

Interleukin –1 Receptor Antagonist Gene Polymorphism and Malignant lymphomas

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Abstract:

Background and objectives: The functional polymorphisms regulating immunologic responses may influence the proliferation or suppression of malignant lymphoma. Polymorphism of a specific gene can have an important effect on gene transcription, the stability of the mRNA, or the quantity and activity of the resulting protein. Interleukin –1 (IL-1) gene cluster polymorphisms have been shown to be important mediators of inflammation. This study aimed to determine whether polymorphisms at IL-1 receptor antagonist (IL-1ra) locus modulate the risk of developing malignant lymphoma. **Methods:** the hospital-based case-control study was conducted in Epidemiology and Genetics Unit, Algernon Firth Building, University of Leeds, LS2 9JT. Genotypes were determined for a variable length polymorphism in intron 2 of the IL-1ra gene (IL-1RN) using PCR based technique. Genotypes were examined in a total of 279 lymphoma cases, 60 Hodgkin's disease (HD) cases and 464 non-cancer control subjects. **Results:** The overall allele distribution of these polymorphisms did not differ substantially between patients and controls; the odds ratio (OR) was 0.72 and 95% confidence interval (CI) was 0.5-1.03 for the allele 2 carriers of IL-1RN. Subgroup analysis according to histology [diffuse large B-cell lymphoma (DLBC) and follicular cell lymphoma (FCC)] failed to illustrate differences except for DLBC which showed a significant deficit of the 2/2 genotype in the older DLBC cases, i.e. that the IL-1RN*2/*2 genotype is protective for cases in the 60-65 years group compared to the 1/1 genotype (OR = 0.25 & 95% CI=0.09 – 0.67). On the other hand the IL-1RN*2/*2 genotype was a risk for HD cases (OR=2.27 & 95% CI=1.22-4.24). **Conclusion:** The data of this study show a limited association between IL-1 RN gene polymorphisms and malignant lymphoma risk in total. IL-1RN*2/*2 is associated with increased risk to HD. The possible protection role/risk association of the IL-1RN*2/*2 genotype and DLBC/HD respectively needs further clarification.

Introduction

Maintaining the physiological balance or homeostasis is of vital importance for any living organism. Various forms of endogenous and exogenous stress constantly interfere with this homeostasis. As a result of genetically determined adaptive potential, the effect of these stress factors is normally compensated, and a new level of homeostasis is usually rapidly achieved. The adaptive potential between the

species and between individuals of the same species has a great variation. One of the most important population genetic mechanisms explaining this natural diversity is genetic polymorphism (Steven et al, 2002). Inflammatory stress is the stress form, which is constantly present. Cytokines are a group of small soluble or cell-membrane-bound protein or glycoprotein messenger molecules with high

potential in the regulation of inflammatory responses. The balance of pro-inflammatory and anti-inflammatory cytokines is essential for normal cellular function. Like most human genes, the cytokine genes are also polymorphic (Mwantembe et al, 2001). Some alleles of polymorphic cytokine genes have been shown to be associated with variation in cytokine production capacity. Moreover, allelic imbalance of several cytokine genes has been described in a number of diseases. These complementary biological and pathological associations of cytokine gene polymorphism make them an interesting subject of research (Santtila et al 1998). Clinically oriented genetic association studies are of great value as genetic risk-assessment based treatment strategies will become applicable in clinical practice in the near future (Sehoulie et al, 2002).

Cytokines play important roles in the hematopoietic and immune systems. Their functions include control of cellular and humoral immune responses, with an impact on inflammation, chemotaxis, tumor regression, hematopoiesis, and acute-phase responses (Hsu et al, 1993). Interleukin-1 (IL-1), which is a prototypic multifunctional cytokine synthesized by a variety of cell types, including activated macrophages and stimulated B-lymphocytes, is an essential mediator of inflammation and immunity (Cantagrel et al, 1999). It can exert either inhibitory or promoting effects on neoplasms including hematologic malignancies (Nicklin et al, 1994).

IL-1 gene cluster is situated on chromosome 2q, and is comprised of three related genes within a 430-kilobase region: IL-1A, IL-1B and IL-1RN, which encode the pro-inflammatory cytokines IL-1 α and IL-1 β and their endogenous receptor antagonist

IL-1ra, respectively (Dinarello, 1996). Five alleles of the IL-1ra gene have been reported, corresponding to 2, 3, 4, 5 and 6 copies of an 86-bp variable number of tandem repeat (VNTR) located in intron 2. The frequency of the individual alleles varies among different ethnic or geographic populations, but allele 1 (IL-1RN*1), is always more common than allele 2 (IL-1RN*2). The remaining alleles, occurs in <1% of most populations. The IL-1RN*2 allele enhances IL-1 β production and is associated with several diseases, however data regarding its effects on IL-1ra production are contradictory (Tountas et al, 1999).

Patients and methods

This study was carried in the Epidemiology and Genetics Unit, Algernon Firth Building, University of Leeds, Leeds, LS2 9JT. There were 139 cases of diffuse large B cell lymphoma (DLBC) (aged 25-65 years), 140 cases of follicular center cell lymphoma (FCC) (aged 31-65 years) and 60 cases of Hodgkin's disease (HD) (aged 28-45 years). All the cases were identified from multiple sources throughout the Yorkshire region, in the North of England. Archived samples, comprising paraffin blocks of biopsies, surgical specimens, and peripheral blood smears were obtained. All cases were histologically validated according to the criteria of the Revised European-American classification of lymphoid Neoplasm (REAL) classification (Isaacson, 1995), using the standard panel of markers utilized by the hematological Malignancy Diagnostic Service (HMDS) at Leeds General Infirmary (LGI) (CD20, CD79, CD10, CD5, CD23, CD3, Ki67 and BCL-2). A reference series of 464 healthy population controls (aged 19-66 years) were collected as part of an ongoing lymphoma study being carried out by

the Leukemia Research Fund (LRF) Epidemiology and Genetics Unit. Each control provided a blood sample. Ethical approval to establish a DNA repository for cases of hematological malignancy and to carry out a case-control comparison of genotypes was obtained from Yorkshire MREC.

IL-1RN VNTR (variable number of tandem repeat) Genotyping.

Genomic DNA was extracted from anticoagulated peripheral blood leukocytes using a standard proteinase K digestion and phenol/chloroform extraction method. DNA amplification was done by polymerase chain reaction (PCR) using Peltier thermal cycle (PTC-225). Each PCR reaction was carried out with 50 ng genomic DNA and 1.0 u Ampli Tag Gold polymerase (Perkin-Elmer). For optimal amplification, the Mg^{2+} concentration of the reaction buffer was adjusted to 1.5mM. Primers: IL-1RA F 5` GGT CAG AAG GGC AGA GA3`. IL-1RA R 5` CCC CTC AGC AAC ACT CC 3`. Negative controls without DNA template were included with each reaction. PCR denaturation 5 min at 94°C, (94°C 1-min, 60°C 1-min, 72°C 1-min) 30 cycles, extension 5-min at 72°C. The PCR products sizing were analyzed by electrophoresis on a 2% agarose gel stained with ethidium bromide (Kenemoto et al, 2000). The products were sized relative to a 100 bp DNA ladder and were coded as follows: allele 1= 4 repeats (442bp), allele 2= 2 repeats (270 bp), allele 3= 5 repeats (528bp), allele 4= 3 repeats (356 bp), allele 5 = 6 repeats (614bp); due to the rarity the 3, 4 and 5 alleles were grouped for statistical analysis.

Statistical Analysis

All odds ratios (OR) and 95% confidence intervals (CI) associated with risk of lymphoma were adjusted for sex as well as age as a continuous

variable, instead of age –sex matching using an unconditional logistic regression model. 95%CI value <1 is protective and >1 is risky. Accordance with the Hardy-Weinberg equilibrium the genotype frequencies among cases and controls were also compared with the X^2 test. All analyses were conducted using STATA version 7 software.

Results

In total there were 464 controls with an age range of 19-66 years, 51.7% of these were males. There were 139 cases of DLBC, with an age range of 25-65 years, and 52.2% of these were males. There were 140 cases of FCC, with an age range of 31-65 years, and 45.6% of these were males.

Age and sex comparison of controls:

When the controls were stratified into 3 age groups, (19-49, 50-59, 60-65 years), no significant differences in genotype frequency in the control population were seen, P=0.52.

When the controls were stratified by sex, no significant differences in genotype frequency in the control population were seen, P=0.99.

Age and sex comparison of cases:

When all cases were stratified by sex, no significant differences in genotype frequency in the case populations were seen, P=0.66, for DLBC only p=0.166, and for FCC only p=0.91.

When the cases were stratified into 3 age groups, (19-49, 50-59, 60-65) (the age groups basically being determined on splitting the data into 3 reasonably even sections), no significant differences in genotype frequency in the case population (DLBC and FCC) were seen, P=0.48. While no significant differences were seen for age in either the DLBC or FCC groups (p=0.19 and p=0.06 respectively), but the IL-1RN2*/*2 genotype was different for older DLBC cases (60-65 years) (table 1) & (Figure 1).

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Table 1: Genotype frequencies for IL-1RN genotypes in combined lymphoma cases, DLBC and FCC cases, stratified by age.

DLBC & FCC (n=279)					
Genotype	19-49y %	50-59y%	60-65y%	OR	95%CI
1/1	26 (49.03)	74 (52.48)	32 (47.06)	1.0	-
1/2	11 (20.75)	40 (28.37)	18 (26.47)	1.29	0.59 – 2.80
1/345	1 (1.89)	4 (2.84)	1 (1.47)	1.22	0.13 – 10.95
2/2	15 (28.3)	23 (16.31)	17 (25.0)	0.65	0.31 – 1.36
DLBC (n=139)					
Genotype	19-49y	50-59y	60-65y	OR	95%CI
1/1	10 (34.48)	38 (51.35)	19 (59.38)	1.0	-
1/2	5 (17.24)	23 (31.08)	5 (15.63)	0.98	0.30 – 3.14
1/345	1 (3.45)	1 (1.35)	1 (3.13)	0.35	0.03 – 4.24
2/2	13 (44.83)	12 (16.22)	7 (21.88)	0.25	0.09 – 0.67*
FCC (n=140)					
Genotype	19-49y	50-59y	60-65y	OR	95%CI
1/1	16 (66.67)	36 (53.73)	13 (36.11)	1.0	-
1/2	6 (25)	17 (25.37)	13 (36.11)	1.63	0.57 – 4.63
1/345	0 (0)	3 (4.48)	0 (0)	0	-
2/2	2 (8.33)	11 (16.42)	10 (27.78)	3.42	0.72 – 16.25

This table shows that there is a significant deficit of the IL-1RN*2/*2 genotype in the older DLBC cases in the 60-65 years group compared to the IL-1*1/*1 genotype.

Table 2: IL-1 RN genotypes in combined lymphoma (DLBC and FCC) cases.

Genotype	Controls (%) (n = 464)	Cases (%) (n = 279)	OR	95% CI
1/1	218 (46.98)	132 (50.38)	1.0	-
1/2	157 (33.84)	69 (26.34)	0.72	0.5 – 1.03
1/345	8 (1.72)	6 (2.29)	1.23	0.42 – 3.64
2/2	81 (17.46)	55 (20.99)	1.12	0.74 – 1.68

There appears to be non-significant protective effect for the IL-RN*1/*2 genotype.

Table 3: IL-1 RN genotypes in DLBC cases (n=139).

Genotype	Controls (%)	Cases (%)	OR	95% CI
1/1	218 (46.98)	67 (49.63)	1.0	-
1/2	157 (33.84)	33 (24.44)	0.68	0.2 – 1.08
1/345	8 (1.72)	3 (2.22)	1.22	0.31 – 4.72
2/2	81 (17.46)	32 (23.70)	1.28	0.78 – 2.10

There appears to be non-significant protective effect for the IL-1RN*1/*2 genotype.

Table 4: IL-1 RN genotypes in FCC cases (n=140).

Genotype	Controls (%)	Cases (%)	OR	95% CI
1/1	218 (46.98)	65 (51.18)	1.0	-
1/2	157 (33.84)	36 (28.35)	0.76	0.48 – 1.21
1/345	8 (1.72)	3 (2.36)	1.25	0.32 – 4.87
2/2	81 (17.46)	23 (18.11)	0.95	0.55 – 1.63

There appears to be non-significant protective effect for the IL-1RN*1/*2 genotype.

Table 5: IL-1 RN genotypes in HD cases (n=60).

Genotype	Controls (%)	Cases (%)	OR	95% CI
1/1	218 (46.98)	26 (43.3)	1.0	-
1/2	157 (33.84)	11 (18.3)	0.58	0.28 – 1.22
1/345	8 (1.72)	1 (1.7)	1.04	0.12 – 8.71
2/2	81 (17.46)	22 (36.7)	2.27	1.22 – 4.24*

This table shows that IL-1RN*2/*2 genotype % is increased in HD cases.

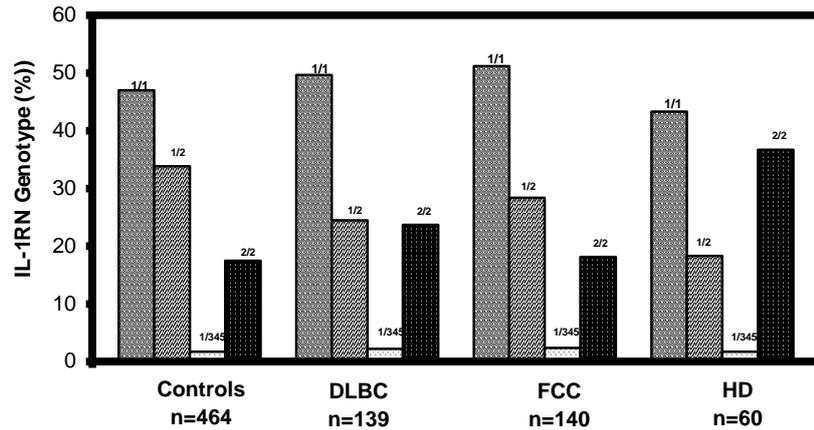


Figure 1: Percentage of IL-1RN genotypes among all studied groups.

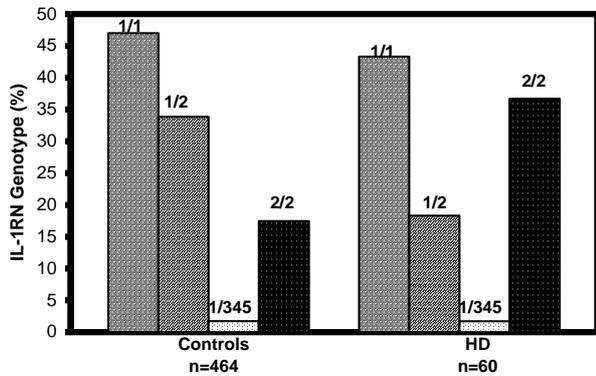


Figure 2: Percentage of IL-1RN genotypes among controls and HD groups.

and
HD

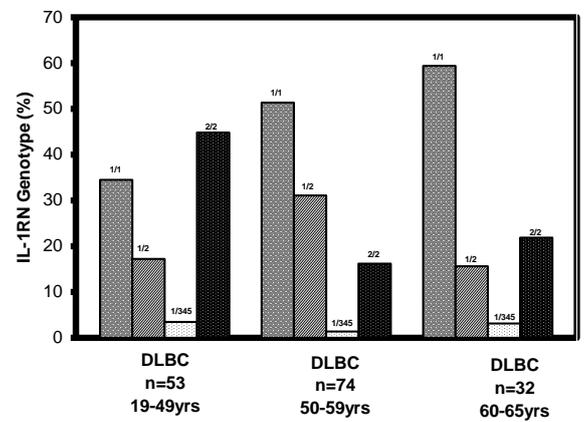


Figure 3: Percentage of IL-1RN genotypes stratified by age among DLBC cases.

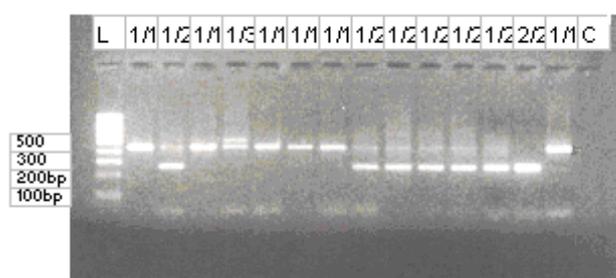


Figure 4: PCR/agarose gel electrophoresis of the IL-1RN VNTR polymorphism illustrating the most common genotypes in this study (L=100 base pair ladder molecular weight marker & C=control).

Discussion

Interleukin (IL)-1 α and IL-1 β are major proinflammatory cytokines that are synthesized during infection and

inflammatory processes. IL-1RA competes for the same IL-1 receptor as that for IL-1 α and IL-1 β , thus

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modulating the potentially injurious effects of IL-1 (Arend et al, 1998). The gene for IL-1ra is located on the long arm of the chromosome 2 on a 430 kb stretch of DNA. Intron 2 of the IL-1ra gene contains a variable number of identical tandem repeats (VNTR) of an 86 base pair length of DNA (Nicklin et al, 1994). IL-1RN*2 has been associated with decreased IL-1ra levels, and consequently increased IL-1 β levels (Andus et al, 1997). The central role of the IL-1 system is protection against many different insults, ranging from microbial colonization to infection to malignant transformation. The relative levels of IL-1-RA and IL-1 at an inflammatory site will thus determine whether a proinflammatory response will be initiated and persist or will be terminated. Biological functions of IL-1 on hematologic malignancies have therefore already pointed out (Dinarelo, 1996). IL-1B has anti-neoplastic activity through activation of the immune system. The higher incidence of hematologic malignancies among immune deficiency individuals suggests that the differences in the status of immune activation may change the predisposition to hematologic malignancy (Santavenere et al 1994).

The data of this study did not find any difference in genotype frequencies of IL-1RN between malignant lymphoma patients as a whole and control subjects. On the other hand this study confirmed a mild over expression of IL-1RN2*/2* in malignant lymphoma cases (DLBC & FCC), which was not statistically significant (controls 17.46%, DLBC 23.7% and FCC 18.11%). The allelic frequency of IL-1RN2*/2* was lower in this study than found in the study of Rollinson et al (2003) (controls 8% and gastric lymphoma 33.9%). Excess in risk of the malignant lymphoma have been desc-

ribed for patients with various conditions involving substantial immune dysfunction, particularly conditions where chronic antigenic stimulation is present (Machado et al, 2001). The consideration of polymorphisms involved in the immune response, in combination with those involved with the prevention of DNA damage, and further those involved in DNA repair, could allow the mechanisms underlying these associations to be explored further.

Subgroup analyses according to histologic subtypes showed a possible difference in DLBC. When DLBC cases were stratified into 3 age groups, the IL-1RN2*/2* genotype in the older cases (60-65 year group) showed a protective effect when compared to IL-1 1*/1* genotype (OR=0.25 and 95% CI=0.09 – 0.67). Steven et al (2002) reported that it might be expected that people who were IL-1RN*2 homozygous (IL-1RN*2*/2*) might have a genetic advantage in cancer prevention because this genotype is associated with low level of IL-1ra and elevated level of IL-1 β (which has anti-neoplastic activity) with prolonged and more severe inflammatory reactions. These results with malignant lymphoma was consistent with the study done by Matsuo et al (2001), who suggested that, the effects of IL-1 family gene polymorphisms in lymphoid malignancies seem to be limited. On the other hand Rollinson et al (2003) reported a significant association between gastric marginal zone lymphoma and inheritance of IL-1RN2*/2* genotype. Also El-Omar et al (2000) reported that IL-1RN*2 homozygous carriers were at increased risk for developing gastric carcinoma. They explained their results by confirming that both IL-1B and IL-1RN polymorphisms enhance IL-1B expression, which are associated with an increased risk of gastric carcinoma. Sehoulie et al

(2002) reported elevated level of IL-1 β at the homozygous for IL-1RN*2 which was associated with gastric cancer. Demeter et al (1996) found in a small group study of patients with secondary AML that the frequency of IL-1RN*4 allele appeared to be significantly increased.

The data of this study revealed that the IL-1RN2*/2* genotype was in 36.7% of HD cases compared to 17.46% in the control reference population, showing an association with increased risk of HD (OR=2.27 & 95% CI= 1.22-4.24). It is postulated that the association between IL-1RN*2 allele with HD was due to impaired production of IL-1ra and the high IL-1 β levels associated with IL-1RN*2 allele (Tountas et al, 1999). IL-1 β favour a proinflammatory response with substantial immune dysfunction, which may therefore increase risk of DNA damage and lymphomagenesis. Although polymorphism in IL-1RA gene was associated with various malignant diseases, data is lacking for HD.

In conclusion, the present study revealed that genotype frequencies of IL-1RN gene polymorphism do not vary between malignant lymphoma patients and non-cancer controls. This is probably due to the genetic background of the patients studied. IL-1RN*2/*2 is associated with increased risk to HD. The biological mechanism of this difference is unclear. The apparent relationship between IL-1RN*2/*2 and both lymphoma and HD raises the possibility that another nearby gene polymorphism might be responsible for some of these associations. The results of this study provide useful information to explore the susceptibility to lymphoma and HD, which it would be valuable to investigate further in larger studies among Egyptians.

Acknowledgements

I thank Prof. Gareth J Morgan for his advise on genetic issues, Mrs Heather Kesby for technical assistance and Dr. Sara Rollinson for technical assistance and statistical analysis. This work was funded by the Leukaemia Research Fund of the Molecular Epidemiology and Genetic Unit, Algernon Firth Building Leeds, LS2 9JT, Leeds University.

References

1. **Andus T, Daig R, Vogl D, Aschenbrenner E, Lock G and Hollerbach S (1997).** Imbalance of the interleukin-1 system in colonic mucosa associated with intestinal inflammation and interleukin-1 receptor antagonist genotype 2. *Gut*; 41: 651-657.
2. **Arend WP, Malyak M, Guthridge CJ and Gaby C (1998):** Interleukin-1 receptor antagonist: role in biology. *Annu Rev Immunol*; 16: 27-55.
3. **Cantagrel A, Navaux F, Lou bet – les cou lie P, Nouzhashemi F, Ernault G, Abbal M, Constantin A, Laroche M and Mazieres B (1999).** Interleukin-1 β , interleukin-1 receptor antagonist, interleukin-4 and interleukin-10 gene polymorphisms. *Arthritis&Rheumatism*; 42: 1093-1100.
4. **Demeter J, Messer G, Ramisch S, Mee JB, Di Giovine FS, Schmid M, Herrmann F and Parzolt F. (1996).** Polymorphism within the second intron of the IL-1 receptor antagonist gene in patients with hematologic malignancies. *Cytokines Mol Ther*; 2: 239-242.
5. **Dinarelo CA (1996).** Biologic bases for interleukin-1 in disease. *Blood*; 87: 2095-2147.
6. **El-Omar EM, Carrington M, Chow WH, Mc Coll KE, Bream JH and Young HA (2000).** Interleukin-1 polymorphisms associated with increased risk of gastric cancer. *Nature*; 404: 398-402.
7. **Hsu SM, Waldron JW, Hsu PL and Hough AJ (1993).** Cytokines in malignant lymphomas: review and

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- prospective evaluation. *Hum Pathol*; 24: 1040-57.
8. **Isaacson PG (1995)**. The revised European–American lymphoma(REAL) classification. *Clin Oncol*; 7: 347-348.
 9. **Kanemoto k, Kawasaki J, Miyamoto T, Obayashi H and Nishimura M (2000)**. Interleukin (IL)-1 β , IL-1 α and IL-1 Receptor Antagonist Gene Polymorphisms in patients with temporal lobe epilepsy. *Ann Neurol*; 47: 571-574.
 10. **Machado JC, Pharoah P, Sausa S, Carvalho R, Oliveira C, Figueiredo C, Amorin A, Seruca R, Caldas C, Carneiro F and sobrinho Sinoes M (2001)**. Interleukin-1 β and interleukin-1RN polymorphisms are associated with increased risk of gastric carcinoma. *Gastroenterology*; 121: 823-829
 11. **Matsuo K, Hamajima N, Suzuki R, Nakamura S, Seto M, Morishima Y and Tajima K (2001)**. No substantial difference in genotype frequencies of interleukin and myeloperoxidase polymorphisms between malignant lymphoma patients and non-cancer controls. *Haematologica*; 86: 602-608.
 12. **Mwantembe O, Gaillard MC and Barkhuizen M (2001)**. Ethnic differences in allelic associations of the interleukin-1 gene cluster in South African patients with inflammatory bowel disease and in control individuals. *Immunol genetics*; 52: 249-54
 13. **Nicklin MJ, Weith A and Duff GW (1994)**. A physical map of the region encompassing the human interleukin-1 alpha, interleukin-1 beta and interleukin-1 receptor antagonist genes. *Genomics*; 19: 382-4.
 14. **Rollinson S, Levene AP, Meusah FK, Roddam PL, Allan JM, Diss TC, Roman E, Jack A, Maclennan K, Dixon MF, Morgan GJ (2003)**. Gastric marginal zone lymphoma is associated with polymorphisms in genes involved in inflammatory response and antioxidative capacity. *Blood*; 102 1007-11.
 15. **Santavenere E, Di Pietro R, Centurione MA, Trubiani O, Zamia L and Roma R (1994)**. IL-1 α antiproliferative and differentiative effects on Daudi Lymphoma cells: multiparametric analysis. *Cell Biol Int*; 18: 777-82.
 16. **Santtila S, Savinainen K and Hurme M (1998)**. Presence of the IL-1RA allele 2 (IL-1RN*2) is associated with enhanced IL-1 production in vitro. *Scand J Immunol*; 47: 195-8.
 17. **Sehoulie J, Mustea A, Kongsen D, Katsares I, Lichtenegger w (2002)**. Polymorphism of IL-1 receptor antagonist gene: role in cancer. *Anticancer Res*; 22: 3421-4.
 18. **Steven S, Gerber S and Ledger WJ (2002)**. Influence of interleukin-1 receptor antagonist gene polymorphism on disease. *Clin Inf Dis*; 34: 204-9.
 19. **Tountas NA, Casini-Raggi V, Yang H, Di Giovine FS, Vecchi M and Kam L (1999)**. Functional and ethnic association of allele 2 of the interleukin-1 receptor antagonist gene in ulcerative colitis. *Gastroenterology*; 117: 806-813.

تعدد شكل مورث مضاد مستقبل إنترلوكين-1 و الأورام الليمفاوية السرطانية منى حلمى الرئيس قسم الباثولوجيا الأكلينيكية و الكيمياء بكلية طب البنات-جامعة الأزهر

لقد وجد أن تعدد الشكل الوظيفي المنظم للإستجابة المناعية قد يتدخل فى تكاثر أو إخماد الأورام السرطانية الليمفاوية. كما أن تعدد شكل مورث معين قد يكون له تأثير هام على نسخ المورث أو إستقرار الحامض النووى المرسل ر ن أ، أو فى كمية و نشاط البروتين الناتج. فقد أظهر تعدد شكل سباطة مورث إنترلوكين-1 أهمية كوسيط فى الإلتهابات.

هدف البحث:

هدف البحث هو تحديد ما إذا كان تعدد أشكال موقع مضاد مستقبل إنترلوكين-1 يعدّ ل إنماء الأورام السرطانية الليمفاوية أو يحمى منها؟

مادة البحث و طريقة:

لقد أجريت الدراسة فى وحدة المورثات و الوبائيات فى جامعة ليدز بإنجلترا. وقد تم تحديد نوع المورث لاختلاف طول و تعدد شكل سباطة مورث مضاد مستقبل إنترلوكين-1 فى إبترون 2 بإستخدام تقنية تفاعل سلسلة البلمرة. كما تم إختيار نوع المورث فى إجمالى 279 حالة للأورام الليمفاوية السرطانية و التى إشملت على مجموعتين فرعيتين: 139 حالة للأورام الليمفاوية المنتشرة للخلايا الكبيرة و 140 حالة للأورام الليمفاوية للخلايا المتوصلة و ذلك بالإضافة إلى 60 حالة لمرض هودجكينز و كذلك 464 شخص من الأصحاء كمجموعة ضابطة.

نتائج البحث:

أظهر البحث أنه لا يوجد على وجة العموم اختلافا جوهريا فى توزيع الملائم بالنسبة لتعدد أشكال سباطة مورث مضاد مستقبل إنترلوكين-1 بين المرضى و المجموعة الضابطة. كذلك فشل التحليل للمجاميع الفرعية طبقا للتنوع النسيجي فى إيضاح اختلافات فيما عدا مجموعة الأورام الليمفاوية المنتشرة للخلايا الكبيرة التى أظهرت عجز مهم فى نوع المورث 2\2 فى المجموعة العمرية الأكبر سنا فى هذه المجموعة الفرعية. بينما و على الجانب الآخر كان هناك زيادة مهمة فى نوع المورث 2\2 فى حالات مرض هودجكينز.

الإستنتاج و التوصيات

أظهرت بيانات هذه الدراسة مصاحبة محدودة بين تعدد شكل مورث مضاد مستقبل إنترلوكين-1 و خطر الأورام الليمفاوية السرطانية ككل. كذلك أظهرت الدراسة خطر مصاحبة نوع المورث 2\2 لمضاد مستقبل إنترلوكين-1 مع مرض هودجكينز. و يوصى بدراسة خطر مصاحبة \ أو حماية نوع المورث 2\2 لمضاد مستقبل إنترلوكين-1 مع الأورام الليمفاوية المنتشرة للخلايا الكبيرة \ و مرض هودجكينز فى المرضى المصريين.