MTHFR gene polymorphisms as risk factors for colorectal cancer among Egyptian patients.

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Abstract

Objective: Polymorphisms in methylenetetrahydrofolate reductase (MTHFR), such as MTHFR C677T and A1298C, are associated with several cancers. This study aimed to evaluate the effects of MTHFR polymorphisms on colorectal cancer risk in a population from Damitta, Egypt. Methods. This hospital-based case-control study was conducted during 2008-2010; 64 colon cancer cases and 90 controls were enrolled. Information was collected and blood samples were obtained for assay of MTHFR C677T and A1298C polymorphisms by polymerase chain reaction-single strand conformation polymorphism (PCR-SSCP) and PCR-restriction fragment length polymorphism (PCR-RFLP) techniques. Associations between variables of interest and colorectal cancer were assessed using conditional logistic regression. Results: Increased risk of colorectal cancer was associated with the MTHFR C677 TT genotype of C677T polymorphism (OR(adj) = 24.0; 95% CI: 1.34-429.1; P value for interaction = 0.001). The 1298AC genotype and C allele was associated with a statistically significant lower risk among subjects (OR, 3.85; 95% CI, 1.78-8.33; P value for interaction=.0005 and OR, 1.88; 95% CI, 1.16-3.059 P value for interaction=0.01), respectively. MTHFR 1298 AA genotype and A allele was found to be associated with a significantly decreased risk for colorectal cancer (OR = 0.25, 95% CI 0.11-0.52; P value for interaction= 0.005 and OR = 0.52, 95% CI 0.32-0.85 P value for interaction=0.01), respectively. There was no clear relation between colorectal adenomas and those with the 1298 CC genotype. The combined CC, AA [corrected] genotypes and the CT+AA [corrected] genotypes and the TT+ AC were associated with a statistically significant lower risk for developing colorectal cancer (P value for interaction= 0.03, 0.02, 0.001), respectively.

CONCLUSIONS: The findings suggest an interaction between the MTHFR genotype and colorectal adenomas among Egyptian patients.

Introduction

Colorectal cancer is one of the most common cancers in the world, accounting for nearly 10% of new cases of all cancers (Parkin, et al., 2005). The incidence of colorectal cancer varies substantially worldwide, with high rates in Western countries and low rates in African and Asian countries in general (Stewart et al., 2006). Methyleneetetrahydrofolate reductase (MTHFR) is an enzyme which catalyzes the reduction of 5, 10-methyleneetetrahydrofolate to 5-methyltetrahydrofolate (MTHF), the predominant circulatory form of folate and carbon donor for the re-methylation of homocysteine to methionine (Donnelly, 2001 and Fowler, 2005). Several polymorphisms have been described that result in amino acid changes, which could...
lead to altered MTHFR enzymatic activity (Donnelly, 2001 & Toffoli and De Mattia, 2008). The best-characterized being the C677T, a valine-to-alanine substitution at amino acid 226, which was reportedly linked with moderate hyperhomocysteinemia when folate levels are low in homozygotes (677T/T) compared with heterozygotes or non-carriers (Friedman et al., 1999 & Almawi et al., 2004). The A1298C missense mutation is an additional MTHFR gene polymorphism located in the enzyme regulatory domain (van der Put et al., 1998 & Friedman et al., 1999), but unlike the C677T found within the enzyme catalytic domain, it does not result in either a thermolabile protein or with increased homocysteine (Hanson et al., 2001 & Friso et al., 2002). It is considered that alteration of DNA methylation is associated with carcinogenesis and is not just an accompanying phenomenon (Choi & Mason, 2002). Two functional, common polymorphisms (C677T and A1298C) are known in the MTHFR gene. In this review, we evaluate the consistency of reported associations of the MTHFR polymorphisms with colorectal cancer.

**Aim of the work:**
The objective of the present work is to detect the relation of MTHFR C677T and A1298C polymorphisms to colorectal cancer.

**Subject and methods**

**Subjects:**
The present study was conducted on a total of 64 adult patients aged between 23 – 73 years with a mean 47.73 ±12.72, the number of males is 38 aged between 23 – 73, and the number of females is 26 aged between 32 - 73. Patients selected from the Cancer Institute, Mansoura University, Egypt in the period between January and September 2009. In addition, 90 normal healthy control randomly selected from blood donors. The samples were taken from the blood of all patients and controls after their routine check up by their own consent. The work took the approval of ethical committee of National Cancer Institute.

**Method:**

**DNA extraction and purification**

After obtaining informed consent from all cases and controls, venous blood samples (3 ml) were collected on EDTA (ethylenediamine tetra acetate) containing tubes, DNA was extracted promptly using DNA extraction and purification kit (Gentra Systems, USA) according to manufacturer's instructions and then stored at -20°C till use.

**PCR amplification**

MTHFR C677T genotype analysis was performed by PCR-RFLP analysis.

The primer sequences for C677T were: Forward: (5´- TGA AGG AGA AGG TGT CTG CGG GA- 3´) and Reverse: (5´-AGG ACG GTG CGG TGA GAG TG- 3´) (Gibeo BRL). For PCR the following procedure was used: preliminary denaturation at 95°C for 3 min, 5 cycles: denaturation at 94°C for 1 min, annealing at 64°C for 1 min, synthesis at 72°C for 30 sec, next 30 cycles were run: denaturation at 94°C for 45 sec, annealing at 62°C for 45 sec, synthesis at 72°C for 25 sec and final synthesis at 72°C for 7 min. Digestion was performed by restrictase HinfI (Fermentas, USA) at 37°C for 4 h. For evaluation of MTHFR A1298C polymorphism, the following primers were used: Forward: (5´- CTT TGG GGA GGT GAA GGA CTA CTA C- 3´) and Reverse: (5´-CAC TTT GTG AGC ATT CCG GTT TG- 3´) (Gibeo BRL). The PCR procedure was composed of preliminary denaturation at 95°C for 2 min, 5 cycles: denaturation at 95°C for 1 min, annealing at 55°C for 2 min, synthesis at 72°C for 2 min, next 32 cycles were run: denaturation at 95°C for 75 sec, annealing at 55°C for 75 sec, synthesis at 72°C for 90 sec and final synthesis at 72°C for 6 min. Restrictase
MboII was used for digestion at 37°C for 7h. (Siemianowicz et al., 2003).

Detection of amplified and digested products

The entire reaction volume plus 5 µl of bromophenol blue track dye were loaded into 2% agarose gel (Boehringer Mannheim) containing ethidium bromide. Gels were electrophoresed for 30 minutes at 100 V, photographed under UV light (320 nm) and then scored for the presence or absence of an allele specific band.

Statistical analysis:

Data were processed and analyzed using the Statistical Package of Social Science (SPSS, version 10.0). The frequency of studied allelic polymorphisms were compared between groups by using Fisher's exact test (modified Chi square test) and Odds ratio. A value of P <0.05 was considered to be significant.

Results

Comparing studied cases of Colorectal cancer to that of Healthy controls regarding the distribution of MTHFR C677T and A1298C genotypes and alleles(table 1, fig.1,2) showed that cases of colorectal cancer had a significantly higher frequency of the homozygous mutated TT genotype of C677T polymorphism (10.9% vs. 0 %, p=0.001). On the other hand they had a significantly lower frequency of heterozygous mutated AC genotype and C allele of A1298C polymorphism compared to controls (82.8% vs. 55.6%, p=0.0005 and 41.4% vs. 27.8%, p= 0.01, respectively), while they had a significantly higher frequency of wild AA genotype and A allele of A1298C polymorphism compared to controls (17.2% vs. 44.4%, p=0.0005 and 58.6% vs. 72.2%, p=0.01, respectively).

Comparing all studied cases of colorectal cancer to that of healthy controls regarding their combined genotypes of C677T and A1298C polymorphisms of MTHFR gene (table 2), it is noted that cases had a significantly lower frequency of combined (CC and AA) genotypes and (CT and AA) genotypes compared to controls (6.25% vs. 18.9%, p= 0.03 & 11.0 % vs. 25.6 %, p= 0.02 ).While they had a significantly higher frequency of combined (TT and AC) genotypes
Table (1): Comparison between all cases colorectal cancer and healthy controls regarding their allele frequency and genotype distribution of C677T and A1298C polymorphisms of MTHFR gene.

<table>
<thead>
<tr>
<th></th>
<th>cases</th>
<th>Healthy controls</th>
<th>(P)</th>
<th>OR(95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>n(%)</td>
<td>n(%)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C677T</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Genotypes</td>
<td>64(100.0)</td>
<td>90(100.0)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>CC</td>
<td>26(40.6)</td>
<td>40(44.4)</td>
<td>0.7</td>
<td>0.87(0.4-1.6)</td>
</tr>
<tr>
<td>CT</td>
<td>31(48.4)</td>
<td>50(55.6)</td>
<td>0.4</td>
<td>0.7(0.38-1.3)</td>
</tr>
<tr>
<td>TT</td>
<td>7(10.9)</td>
<td>0(0.0)</td>
<td>0.001**</td>
<td>24.0(1.34-429.1)</td>
</tr>
<tr>
<td>Alleles</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>C</td>
<td>83(64.9)</td>
<td>130(16.7)</td>
<td>0.2</td>
<td>0.7(0.43-1.17)</td>
</tr>
<tr>
<td>T</td>
<td>45(35.1)</td>
<td>50(27.8)</td>
<td>0.2</td>
<td>1.3(0.85-2.27)</td>
</tr>
<tr>
<td>A1298C</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Genotypes</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>AA</td>
<td>11(17.2)</td>
<td>40(44.4)</td>
<td>0.0005**</td>
<td>0.25(0.11-0.52)</td>
</tr>
<tr>
<td>AC</td>
<td>53(82.8)</td>
<td>50(55.6)</td>
<td>0.0005**</td>
<td>3.85(1.78-8.33)</td>
</tr>
<tr>
<td>CC</td>
<td>0(0)</td>
<td>0(0)</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Alleles</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>A</td>
<td>75(58.6)</td>
<td>130(72.2)</td>
<td>0.01*</td>
<td>0.52(0.32-0.85)</td>
</tr>
<tr>
<td>C</td>
<td>53(41.4)</td>
<td>50(27.8)</td>
<td>0.01*</td>
<td>1.88(1.16-3.05)</td>
</tr>
</tbody>
</table>

n= number of cases, (%)= percentage of cases, OR(95% CI) = Odds ratio & 95% confidence interval
Significance using Fisher's Exact test: * p< 0.05 (significant)
** p < 0.001 (extremely significant)
Table (2): Comparison between all cases of colorectal cancer and healthy controls regarding the frequency of their different combined genotypes of C677T and A1298C polymorphisms of MTHFR gene.

<table>
<thead>
<tr>
<th>Combined genotypes</th>
<th>cases n(%)</th>
<th>Healthy controls n(%)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total</td>
<td>64(100.0)</td>
<td>90(100.0)</td>
<td></td>
</tr>
<tr>
<td>CC and AA</td>
<td>4(6.25)</td>
<td>17(18.9)</td>
<td>0.03*</td>
</tr>
<tr>
<td>CC and AC</td>
<td>22(34.4)</td>
<td>23(25.6)</td>
<td>0.28</td>
</tr>
<tr>
<td>CT and AA</td>
<td>7(11.0)</td>
<td>23(25.6)</td>
<td>0.02*</td>
</tr>
<tr>
<td>CT and AC</td>
<td>24(37.5)</td>
<td>27(30.0)</td>
<td>0.38</td>
</tr>
<tr>
<td>TT and AC</td>
<td>7(11.0)</td>
<td>0(0.0)</td>
<td>0.001**</td>
</tr>
</tbody>
</table>

n= number of subjects, (%)= percentage of subjects. * p = < 0.05 (significant) ** p = < 0.001 (extremely significant)

**Fig (1)** Digestion of PCR product of C677T polymorphism of MTHFR gene using HinfI enzyme. which digests the 198- bp fragment into 175- and 23- bp fragments; the small 23- bp fragment has run off the gel. Lane M: DNA size marker (50 bp), lane 1,4, and 6 (wild type CC is found which appears at 198 bp), lane 2,5, and 7 (heterozygous mutated genotype CT which has 198, 175, 23 bp fragments), lane 3 (homozygous mutated genotype TT which has 175, 23 bp fragments); the small 23- bp fragment has run off the gel.
Fig (2): Digestion of PCR product of A1298C polymorphism of MTHFR gene using MobII enzyme which digests the 163 bp fragment the wild-type AA into five fragments 56, 31, 30, 28 and 18; the small 18, 28, 30, 31 bp fragments have run off the gel. Lane M: DNA size marker (50 bp), Lane 2, 5, and 6 (wild type AA), lane 1, 3, 4 and 7 (heterozygous mutated type AC).

Discussion

Colorectal cancer is the third most common cancer and the fourth most frequent cause of cancer death world-wide, accounting for 9.4% and 7.9% of all cancer cases and deaths, respectively. In more developed countries, colorectal cancer is the second most frequent cancer type and cause of cancer death, accounting for 13.3% and 11.7% of the cases, respectively. There is up to a 20-fold difference in incidence between the high rates experienced in the economically developed countries of Australia, Europe, Japan and North America versus those of central Africa, the Indian sub-continent and southern Asia (Ferlay et al., 2004).

Methylenetetrahydrofolate reductase (MTHFR) is a key enzyme regulating folate metabolism. It irreversibly converts 5,10-methylenetetrahydrofolate to 5-methyltetrahydrofolate, which is the major form of folate in blood (Sharp & Little, 2004). The substrate of MTHFR, 5,10-methylenetetrahydrofolate, is required for conversion of deoxyuridylate to thymidylate. Depletion of 5,10-methylenetetrahydrofolate results in uracil misincorporation into DNA, and removal of this abnormal base may lead to single and double strand breaks (Blount et al., 1997; Duthie, 1999). Furthermore, insufficient thymidylate can increase DNA misrepair, resulting in overall DNA damage in the cell (Duthie et al., 2000). On the other hand, 5-methyltetrahydrofolate provides the methyl group for methylation of homocysteine to methionine. Imbalanced DNA methylation, i.e., global genomic hypomethylation and methylation of usually unmethylated C—phosphate—G—site (CpG) sites, has been implicated in colorectal carcinogenesis (Toyota et al., 1999 and van Rijnsoever et al., 2002).

DNA methylation is a crucial epigenetic determinant in gene expression, maintenance of DNA integrity and stability, chromatin modifications and development of mutations did find a significant excess of 677TT individuals among patients with colorectal carcinomas (CRC). Indeed, induction of DNA damage and disruption of its DNA integrity, impaired DNA repair and hypermutability are generally considered to be the primary mechanisms by which folate deficiency enhances colorectal carcinogenesis (Duthie, 1999; Choi & Mason, 2002).

The current study aims mainly to investigate the association of C677T and A1298C polymorphisms of Methylenetetrahydrofolate reductase (MTHFR) gene and their combined effect with colorectal cancer among Egyptian cases of colorectal cancer.
This data showed that homozygous mutated TT genotype of C677T polymorphism had a significantly higher frequency among cases of colorectal cancer compared to controls. At position 677, TT genotype frequency was 10.9% with an odds ratio (OR) of 24.0 [95% confidence interval (CI) 1.34 - 429.1, P = 0.001]. Thus TT genotype may be considered as genetic risk factor for colorectal cancer. This results are in agreement with the following results: El Awady et al. (2009), Levine et al. (2010), Cui et al. (2010). El Awady et al. (2009) study among 35 cases of Colorectal carcinomas (CRC) and 68 healthy controls suggested that at least one C to T polymorphism at 677MTHFR gene is required to significantly increase the risk for CRC development. Further large scale studies are required to confirm the present findings. Frequencies of heterozygous and homozygous polymorphisms of MTHFR at positions 677 in carcinoma tissues were always the highest. At position 677, TT and CT genotype frequencies were 17% and 66% with an odds ratio (OR) of 11 [95% confidence interval (CI) 2.39-50.59] and OR 8.34 [95% CI 2.97-23.92], respectively, in carcinoma tissues. While in the germ line of patients the genotype frequencies of 677TT and CT were 6% and 54% with OR 1.57 [95% CI 0.26-9.51] and 2.99 [95% CI 1.25-7.12], respectively, compared to controls (6% and 29%, respectively) in Egypt. Levine et al. (2010) study in, Los Angeles, USA. Suggested that the MTHFR C677T TT genotype is associated with a 15% to 18% reduction in colorectal cancer risk, but it is not clear if other variants of the gene are associated with colorectal cancer risk. The 677 TT genotype was associated with a decreased risk of microsatellite-stable/microsatellite-low tumors (OR, 0.69; 95% CI, 0.49-0.97) and an increased risk of microsatellite-high tumors (OR, 2.22; 95% CI, 0.91-5.43; P(interaction) = 0.01), as well as an increased risk of proximal cancers and a decreased risk of distal and rectal cancers (P(interaction) = 0.02). No other single nucleotide polymorphism was associated with risk overall or within subgroups.

Cui et al. (2010) study was designed to investigate an association between the methylenetetrahydrofolate reductase (MTHFR) C677T polymorphism and the risk of gastric and colorectal cancer in the Korean population. The MTHFR 677TT genotype showed a weak opposite association with colorectal cancer compared to the homozygous CC genotype [adjusted age and sex odds ratio (OR) = 0.792, 95% confidence interval (CI) = 0.638-0.984, P = 0.035]. Subjects with the MTHFR 677CT showed a significantly reduced risk of gastric cancer compared those with the 677CC genotype (age- and sex-adjusted OR = 0.810; 95% CI = 0.696-0.942, P = 0.006). It is observed no significant interactions between the MTHFR C677T polymorphism and smoking or drinking in the risk of colorectal cancer. The T allele was found to provide a weak protective association with gastric cancer and colorectal cancer. In contrast, these results are in disagreement with the present results, Le Marchand et al., (2005), Kono & Chen, (2005), Mokarram et al (2008), Le Marchand et al. (2005) founded that an inverse association between colorectal cancer risk and the TT genotype, with odds ratios (OR; and 95% confidence intervals) for the CC, CT, and TT genotypes of 1.00, 1.01 (0.84-1.21), and 0.77 (0.58-1.03), respectively. This association was similar in both sexes in Hawaii, USA. Such an association had been seen in some previous studies (Curtin et al., 2004, Ulrich et al., 1999). Kono & Chen (2005) founded that a Decreased risk of colorectal cancer associated with the 677TT genotype has fairly consistently been observed in Japan. Mokarram et al. (2008) showed an increased risk of tumor methylation was associated with the CT genotype (OR = 2.5; 95% CI, 1.1-5.6) in comparison to controls with the CC genotype in Iran. Regarding genotypes and alleles of A1298C polymorphism, this data showed that a significantly lower frequency of heterozygous mutated AC genotype and C allele of A1298C polymorphism compared to controls (82.8% vs. 55.6%, p=0.0005 and 41.4% vs. 27.8%, p= 0.1, respectively),
while they had a significantly higher frequency of wild AA genotype and A allele of A1298C polymorphism compared to controls (17.2% vs. 44.4%, p=0.0005 and 58.6% vs. 72.2%, p=0.01, respectively). There is no significant frequency in homogenous mutated CC of 1298 genotype. Our results are in agreement with results of El Awady et al.(2009), Chandy et al., (2010). El Awady et al. (2009) founded that Frequencies of MTHFR677T and 1298C alleles were significantly higher among cases of CRC tumor tissues (50% and 56%, respectively) than germ line alleles in CRC patients (33% and 41%, respectively) and healthy controls (21% and 35%, respectively) in Egypt. Chandy et al. (2010) study was under taken to analyse the association of the MTHFR gene polymorphisms 677 C-->T and 1298 A-->C and risk of colorectal cancer (CRC) in India. The OR for 677 CT was 1.18 (95% CI 0.59-2.32, P = 0.642), OR for 1298 AC was 1.68 (95% CI 0.92-3.08, P = 0.092) and for 1298 CC was 0.45 (95% CI 0.18-1.12, P = 0.081). The OR for the combined heterozygous state (677 CT and 1298 AC) was 1.18 (95% CI 0.52-2.64, P =0.697).

Regarding genotypes and alleles of A1298C polymorphism, we found our cases of colorectal cancer are in disagreement with the results of, Curtin et al.(2004) who founded that the 677CC/1298CC (wild-type/variant) genotype was associated with a statistically significant lower risk among women (OR, 0.6; 95% CI, 0.4-0.9) but not men. When the polymorphisms were considered individually, for A1298C a significant risk reduction associated with the homozygous variant CC genotype was seen among women only (OR, 0.6; 95% CI, 0.5-0.9), this study supports recent findings that the MTHFR A1298C polymorphism may be a predictor of colon cancer risk and have functional relevance. The possible interaction with hormone-replacement therapy warrants additional investigation and these results are in agreement with our result for AA and AC genotypes.

Zhang et al. (2007) supported the role of the A1298C polymorphism in MTHFR as prognostic marker in female patients with metastatic colon cancer. Further studies are needed to confirm these findings. The A1298C polymorphism showed statistically significant differences in overall survival (OS) in female, but not male, patients with metastatic colon cancer (log-rank test, P = .038). Among females, Overall survival (OS) was greater for patients with the A/A genotype (n = 67; median OS, 18.4 months) compared with patients with the A/C genotype (n = 50; median Overall survival (OS), 13.9 months) or C/C genotype (n = 10; median OS, 15.6 months) in USA. Sharp et al. 2008) founded (that For A1298C, compared with AA subjects, CC subjects had modest, non-significant, reduced risk (multivariate OR = 0.81 (95 % CI 0.45, 1.49) in UK. These results are in agreement with our results for CC subjects, but disagree for AA subjects.

The present results showed that Comparing all studied cases of colorectal cancer to that of healthy controls regarding their combined genotypes of C677T and A1298C polymorphisms of MTHFR gene, it is noted that cases had a significantly lower frequency of combined (CC and AA) genotypes and (CT and AA) genotypes compared to controls (6.25% vs. 18.9%, p= 0.03 & 11 % vs. 25.6 %, p= 0.02).while they had a significantly higher frequency of combined (TT and AC) genotypes (11% vs. 0 %, p= 0.001). These results were in agreement with our results for the combined genotype 677CT plus 1298AA .

El Awady et al. (2009) founded that The combined genotype MTHFR 1298CC + AC frequencies were 86% with OR 3.71 [95% CI 1.28-10.78] in carcinoma tissues, 69% with OR 1.35 [95% CI 0.57-3.21] in germ line of patients and 62% in controls. The combined genotype 677CT plus any of the following genotypes 1298AA, AC or CC enhanced risk of Colorectal carcinomas (CRC), when comparing germ line DNA polymorphism of patients versus peripheral blood DNA of control subjects with OR 4.5 [95% CI 0.94-21.56], OR 3.12 [95%CI 0.79-12.36] and OR 18 [95%CI 1.56-207.5], respectively, suggesting strong genetic predisposition of certain Egyptian population to Colorectal carcinomas (CRC) in Egypt. These results are in agreement
with our results for (677 CC+1298 AA). Eaton et al. (2005) found that strong inverse associations between combined MTHFR genotypes and MSI (677 CC+1298 AC/CC, OR, 0.09; 95% CI, 0.01-0.59; 677 CT/TT+1298 AA, OR, 0.13; 95% CI, 0.02-0.85) compared with the combined wild-type genotypes (677 CC+1298 AA) in USA.

Regarding normal and abnormal cases of colorectal cancer among tumor marker, hepatic and completely blood count analysis, no higher association between C677T, A1298C polymorphisms and those found in the current study. Possible explanation for these variations in polymorphisms may be due to variations in ethnic background of the cases and controls or due to variation in cases presentations. Conclusions: Our findings indicate that C677T and A1298C polymorphisms of MTHFR gene were found to be associated with development of colorectal cancer among Egyptian patients. Mutant TT genotype of C677T, mutant, AC and wild AA genotypes of A1298C, C and A alleles of A1298C may be considered genetic risk factors for development of colorectal cancer. Maximum risk combined genotypes included (CC and AA), (CT and AA) and (TT and AC).

References

التحولات القاعدية في جين الميثيلين تتراهايدروفوليت ردكتيز كعوامل خطيرة لحدث سرطان القولون والمستقيم بين المرضى المصريين

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سرطان القولون والمستقيم هو مرض يصيب الخلايا الطبيعية حيث تبدأ بطانة القولون أو المستقيم في التغير، ونمو دون حسب ولا رقيب وفي بعض الحالات تكون هذه التغييرات نتيجة لأسباب وراثية. استهدفت هذه الدراسة إثان من التحولات القاعدية المفردة في جين الميثيلين تتراهايدروفوليت ردكتيز (سيتوزين 766 ثايمين) و (ادينين 1221 سيتوزين) وعلاقتهما بمرض سرطان القولون والمستقيم بين مرضى المصريين.

أوضح هذه الدراسة وجود نسبة إحصائية معنوية عالية بالتركيب الجيني (سايمين 767 ثايمين) الخاص بالتحول القاعدي المفرد (سيتوزين 1221 سيمين) كما يوجد نسبة إحصائية معنوية عالية خطيره بالتركيب الجيني (أدنين سيمين) واللايل (سيتوزين) الخاص بالتحول القاعدي المفرد (أدنين 1289 سيمين) والتركيب الجيني (أدنين أدنين) واللايل (أدنين) الخاص بالتحول القاعدي المفرد (سيدزين 1289 سيدزين) كما يوجد أيضا نسبة إحصائية عالية بالتركيبات الجينية المشتركة (سيدزين سيدزين وأدنين أدنين) و (أدنين سيمين سيمين) أو (سيدزين ثايمين) أو (أدنين أدنين) بين حالات مرضى سرطان القولون عنها في الاصحاب.