

Protective Effect of Vitamin C and Glutathione Against the Histopathological Changes Induced by Imidacloprid in the Liver and Testis of Japanese Quail.

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Abstract

Pesticides are double edged weapons. They are considered to be the heavy cost of civilization. They are widely used in our daily life. Recently their effects on male fertility have attracted attention. One of these pesticides is imidacloprid. This work was designed to investigate the histological changes in liver and testis of Japanese quail treated with imidacloprid for different periods as well as the reversibility of such changes after arrest of the treatment for recovery period. The effect of vitamin C and glutathione as a protective agent against the action of imidacloprid on liver and testis was also determined.

Introduction

Pesticides are widely used for control of pests in many fields to protect public health, subsistence of crops, food and other materials essential to mankind and animals.

Imidacloprid is a new insecticide and it belongs to a new active group nitroguanidine. Many representatives of this class of compounds, imidacloprid in particular, have an excellent insecticide action (Brocksma *et al.* 1993). The action mechanism of imidacloprid differs not only from that of the organophosphorous compounds and carbamates, but also from that of the pyrethroids. (Soloway *et al.*, 1978).

It is of interest to study the effect of imidacloprid on one of the most important economic birds, the Japanese quail (*Coturnix Coturnix japonica*). Dieher *et al.* (1991) found that after the discovery of the nitromethylene and chloropyridyl insecticidal compounds, which are considered as powerful insecticides and perform an important mechanism of action more over than phosphates, carbamates and pyrethroid insecticides groups. Another potent insecticides, in addition to the nitromethylene compounds, are the guanidine derivatives.

Among the various problems associated with pesticide use is the possibility of its biological accumulation which produces real problems, considering that many animal tissues and milk are ingested by human beings (Kutches *et al.*, 1970), which may cause clinical and subclinical effects leading to loss in animal performance or in residual contamination of animal deviation which may later be consumed by humans (Ceron *et al.*, 1995). Thus, the disappearance of pesticide residues at a given location does not mean the end of the problem, but it might be translocated, bioconcentrated or converted into more dangerous compounds (Matsumura, 1985).

Zafeiridon and Theophilids (2004), described the mode of action of imidacloprid as agonist on the nicotinic acetylcholine receptors of motor neuron, i.e. prevent the binding of acetylcholine to certain acetylcholine receptors (AChRs) by binding to these receptors themselves, in other words, imidacloprid acts as nicotinic acetylcholine receptors stimulator (nAChRs). The selectivity of this molecule and analogue may be due to the insensitivity of mammalian nicotinic

acetylcholine receptors (nAChRs) and the high sensitivity of analogous receptors in insects.

Imidacloprid recorded as one of the best protecting insecticide against ctenocephalides flea on cats and dogs, it provided a high level of residual activity in a very low time, either in laboratory condition or in home environment model (Dennis *et al.*, 2000 & Hutchinson *et al.*, 2001) and Liburd *et al.*, (2004).

Philippe *et al.* (1999), reported that when pigeon feed on potential ingestion of imidacloprid coated seeds, the primary metabolite shape of imidacloprid was 6-chloronicotinic acid in the tissues and organs of affected animals. Birds found dead after exposure to imidacloprid, toxic residues detected in the liver of affected pigeons.

Pfluger and Schmuck (1991), studied the possibility of contact with the active substance, and hence the risk of side effects of imidacloprid insecticide is greatly restricted for many organisms in the environment. Imidacloprid has to be classified as toxic agent to birds with an LD50 ranges between 25-50 mg/kg.

The mammalian toxicity of imidacloprid is low. The lowest LD50 value of 131-168 mg/kg was determined in mice. The subacute toxicity for a mammal is also outside the hazardous range. The reproduction toxicity in mammals is in the same favorable range as in birds. The risk of imidacloprid was increased during its application in a seed dressing or in granules more than the residues of spray application (Shull and Coheeke 1983).

Abbink (1991), discovered the properties of the new insecticide imidacloprid and its analogues and has shown that it had an excellent insecticidal action which can similarly be achieved by interfering with the postsynaptic receptors. It has been released into the synapse (the neurotransmitter cannot exert an effect unless it binds to the receptors in the postsynaptic membrane.).

Imidacloprid and its derivatives prevent the binding of acetylcholine to certain acetylcholine receptors by binding

to those receptors themselves (Abbink, 1991).

Insecticides transformed into chemically reactive metabolites in the liver is either irreversibly binds to hepatic glutathione to be detoxicated or bind with hepatic proteins and produced hepatic toxicity. Then the hepatic glutathione content can be greatly depleted after exposure to insecticides, and this leads to damage of the hepatic tissues. Then the addition of external glutathione as antidotes may make a good protection against insecticide toxicity. (Abd-Elghaney 2002).

Glutathione acts as an antioxidant and protective agent against insecticide. Glutathione also plays an important role in removal of hydroperoxides and hence protection from ionizing radiation and its maintenance of the sulfhydryl status of proteins and transport of Y- glutamyl amino acids (El-Naggar and Salah 1997).

Glutathione (Y-glutamyl-cysteinylglycine) is a typical amino acid in which the N-terminal glutamate is linked to cysteine via a non-peptidyl bond, is present in all forms of life. In humans and animals, glutathione is required for the action of several enzymes (Lehninger, *et al.*, 1993).

The amino acid cysteine through its active reducing sulphhydryl radicals (-SH) is the effective component of glutathione. This active group has proved its protective function in many toxic states (Yamada *et al.*, 1964; Horiuchi *et al.*, 1979 and Cicchella, 1972). These investigators demonstrated the safety of this compound as a natural body synthesized compound. Even at neoplasms level, a high protective dose of Glutathione administered with the antimetabolic agent, cisplatin has rendered the tumor less bulky and easily removed (Fontanelli *et al.*, 1992).

Vitamin C is not present in all birds' tissues so that it is considered as external antioxidants. It is an available substance, which passes from the plasma into the body tissues, where it plays an important role in liver metabolism, haemopoiesis, collagen formation, endocrine control, ovulation and other metabolic processes. Vitamin C plays an important role in insecticide toxicity

protection especially in the hepatic toxicity as antioxidant agent and prevents the effect of free radicals for the vital cells (Abd-Elghaney 2002).

Materials and Methods

1- Experimental animals:

A total of 90 male Japanese quails (*Coturnix coturnix japonica*) were kept under normal laboratory condition.

Birds were divided randomly into equal 6 groups of 20 birds in each, as the following:

Group 1: Control group.

Group 2: Injected with vitamin C (0.08 mg/kg body weight).

Group 3: Treated with glutathione (0.55 mg/kg body weight).

Group 4: Treated with 1/50 LD50 imidacloprid.

Group 5: Treated with imidacloprid and vitamin C (1/50 LD50 and 0.08 mg/kg body weight).

Group 6: Treated with imidacloprid and glutathione (1/50 LD50 and 0.55 mg/kg body weight).

Birds were orally treated daily with 1/50 LD50 imidacloprid insecticide via stomach tube, the antidotes were used to defense quails against insecticide effect. Vitamin C and glutathione were used to be antidotes on quails treated with imidacloprid, for 3 and 6 weeks followed by another 3 weeks as recovery periods.

Glutathione was available to be dissolved in water and administrated orally via stomach tube, in a dose equal to 0.55 mg/kg of quail.

Birds were scarified at 3 and 6 weeks followed by another 3 weeks as a recovery period. The scarified animals were quickly dissected and small pieces from liver, and testes were rapidly removed and then fixed in 10% neutral buffered formal for 24h. Following fixation the specimens were carefully washed in running tap water, dehydrated in ascending series of alcohol, cleared in xylene and then embedded in paraffin wax. Sections of 5 μ thickness were cut and stained with haematoxylin and eosin according to method adopted by Drury and Wallington (1980).

2- Insecticide used:

Common name: Imidacloprid

Trade name : Gaucho, admir, confidor 20% EC

Chemical name: 6-chloro-3-pyridnylmethyl; 4,5dihydro-N-nitro-0-Himidazol-2-omine

Chemical class: nitro guandine group

Company : Bayer Company Egypt

LD50 : 31 mg/kg body weight

Dose : 1/50 LD50 daily administration via stomach tube

Time : 3, 6 weeks of treatment followed by recovery period (3 week)

Results:

Normal structure of hepatocytes, endothelial lining of the central vein and sinusoidal spaces can be observed in Fig (1), Some degenerative changes could be observed in liver of Japanese quail treated with imidacloprid for 3 & 6 weeks including highly dilated portal spaces, large degenerated area, faintly stained cells and nuclei Fig (2).

Irregular arrangement of hepatocytes with abnormal architecture, large area of necrosis, dilated sinusoidal spaces, large and small areas of degeneration with faintly stained cytoplasmic nuclei were seen after 3 weeks recovery periods. Figs. (3, 4).

Normal well developed hepatocytes, observed in liver of quail injected with vitamin C for 3 & 6 weeks and a recovery period Fig (5) when quail treated with insecticide and vitamin C for 3&6 weeks, the liver tissues showed, highly dilated HPV & hepatic artery and degenerated bile duct, some cells appeared vacuolated (v) with dilated sinusoidal spaces(s). Figs. (6,7),. Signs of recovery could be observed after 3-week recovery period with good architecture of hepatocytes. Fig. (8). Liver of Japanese quail treated with glutathione for 3 & 6 weeks and 3 weeks recovery showed normal arrangement of hepatocytes as well as, normal portal space. Fig. (9).

Faintly stained hepatocytes, ballooning, (b) enlarged hepatocytes, wide sinusoidal spaces, pyknotic nuclei (P) surrounded by vacuolated cytoplasm, and some affected cells contained karyolytic

nuclei (K) could be observed in quail treated with insecticide and glutathione for 3 and 6 weeks Figs (10, 11).

After 3 weeks recovery from treatment with imidacloprid and glutathione marked recovery with normal architecture of hepatocytes, portal space containing normal HPV, artery and bile duct were observed while some hepatocyte appeared vacuolated (v) with signs of ballooning (b) Fig. (12)

The normal histological structure of the testis is shown in Fig. (13). Microscopic examination of testis taken from Japanese quail 3 and 6 weeks following treatment with imidacloprid revealed different degenerative changes included bizarre cells, disappearance of spermatogenic cells, thickened tunica albuginea, the tubules are devoid of sperms and some pyknotic nuclei could be noticed. No recovery signs could be observed after 3 weeks recovery periods where debris of sperms could be noticed, vacuolation of spermatocytes and rupture of basement membrane could be detected. Figs. (14-15-16).

The treatment of quails with vitamin C or glutathione alone showed normal appearance.

In testes of Japanese quail treated with imidacloprid and vitamin C for 3&6 weeks, most of spermatogenic layers appeared affected, debris of sperms, ruptured basement membrane could be observed. Figs.(17,18) as well as few Leydig cells could be seen, some spermatogenic layers could be observed, some tubules contained sperms.

Noticeable recovery could be detected in group treated with imidacloprid and vitamin C after 3 weeks recovery period. Fig. (19)

Group of quail treated with imidacloprid and injected with glutathione showed mild signs of improvement after 3 and 6 weeks. Appearance of some sperms and mass of undifferentiated cell could be observed while sign of degeneration and vacuolation were still noticed Figs (20, 21). While after 3 weeks recovery periods, remarkable recovery was observed where nearly most of the seminiferous tubules retained their normal architecture while vacuolation was still observed and some tubules have abnormal arrangement of spermatogenic layers and few Leydig cells were appeared Fig (22).

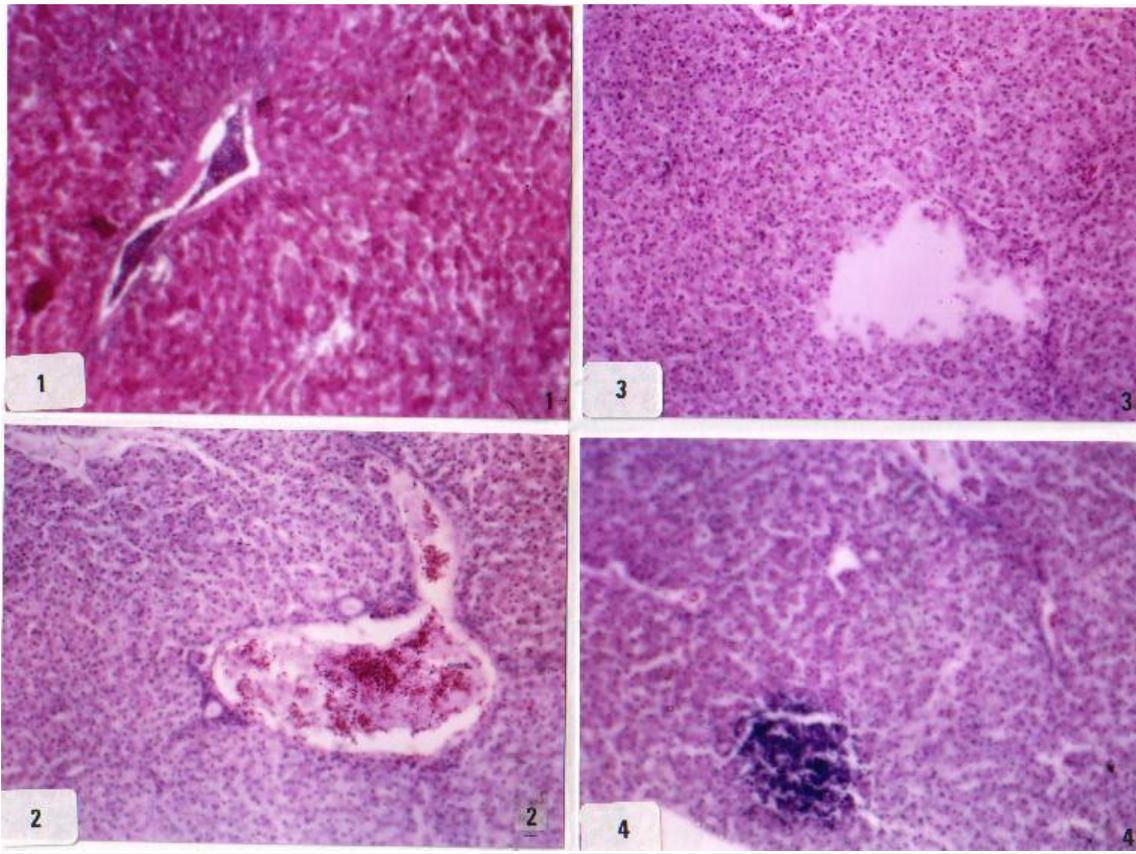


Fig. 1: section in control liver of quail showing normal structure of hepatocytes, endothelial lining of the central vein and sinusoidal spaces.

(Hx & X 250)

Fig. 2: section in liver of quail treated with imidacloprid for 3 & 6 weeks showing highly dilated portal space, large degenerated area, faintly stained cells and nuclei.

(Hx & X 250)

Figs. 3,4: section in liver of quail treated with imidacloprid after 3 weeks recovery, showing large area of necrosis, dilated sinusoidal spaces, large area of degeneration and faintly stained cytoplasmic nuclei.

(Hx & X 250)

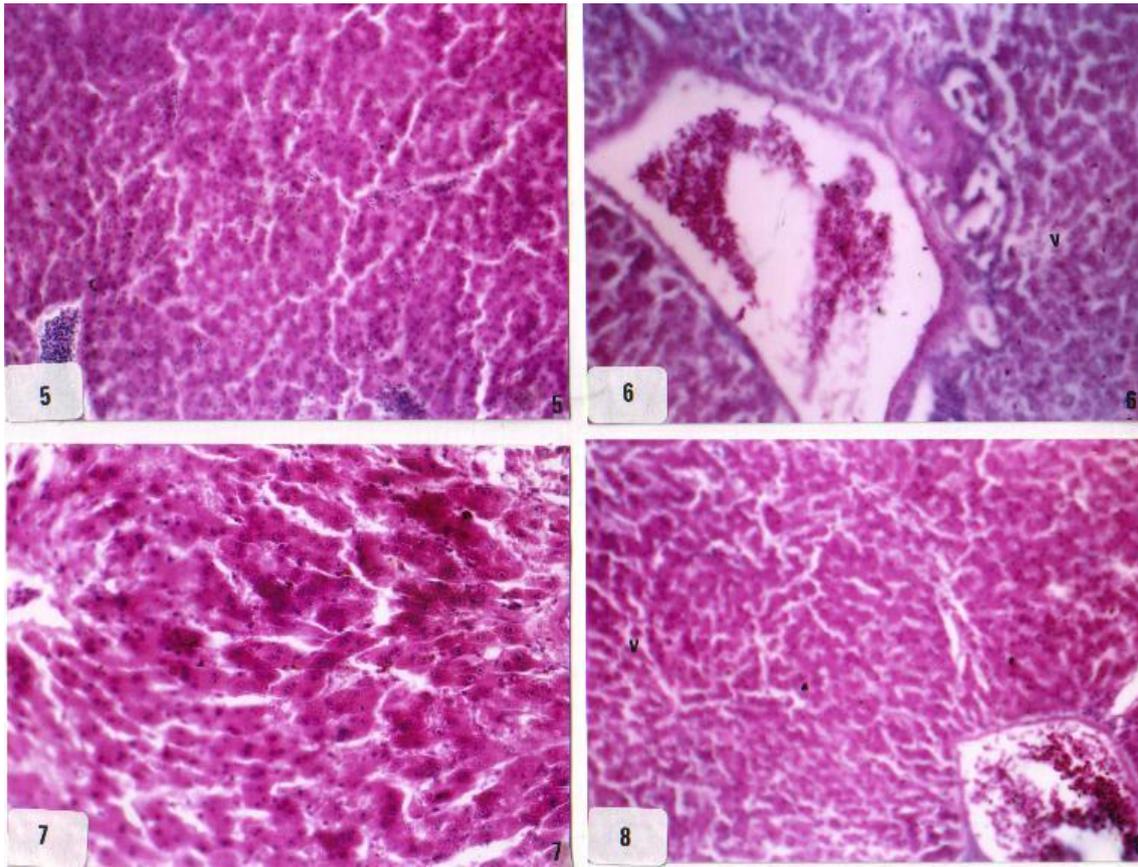


Fig. 5: section in liver of quail treated with vit. C for 3 & 6 weeks and recovery period showing normal hepatocytes.

(Hx & X 250)

Figs. 6,7: section in liver of quail treated with imidacloprid and vit. C showing highly dilated HPV, hepatic artery and degenerated bile duct, vacuolated cells and dilated sinusoidal spaces.

(Hx & X 250)

Fig. 8: section in liver of quail treated with imidacloprid and vit. C after recovery period, showing good architecture of hepatocytes.

(Hx & X 250)

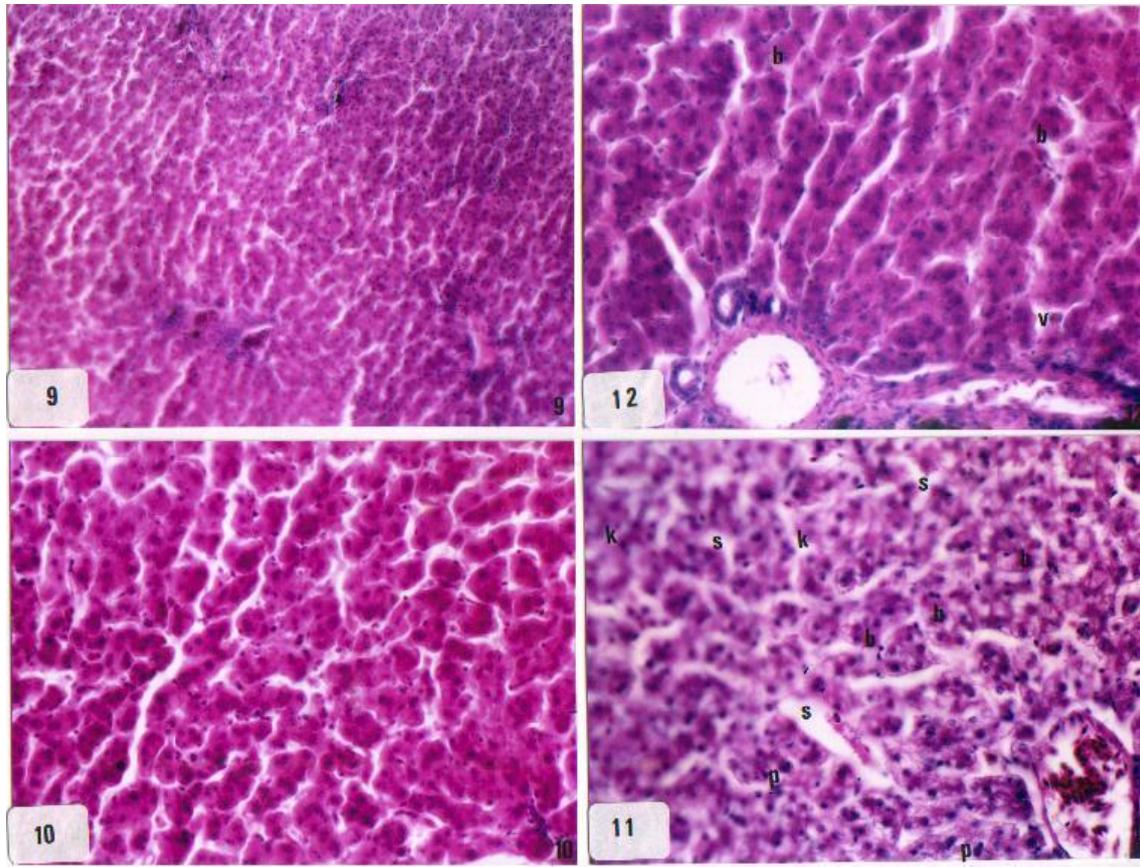


Fig. 9: section in liver of quail treated with glutathione for 3 & 6 weeks and 3 recovery weeks, showing normal liver structure.

(Hx & X 250)

Figs. 10,11: section in liver of quail treated with imidacloprid and glutathione for 3 weeks showing, ballooning, enlarged hepatocytes and faintly stained hepatocytes, wide sinusoidal spaces, pyknotic nuclei, vacuolated cytoplasm and nuclei.

(Hx & X 250)

Fig. 12: section in liver of quail treated with imidacloprid and glutathione for 3 recovery weeks showing portal space containing normal hepatic portal vein, hepatic artery and bile duct with some vacuolated hepatocytes.

(Hx & X 250)

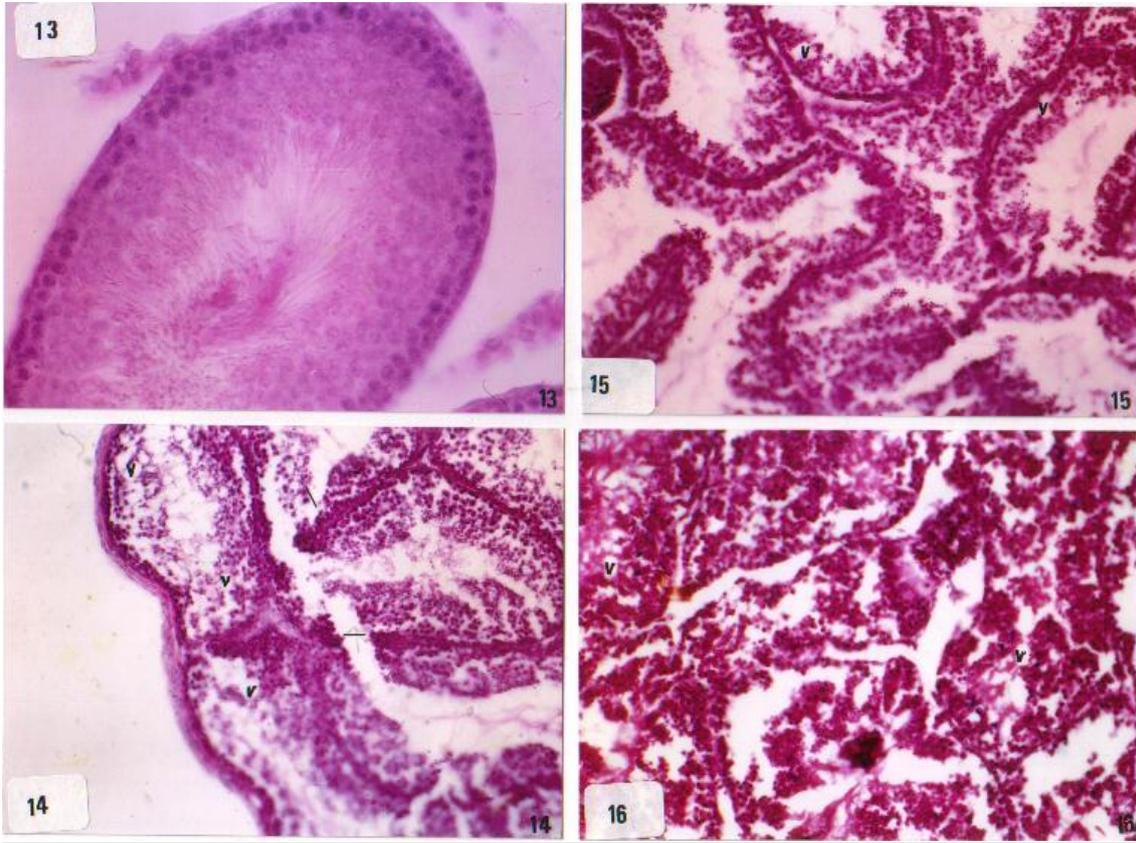
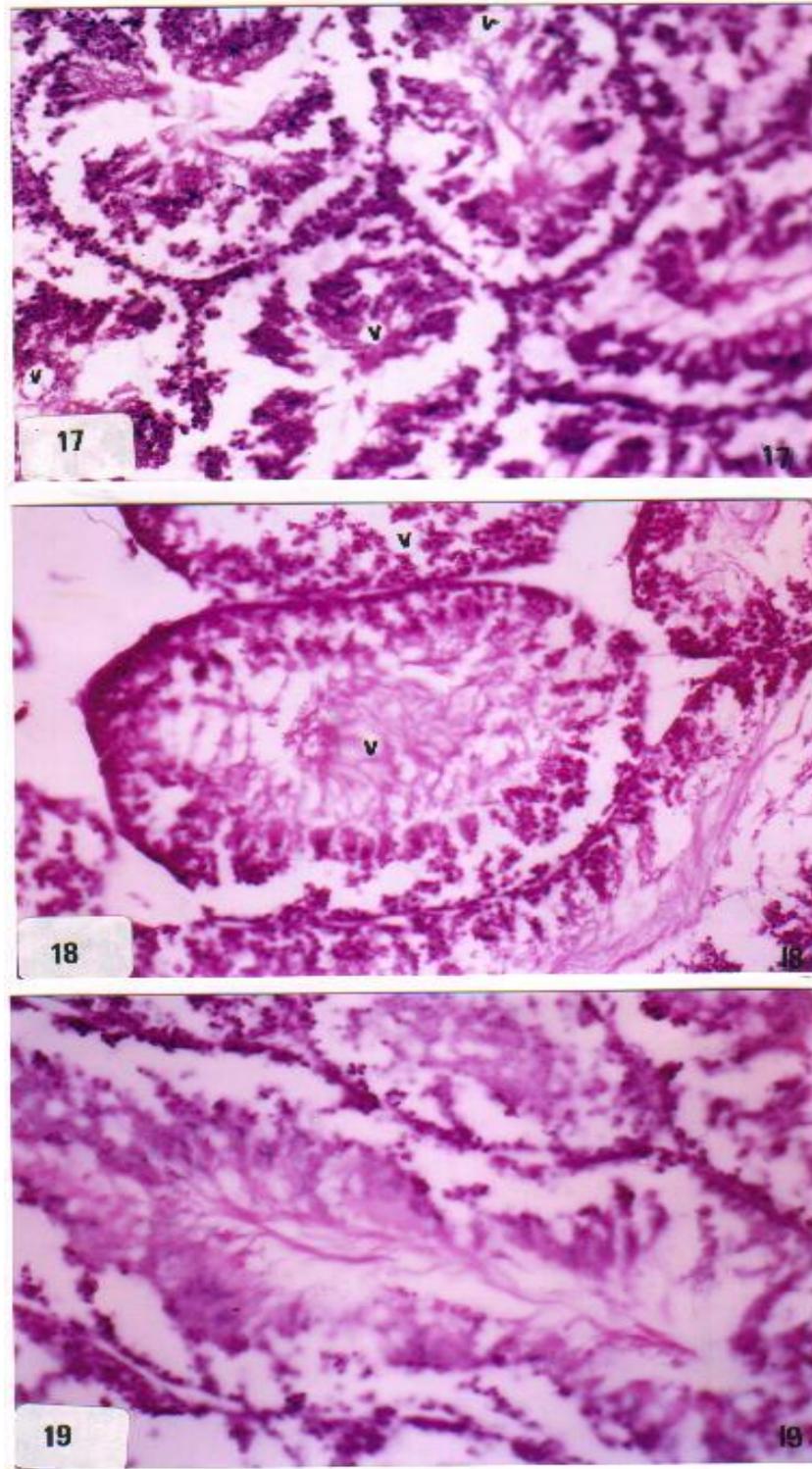


Fig. 13: section in control testis of quail showing normal structure of the testis.
(Hx & X 250)

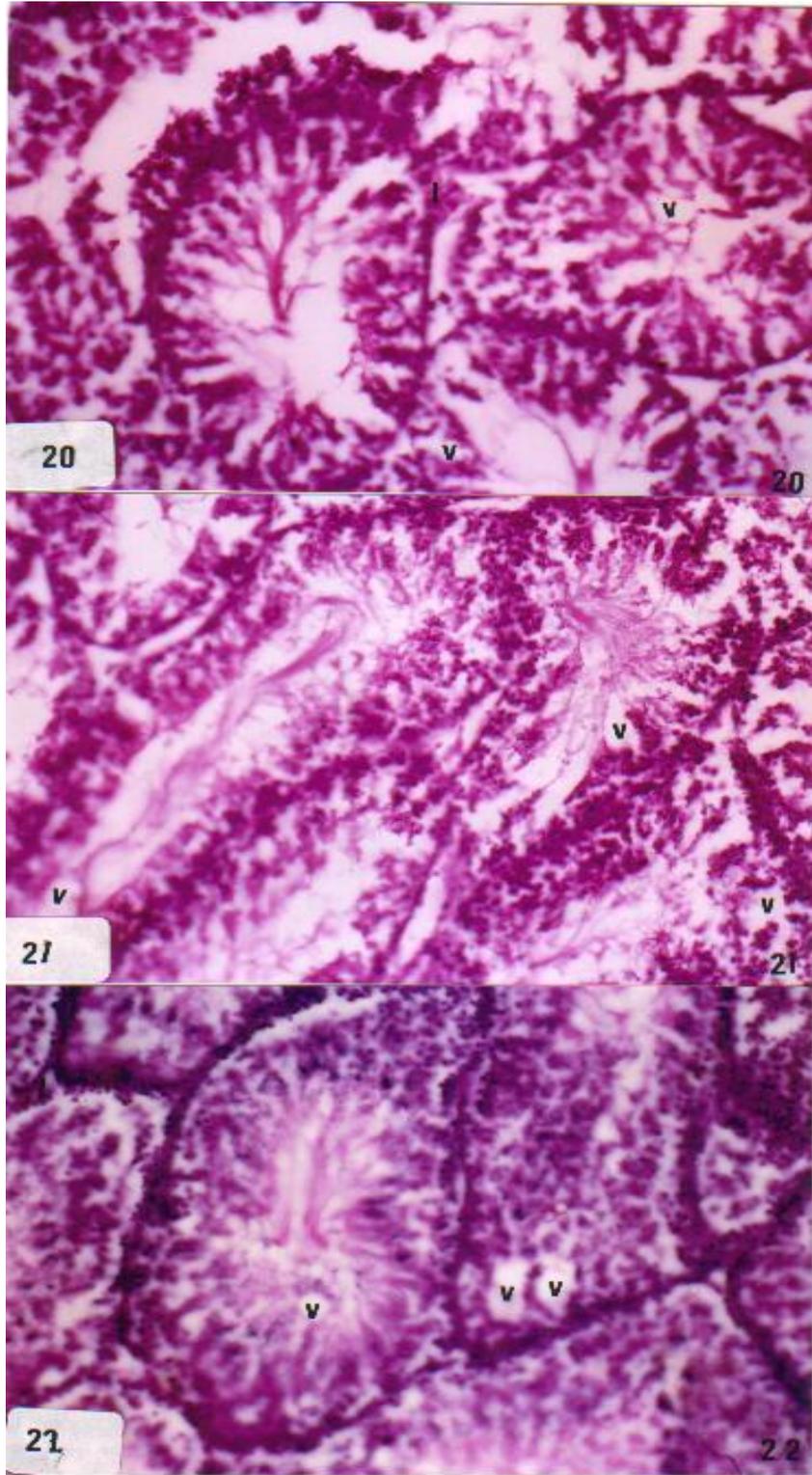
Figs. 14,15,16: section in testis of quail treated with imidacloprid for 3 & 6 and 3 recovery weeks showing bizarre cells, thickened tunica albuginea, disappearance of spermatogenic cells. The tubules are devoid of sperms. Some pyknotic nuclei could be observed. Rupture of basement membrane, vacuolation and debris of sperm.

(Hx & X 250)



Figs. 17,18,19: section in testis of quail treated with imidacloprid and vit. C for 3 & 6 and 3 recovery weeks showing debris of sperms and ruptured basement membrane, few Leydig cells, some tubules contained sperm.

(Hx & X 250)



Figs. 20,21: section in testis of quail treated with imidacloprid and glutathione for 3 & 6 weeks showing undifferentiated cells, vacuolation and debris of sperms.

(Hx & X 250)

Fig. 22: section in testis of quail treated with imidacloprid and glutathione for 3 weeks recovery showing good architecture, Leydig cells while signs of vacuolation could be detected.

(Hx & X 250)

Discussion

Numerous studies had been performed to demonstrate the effect of insecticides on birds and animals, different pathological changes were noticed in liver and testis of Japanese quail treated with different dose levels, these changes included: large areas of vacuolation, fatty degeneration, large areas of necrosis, congested sinusoidal spaces and different types of affected nuclei. Our results are in accordance with those reported by Abo El-Ezz *et al.* (1986), El-Sayed (1998), Ahmed, (2000) and El-Khatib *et al.* (2003).

El-Swak (1990), treated hens with an organophosphorous insecticide profenofos and he noticed that liver showed congestion and degeneration. El-Khatib *et al.* (2003), observed that liver of treated rats exhibited severe histopathological alteration manifested by kupffer cells activation, necrotic changes in hepatocytes, small focal areas of hepatic necrosis and infiltration with mononuclear cells after treated with butachlor.

Environmental insecticide pollution is considered a great problem in different countries. Many cases of poisoning have been reported in man, animal and birds fed on diets polluted with such compounds (Kerr *et al.*, 1987). The liver is the major site of metabolism including detoxification and activation of many compounds (Guyton 1995). The present study showed that imidacloprid produced severe structural alteration in hepatocytes and testis of Japanese quail.

Sammour and Hamouda (1997) concluded that Organo-phosphorous insecticides have more or less common toxic effects which are not species specific and therefore are very liable to occur in man if exposed to such compounds.

The swelling and vacuolation of the cells are most probably due to the retention of fluid inside the hepatocytes resulting in what is known as cloudy swelling or hydropic degeneration which is said to be either due to reduction of the energy necessary for regulation of ion concentration of the cells (Elwi, 1967),

mild or short term anoxia (Hruban *et al.*, 1972) or metabolic stress (De Duve & Wattiaux, 1996).

The earliest stage of degeneration in the study is most probably represented in the enlarged distorted hepatocytes which contained deeply stained eosinophilic cytoplasm and deeply stained nuclei in few cells denoting swelling and decreased activity of these cells. Similar cells were described by Abu-Hadeed (1978) in the liver of the Nile fish when exposed to the organophosphorous compound Curacron. However, he attributed them to coagulative necrosis of the cells due to chronic toxicity.

According to Walter & Israel (1974), cellular infiltration may be due to the presence of necrotic cells which act as an irritant substance attracting the inflammatory cells.

Mahiuddin & Ahmed (1986), described bile duct proliferation and attributed it to second grade liver cirrhosis.

Concerning the efficiency of glutathione in the reduction of toxicity, the study revealed that this biochemical compound has provided protection against imidacloprid toxicity as indicated by improvement of the histological changes and in the serum chemistry of SGOT and SGPT. The explanation of this improvement is linked to the sulphhydryl group of the amino acid cysteine, which prevents oxidation of endogenous mitochondrial, and microsomal enzymes which participate in the toxicity production (Bhanumathi and Devi, 1994).

The liver necrosis is caused by the covalent binding of reactive electrophilic metabolites to liver macromolecules. Normally, these reactive metabolites are detoxified by combining with hepatic glutathione, which when becomes exhausted, the metabolites bind to liver macromolecules which resulted in hepatocytes damage. This biochemical entity has been confirmed by Gardner and Cluff (1970) who stated that hepatic necrosis produced by the ingestion of large quantities of toxic agents can be prevented

or at least attenuated, by the administration of substances such as cysteamine, which bind the electrophilic metabolites and prevent them from reacting with hepatic proteins with resultant hepatic necrosis. Histologically, in this study, glutathione resulted in normal appearance of liver and testis tissues.

Moldeus and Jernstrom (1983), stated that when hepatic tissue is incubated in a medium containing sulphhydryl containing amino acids, the hepatic tissue viability would be maintained. Thus, it might be suggested that glutathione protection against tissue toxicity depends on increased glutathione availability. Many authors observed the potency of glutathione in reducing tissue damage when provided prior or even with toxic agents. Bhanumathi and Devi (1994), concluded that not only the sulphhydryl group mechanism that protect tissue damage, but glutathione peroxidase enzyme, of which selenium is an integral component, provides a second line of defense. They added that the biochemical action of glutathione seems to be prevention of peroxidative damage to cellular and subcellular elements.

Vitamin c is a water-soluble antioxidant, which can directly scavenge superoxide and hydroxyl radicals; it is primarily active in the cytosine and plasma (Guyton 1995).

The imidacloprid action potential at the presynaptic ending could be explained by the release of the transmitter of neurons from these pools and binds to receptors on the postsynaptic membrane creating a specific change in the conductivity of the membrane for certain ions. All pre and postsynaptic effects are ultimately due to changes in the permeability of the neural membrane.

The semipermeable membrane of a resting cell is polarized as a result of specific distribution of ions between intra and extra cellular space. The ions responsible for polarization are Na⁺, K⁺, Cl⁻ and sometimes Ca⁺⁺ (Schroeder and Flattum, 1984).

El-Kashoury (1999), evaluated the effect of imidacloprid at low concentration in male albino rats, either alone or

combined with another insecticide as Profenofos or carbosulfan, the obtained results recorded a reduction in liver, kidney function. Also, on cholinesterase activity, total lipids level and no change in plasma glucose level could be noticed. In case of insecticides combination a remarked elevation in glucose level, decrease in ALT and ALP activities, and no change in total protein and albumin could be recorded.

El-Hamady, *et.al.*, (2001), evaluated the effect of imidacloprid insecticide at acute and subacute doses on japanese quail for 30 days. These results showed hepatic and renal toxicity, in addition no haematological, biochemical changes were observed on birds exposed to sublethal doses of insecticide even at the higher tested dose.

From the above-mentioned results, it was clear that the exposure of Japanese quails to the applied insecticide (imidacloprid) caused many histopathological changes in liver and testis. Finally, it is recommended that the use of imidacloprid insecticide must be limited due to its hazardous effect to the non-target species including the farmers and the farm birds even the technicians who exposed to it and also residue in the agricultural products.

Mild recovery was observed in liver and testis treated with vitamin C and imidacloprid which indicating the antioxidant effect of vitamin C. It's well recommended to use either vitamin C or glutathione as antioxidants to reduce the toxic effect of imidacloprid.

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التأثير الوقائي لفيتامين (ج) والجلوتاثيون على التغيرات المرضية النسيجية التي يحدثها المبيد الحشري (اميداكلوبريد) في كبد وخصية السمان الياباني

أميمة سليمان عيسى

المركز القومي لبحوث وتكنولوجيا الإشعاع -هيئة الطاقة الذرية

لا شك أن استخدام المبيدات الحشرية من المشاكل التي تواجهنا الآن وذلك لما تحدثه من أضرار كثيرة سواء للبيئة أو للكائنات الحية حيث تستخدم هذه المبيدات فى رش كثير من الخضروات والفاكهة والحبوب ويتأثر بها كل الكائنات الحية التي تتعامل مع البيئة.

ويطالعنا العلماء يوميا باكتشاف مبيد جديد حيث تتطور قدرة الحشرات الضارة والفطريات على مقاومة كثير من المبيدات القديمة وتكتسب مناعة ضدها. وقد أجرى هذا البحث لدراسة بعض المخاطر التي تتعرض لها الطيور ومنها السمان الذى يعد واحدا من أفضل نماذج الطيور التجريبية المنتشرة فى مصر وذلك بتعرضها إلى جرعات مخففة من المبيد الحشري (إميدا كلوبريد) لذى يعتبر واحدا من أحدث المبيدات الحشرية الموجودة والشائعة الإستخدام حاليا فى مصر .وفى محاولة لتفادى سمية المبيد على الطائر فقد إستخدم نوعين مختلفين من مضادات الأكسدة أحدهما كمادة خارجية (فيتامين ج) والآخر كمادة طبيعية يفرزها الكبد (الجلوتاثيون فى محاولة لمعرفة مدى قدرتهم لتفادى أو تقليل سمية المبيد. أستخدم فى هذه التجربة عدد 90 من ذكور السمان قسمت إلى ستة مجاميع كالتأتى: الأولى :مجموعة ضابطة.

الثانية :مجموعة معالجة بفيتامين ج.

الثالثة :مجموعة معالجة بالجلوتاثيون

الرابعة :مجموعة معالجة بجرعة 50/1 من الجرعة النصف مميتة للمبيد الحشري أميداكلوبريد.

الخامسة :مجموعة معالجة بالمبيد الحشري أميداكلوبريد بالإضافة إلى فيتامين ج.

السادسة :مجموعة معالجة بالمبيد الحشري أميداكلوبريد بالإضافة إلى الجلوتاثيون.

وقد تم معالجة المجموعات لمدة 3 ، 6 أسابيع على التوالى وتم ذبح خمسة من الطيور من كل مجموعة فى نهاية فترة العلاج وتركت بقية الطيور لمدة 3 أسابيع بدون أى معالجة كفترة إستشفاء.

وقد أظهرت النتائج أن المبيد قد أحدث تأثير بالغ على تركيب الكبد حيث وجدت مظاهر للتحلل فى العديد من الخلايا على شكل فراغات فى السيتوبلازم وتقلص فى الأنوية كما حدث إحتقان فى الأوردة البابية وكذلك وجود العديد من الفراغات الدموية . أما عن الخصية فقد ظهرت علامات التحلل النووى فى بعض خلايا الخصية

كما ظهرت بعض تجمعات من النفايات الخلوية فى البعض الآخر من الخلايا وخاصة خلايا لايدج.

ولقد لوحظ أن إستخدام فيتامين ج أو الجلوتاثيون يقلل كثيرا من أخطار المبيد خاصة بعد مرور فترة الإستشفاء مما يوضح أن إستخدام إحدى هاتين المادتين فى مزارع الطيور له تأثير إيجابى يمكنه من وقاية الأنسجة التى تواجه مثل هذه المبيدات .