patient satisfaction with dental implant treatment modalities must include prevention and treatment of peri-implant infections (25). Peri-implantitis and periodontitis are often described as inflammatory diseases with oral pathogens emerging either at implants, within the intraimplant components or teeth. Peri-implantitis has been associated with gram-negative bacteria similar to periodontitis (5). Peri-implantitis has been associated with gram-negative bacteria similar to periodontitis (20). The goal of implant dentistry today should be the control of periodontal pathogens in the oral cavity. Once a peri-implant inflammatory process starts, implant sites get compromised by severe loss of peri-implant supported bone.

Meffert et al, (21) classified implants with complication into three types: Ailing implant in which bone loss accompanies
pocket formation, but the implant is static at maintenance checks; failing implant with bone loss irrespective of therapy, and purulent exudate; and failed implant, showing mobility, dull sound on percussion and peri-implant radiolucency. Ailing and failing implants may be treated but failed implants must be removed because bone loss will continue. The failing implant is non-mobile with bone loss due to the following causes of peri-implantitis overheating of bone during placement, lack of sufficient bone.

To enhance the level of patient expectations and compliance, it is of special interest to determine whether it is possible to maintain the affected implant by bone grafting techniques or guided bone regeneration GBR procedures and rebuild the previously peri-implant tissues (18). Longitudinal studies have reported survival and success rates around 90-95% for osseointegrated titanium implants (10). Peri-implantitis has been reported to be prevalent in 4-15% of individuals with implants (30). Misch (23) suggested the use of bone grafts for management of failing implants. Various graft materials are being utilized today in an attempt to repair osseous defects around implants. Limited data are available regarding treatment of peri-implantitis in humans. The use of local or systemic antibiotics, mechanical debridement, guided tissue regeneration and autogenous bone grafts have been described in the literature (24, 27, 30). In humans, only case reports are available (14) and longitudinal studies evaluating different treatment approaches in peri-implant diseases are lacking. Moreover, the studies performed so far are all short-term studies (less one year) and little is known about the implant survival rate on a long-term basis (6 years). The aim of this study was to evaluate the clinical, microbiological and radiological outcomes of the use of a composite graft as a treatment modality aimed at reconstructing intraosseous peri-implant intrabony defects over a 6 year maintenance period in patients with loss of peri-implant supporting bone.

### Material and Methods

#### Subjects

The data of this study were taken from six patients attending the outpatient clinic of Dental department Al Zahraa University Hospital, Faculty of Dentistry, Al Azhar University and the outpatient clinic of Oral Medicine & Periodontology and Diagnosis Department, Faculty of Oral and Dental Medicine, Cairo University. The participants were four males and two females. Their ages ranged from 35-55 years, they suffered from peri-implant hard tissue break down around single tooth replaced implants*. The patients were systemically free from any systemic disease and evaluated by the aid of dental modification of Cornell Medical Index (17).

#### Peri-implant parameters

Subjects fulfilling the inclusion criteria were invited to participate in the study and if accepted signed an informed consent form (baseline). All patients were monitored one year and 6 years after treatment. Gingival index, probing depths (the distance between the peri-implant gingival margin and the bottom of the peri-implant pocket) and probing bone levels (under local anaesthesia; the distance from the implant shoulder to the deepest depth at which the probe meets strong resistance from contact with bone) were measured at 4 sites around the implant (midbuccal, distobuccal, midlingual, distolingual) Using a periodontal probe 0.4 mm diameter tip. The probing depths and probing bone level measurements (reproducibility + 1mm in greater than 95%) were performed by the same calibrated examiner.

Microbiological examination was performed at baseline (immediately when the patients attended to the study and before any hygienic treatment was carried out), then at one years and 6 years to detect any anaerobic microorganisms that may be located subgingivally. Each implant was isolated with cotton rolls after removing the supragingival plaque with a plastic curette. After air drying, a sterile paper point was inserted subgingivally and left for 30 seconds then dipped in 1 ml of thioglycolate fluid medium using a calibrated loop.

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* Paragon Implant: Core Vent 15821 Ventura Boulevard, USA
A sample of one micron was inoculated into a non-selective media of schaedler anaerobic agar plate enriched with sheep blood.

Selective media included a selective medium for non-sporing anaerobes NS and a selective medium for gram negative anaerobes (GN). All plates were incubated at 37 C up to 15 days to allow the growth of slowly growing anaerobes. The plates were inspected every 48 hours to detect a newly growing colony type. Each colony that appeared was subjected to the following: Gram stained smear, subculturing on anaerobic plates that were incubated anaerobically to get a pure growth of strict anaerobic organisms.

The identification of anaerobes was made using the analytical profile index. Microbiological examination comprised the incidence: which is the number of sites colonized by each member of the bacterial group.

Radiographic assessment

Radiographic follow-up was performed at baseline, 1 and 6 years (fig. 1,2&3) in order to evaluate the amount of bone formation. It depended on evidences of calcification and bridging on the peri-implant bone defect by newly formed bone. Radiovisioigraphy RVG was used in this study using the paralleling technique, which required a film holder that kept the sensor in a proper position in the mouth. The density value within marked points were measured at the crestal area of interest AOI, mid and lateral apical area of interest AOI.

Fig 1: baseline radiograph showing osseous defect along the implant
Surgical Procedures

Following the baseline examination thorough rinsing of the peri-implant pocket was performed with hydrogen peroxide and 2% aqueous solution of chlorhexidine gluconate; systemic metronidazole (250mg once a day) was administered and maintained for 2 weeks. After 2 weeks of this hygienic phase the patients were prepared for the surgical procedure. Following local anaesthesia, full thickness flaps were elevated buccally and lingually. Granulation tissue was carefully removed by plastic curettes which was performed to remove all soft tissue from the defect. The implant surfaces were cleaned using 10% hydrogen peroxide followed by careful rinsing with saline. The bony defect in most instances was vertical and infrabony in nature. A composite bone graft formed of autogenous bone harvested from the chin and mixed with cancellous bovine porous bone mineral* “particle size 0.25 to 1.0 mm” was carefully packed using new plastic amalgam condensers to the level of the surrounding bone into the bony defect. The flap was then repositioned to completely cover and to carefully and tightly adapt around the neck of the implants using silk sutures. An antibiotic regimen was prescribed (3 times daily 250 mg of metronidazole and 375mg of amoxicillin) for ten days. The patient was instructed to rinse twice daily with 0.02% chlorhexidine. One week later, the sutures were removed. Regular check-up was performed monthly on the first year for professional cleaning and motivation of oral hygiene measures and yearly thereafter. The patients were recalled for reassessment of the clinical, radiographic and

*Bio-oss, Geistlich, Wolhusen, Switzerland
microbiological examination. The clinical parameters included probing depths, and probing bone level whereas grey level of bone around the implants were evaluated radiographically. All the clinical data was statistically analyzed using the student t test while the microbiological data was analyzed by the chi square test.

Results

Clinical Results

Statistical analysis of change in the gingival bleeding showed a highly significant difference between the baseline and 1 year and with the baseline and 6 years. However, there was no significant difference between 1 year and 6 years table (1) fig. (4). The results of the present study revealed that all the cases showed favorable clinical and radiographic changes. The peri-implant bony defects were apparent within the first 2 years of the functional phase. Sites with gingival bleeding decreased dramatically from 100% at the baseline to 10% after 1 year and 3% at the six-year registration. Statistical analysis of change in the gingival bleeding showed a highly significant difference between the baseline, 1 year with the baseline and 6 years follow-up period while there was no significant difference between 1 and 6 years follow-up period. The mean probing depth over the experimental period was reduced from 6.94 ±1.16 to 3.00 ±1.36 after one year, and it reached 1.91 +1.21 at the end of the study table (2) fig. (5). Moreover Statistical analysis of the bone level revealed highly significant differences between baseline and 6 years follow-up period p<0.01 and between baseline and 1 year while there was no significant difference between 1 and 6 years. The baseline decreased from 7.02 ±1.71 to 4.32 ±1.67 and 1.90 ±1.13 at the end of the study period. Table (2) and fig.(6)

Radiological Results

Table (3) presents the comparison of bone density at baseline, 1 year, 6 years. The mean bone density at the crestal area was 32.0 ± 0.87 at baseline, it was also 109.5 ± 0.53 and 108.38 ± 1.41 at 1 year. The statistical difference was highly significant between baseline and 1 year, and between baseline and 6 years follow-up period while there was insignificant differences between 1 and 6 years follow-up period. The mean value of bone density at the mid-implant AOI was 72.91 ± 0.83 at baseline reached 141.63 ± 1.19 and 140.5 ± 1.07 at the end of the study. The bone density was at lateral apical AOI at baseline was 63.0 ± 0.94 and it was 144.0 ±0.47 at 1 year. At 6 years follow-up period it was 143.0 ± 0.94 at the end of the follow-up period. The statistical analysis revealed significant difference between the baseline and the end of the study. Fig. (7) shows the comparison between the bone density at crest, mid, lateral apical AOI at baseline, 1 and 6 years.

Microbiological results

The composition of subgingival microbiota obtained from peri-implantitis sites and cultured for their identification is presented in table (4) and figure (8) Peptostreptococci was detected in 18.9% of sites at baseline and 12.1% after 6 years. Sixty five percent of Bacteroid fragilis was found at baseline and 21.7% was found at the end of the follow-up period. Porphyromonas gingivalis was detected in 75% at the baseline while it decreased to 13.4% at the end of the study. Similarly 80% of the sites showed the presence of Provetlla intermedia at baseline while it reached 22% at the end of the study period. 33.3% of the sites harboured enteric rods (Escherichia coli or Escherichia cloacae at the beginning of the study while at the end of the 6 year follow-up registration these species were recovered in 5.2% of sites. Fusobacterium nucleatum was detected at baseline in 50% of sites and it was not detected at the end of the study. Lactobacillus acidophilus was recovered in 33.3% of sites at baseline and dropped to10.3% after 6 years. Conclusively, all patients harboured at least one of the marker organisms in a high percentage at baseline while the percentage of
microorganisms declined at the end of the follow-up period. There was a statistical significant difference between the baseline and the end of the study period regarding all the microbiota harbouring the peri-implant sites.

Table (1): Gingival Index

<table>
<thead>
<tr>
<th>Site</th>
<th>Baseline</th>
<th>1 year</th>
<th>6 years</th>
<th>P value</th>
<th>Significance</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>100%</td>
<td>10%</td>
<td>3%</td>
<td>0.000</td>
<td>HS</td>
</tr>
</tbody>
</table>

P value: level of significance, Baseline: immediate postoperative, HS: highly significant

Table (2): Mean probing depth and bone level at the implant sites at different time intervals

<table>
<thead>
<tr>
<th>At Baseline</th>
<th>Probing depth</th>
<th>Bone level</th>
</tr>
</thead>
<tbody>
<tr>
<td>6.94 + 1.16</td>
<td>7.02 + 1.71</td>
<td></td>
</tr>
<tr>
<td>3 + 1.36</td>
<td>4.32 + 1.67</td>
<td></td>
</tr>
<tr>
<td>1.91 + 1.21</td>
<td>1.90 + 1.13</td>
<td></td>
</tr>
</tbody>
</table>

P value baseline & 1 year

<table>
<thead>
<tr>
<th>P value baseline &amp; 6 years</th>
<th>Significance</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.000</td>
<td>HS</td>
</tr>
</tbody>
</table>

P value baseline & 6 years

<table>
<thead>
<tr>
<th>P value 1 year &amp; 6 years</th>
<th>Significance</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.075</td>
<td>NS</td>
</tr>
</tbody>
</table>

Gain in Bone Level

<table>
<thead>
<tr>
<th>Reduction in the Mean Pocket Depth</th>
<th>5.03+0.05</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gain in Bone Level</td>
<td>5.12+0.5</td>
</tr>
</tbody>
</table>

P value: level of significance, Baseline: immediate postoperative, NS: non-significant, HS: highly significant
Table (3): Bone density around the implants at different sites and different time intervals

<table>
<thead>
<tr>
<th>Mean Bone Density</th>
<th>At Crest</th>
<th>Mid-implant</th>
<th>Lateral-apical</th>
</tr>
</thead>
<tbody>
<tr>
<td>At Baseline</td>
<td>32.0 + 0.87</td>
<td>72.91 + 0.83</td>
<td>63.0 + 0.94</td>
</tr>
<tr>
<td>1 year</td>
<td>109.5 + 0.53</td>
<td>141.63 + 1.19</td>
<td>144.0 + 0.47</td>
</tr>
<tr>
<td>6 year</td>
<td>108.38 + 1.41</td>
<td>140.5 + 1.07</td>
<td>143.0 + 0.94</td>
</tr>
<tr>
<td>P value (baseline &amp; 1 year)</td>
<td>0.0000</td>
<td>0.000</td>
<td>0.000</td>
</tr>
<tr>
<td>Significance</td>
<td>HS</td>
<td>HS</td>
<td>HS</td>
</tr>
<tr>
<td>P value (baseline &amp; 6 year)</td>
<td>0.000</td>
<td>0.000</td>
<td>0.000</td>
</tr>
<tr>
<td>Significance</td>
<td>HS</td>
<td>HS</td>
<td>HS</td>
</tr>
<tr>
<td>P value (1 year &amp; 6 year)</td>
<td>0.92</td>
<td>0.959</td>
<td>0.979</td>
</tr>
<tr>
<td>Significance</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
</tr>
</tbody>
</table>

P value: level of significance, Baseline: immediate postoperative, NS; non-significant, HS: highly significant.
Table (4): The incidence of different microbes identified over the study period

<table>
<thead>
<tr>
<th>Microbiota</th>
<th>Baseline</th>
<th>1 year</th>
<th>6 years</th>
<th>P value</th>
<th>Significance</th>
</tr>
</thead>
<tbody>
<tr>
<td>Peptostreptococci</td>
<td>18.9</td>
<td>15.3</td>
<td>12.1</td>
<td>0.01</td>
<td>HS</td>
</tr>
<tr>
<td>Bacteroid Fragilis</td>
<td>65</td>
<td>33.3</td>
<td>21.7</td>
<td>0.00</td>
<td>HS</td>
</tr>
<tr>
<td>Enterorods</td>
<td>33.3</td>
<td>22.2</td>
<td>5.2</td>
<td>0.23</td>
<td>NS</td>
</tr>
<tr>
<td>Porphyromonas Gingivalis</td>
<td>75</td>
<td>20.7</td>
<td>13.4</td>
<td>0.00</td>
<td>HS</td>
</tr>
<tr>
<td>Prevotella Intermedia</td>
<td>80</td>
<td>50</td>
<td>22</td>
<td>0.00</td>
<td>HS</td>
</tr>
<tr>
<td>Fusobacterium Nucleatum</td>
<td>50</td>
<td>16.7</td>
<td>0</td>
<td>0.00</td>
<td>HS</td>
</tr>
<tr>
<td>Lactobacillus Acidophilis</td>
<td>33.3</td>
<td>10.3</td>
<td>7.1</td>
<td>0.00</td>
<td>HS</td>
</tr>
</tbody>
</table>

P value: level of significance, Baseline: immediate postoperative, NS: non-significant, HS: highly significant

Discussion

The present study evaluated the long-term outcome of treatment of peri-implant destruction, which showed predictable results of maintaining implants treated for peri-implantitis using a treatment model consisting of surgical cleaning and guided bone regeneration for 6 years. Only the peri-implantitis cases with more than 5mm probing depth that were in function for a minimum of 2 years were included.

Following elevation of a full thickness flap a transient burst of regional remodeling occurs. Starting with accelerated resorption activity followed by a slow process of bone regeneration (1, 35). In order to avoid this phenomenon, the flap in the present study was performed in a conservative manner to expose the area of the defect. Moreover, bone loss and remodeling after flap elevation without osseous surgery in periodontitis treatment, ranged from no resorption to 0.8mm.

At the time this study started, little was known about how to deal with infections around implants. Due to ethical reasons and the severity of the included cases, a control group was not included. Several animal studies and case reports investigated the possibility of regenerating new supporting bone around implants using guided bone regeneration (11) these studies produced contradictory results regarding GBR, ranging from no “reossesointegration” (13) to minimal reosseointegration (26) to clinically significant reossseointegration (15).

Infections around titanium implants is accompanied by plaque accumulation and
are difficult to treat (34). From a clinical viewpoint, it may be insufficient to restrict treatment to scaling and polishing only. Therefore, the treatment approach in this study involved flap surgery in order to expose the bone and the fixture for more accessibility to mechanically clean the infected area by using 0.2% chlorhexidine, 10% hydrogen peroxide. The aim of this procedure was to disrupt the integrity of the biofilm, remove and to prevent re-colonization of the vast majority of the microorganisms. This procedure was combined with the use of GBR. The GBR aimed at reconstruction of the lost bony support that was destroyed by the disease process. Several treatment procedures including the use of various bone grafts (27), bone substitutes and GTR (16), and growth factors were suggested for active regeneration of the tooth supporting apparatus. The ideal graft material is the one that is osteoinductive, absorbable, easy to handle, and available in large quantities (6). Autogenous bone meets the first three requisites, but obtaining the needed amount from the intraoral site is often problematic besides donor site morbidity. Allografts and alloplastic materials have been extensively used in regenerative surgery but their osteoinductive capabilities have been shown to be quite variable (31). More recently, a bovine derived xenograft has been introduced into periodontics. This material is available in an unlimited supply and proven safety (33). It is prepared by protein extraction of bovine bone but maintains the natural structure of bone (12).

When evaluating parameters such as inner surface area, porosity, crystalline size, and calcium-to-phosphorus ratio, bovine porous bone mineral BPBM most closely resembles human cancellous bone as compared to other allografts or synthetic hydroxyapatite materials (36). Several studies documented the ability of BPBM to enhance bone formation in sinus elevation procedures (37) around implants (4), and in critical-sized osseous defects (32). True histologic regeneration has been demonstrated in humans when using BPBM alone (22), but more complete and more predictable histologic regeneration was achieved when BPBM was used in combination with bioabsorbable membranes (7) and even more with autogenous bone alone (8). From the abovementioned reasons the composite graft used in the present study was BPBM and autogenous bone. Beheneke et al (3), reported a 3.5mm reduction in probing depth and a 2.2 mm gain in clinical attachment level, the study utilized autogenous bone graft as an augmentation material owing to the possibility of cellular viability and consequent rapid vascularization. Ragy et al (29) reported 2.28 mm reduction in probing depth & 2.44 mm gain in bone level in the treatment of peri-implantitis using alloplast bone graft. Our results showed a reduction in probing depth of 5.03+0.05mm and a gain in bone level of 5.12+0.5mm with the use of composite bone graft which proved to be statistically significant and in accordance with previous studies (3, 29). However, Grunder et al (13) showed a lower potential for bone regeneration, which could be explained partly by the anatomic variations of the bony defects, which were horizontal bony defects as opposed to the intrabony defects in the present study. On the other hand the superior results obtained in this study may be attributed to the combination between autogenous bone & BPBM graft materials. The results of the present study revealed that it was not difficult to restore a healthy situation in conjunction with implants over a 6 year period. The gingival inflammation was resolved and new bone formation occurred.

The changes in the bone density around the implants were studied radiographically which revealed that all the treated implants were well integrated with no further bone loss or periapical radiolucencies. Moreover there was a significant increase in the density of bone at the three areas of interist around the treated implants. This increase in bone density was significantly higher at one and six years follow up period. The result of the change in bone density was in accordance with Ragy et al, 2001 who reported favorable osseous changes in response to bone graft in the treatment of periimplantitis.
Regarding the microbiological findings in the present study there was a significant reduction and total elimination of other pathogenic bacteria which were detected from the beginning of the study. The results were in accordance with the results of Leonhardt et al (19) who reported a total elimination of actinomycetem comitance and the enteric rods in long-term peri-implantitis treatment. The obvious decrease in the percentage of microorganisms which harboured the sites at the end of the study highlighted the need for long-term plaque control to maintain the health of the implant soft and hard tissue interface.

Conclusions
The results of the present study suggest that the combination between autogenous and BPBM bone grafts appears to be an efficacious treatment approach for restoring bone around dental implants following progressive bone loss caused by peri-implantitis. The present method resulted in successful clinical and radiographic bone fill and resolved the peri-implantitis lesion over an observation period of 6 years.

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تقييم نتائج ستة أعوام من الدراسة الإكلينيكية و الميكروبيولوجية والفحص
بالأشعة العلاجية الالتهاب حول الغرسات

عبير سعد جاويش ، سوزان عبد الحكيم حسان
كلية طب الأسنان بنات – جامعة الأزهر

تقوم هذه الدراسة بتحليل نتائج على المدى الطويل لعلاج غرسات الأسنان باستخدام الجراحة مع وضع مخلوط من العظام الطبيعية المأخوذ من عظام الذقان مع جزئيات مفرغة من العظام البقرية.

ضم هذا البحث ست أفراد يعانون من تأكل في العظام حول غرسات بالتيمون. تم اخذ قياسات لمدى عمق الجيوب اللثوية حول الغرسات وكذلك حالة التزيف اللثوي. وقد تم أخذ نسج بكثيرية من حقول الغرسات وعمل مزرعة لمعرفة نوع البكتيريا المحيطة بالغرسات وكذلك تم قياس كثافة لعظام حول الغرسات. أخذت كل هذه المعلومات عند حضور المرضى أول مرة وبعد عام ثم بعد ستة أعوام لاحق.

وقد أثبتت النتائج أن هناك تغير إحصائي إيجابي مؤثر في حالة تجمع البلاك وحالة التزيف اللثوي وكذلك في عمق الجيوب اللثوية حول الغرسات المعالجة. أما التقييم بالأشعة لكثافة العظام حول الغرسات فقد وجد زيادة إحصائية مؤثرة إيجابية لزيادة كثافة العظام حول الغرسات المفعلة وقد لوحظ نقص ملحوظ في عدد ونوع البكتيريا المرضية عند نهاية البحث.

ومن ذلك نستنتج أن استخدام مخلوط العظام كخطط علاجية للالتهابات والتآكل العظمي حول غرسات الأسنان مع الحفاظ بالاهتمام بنظافة الفم والأسنان وصحتها لها تأثير إيجابي للحفاظ على الأنسجة العظمية التي قد عم غرسات الأسنان خلال فترة الدراسة طويلة الأجل.