

Study of Erythropoietin on IGM serum levels in HCV positive patients on regular HD

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Abstract

Both uremia and HD process cause immunosuppression in HD patients. There was significant increase of total serum IgG and IgM levels found in patients with chronic HCV compared with healthy controls. There is evidence pointing to direct effect of rHuEPO upon B cells. High doses of recombinant human erythropoietin (rHu EPO) enhanced in vitro Ig production and proliferation of various plasma cell lines, as well as human plasma cells generated in vitro. Study was conducted at hemodialysis Unit of Shubra Municipal hospital between August 2010 to February 2011. 30 HCV positive patients on regular hemodialysis were included in study, using bicarbonate dialysate and polysulfone membrane dialyser, for 4 hours 3 times weekly. Patients were divided into 2 groups: first group: 15 patients on EPO therapy. 4000 IU/week and second group not taking EPO for all patients full clinical examination was done, CBC, BUN, serum creatinine, ALT, AST, serum albumin and serum IgM by ELISA (quantitative assay), were done.

There was no significant difference between 2 groups as regards age, sex distribution, WBC count, ALT, AST, serum creatinine, BUN and IgM serum level. First group had borderline significant higher Hgb and Hct than second group ($p = 0.056$). Females didn't have higher serum IgM level than males ($p = 0.403$). All correlations of IgM serum level to other parameters of study were irrelevant. Uremia seems to protect ESRD patients on regular HD from complications of HCV and also EPO effect on Ig serum levels.

Key words : Erythropoietin- IgM –HCV – Hemodialysis.

Introduction

Uremia is associated with a state of immune dysfunction characterized by immunodepression that leads to high prevalence of infections as well as by immune activation resulting in inflammation (*Kiechl et al., 2002*).

Improper immunological parameters of both humoral and cellular immunity in CKD patients seem to be deepened by hemodialysis (HD) process (*Liwosca et al., 2011b*). Patients with renal disease have been at increased risk of acquiring HCV because of prolonged vascular access as well as the potential for exposure to infected patients and contaminated equipment (*Fabrizi et al., 2007*).

Several studies have provided experimental evidence of disorders of both cellular and humoral immunity in chronic hepatitis C patients (*Lotfy et al., 2006*).

HCV infection is strongly associated with mixed cryoglobulinemia (MC), a benign disorder characterized by the proliferation of B lymphocytes producing polyclonal IgG or monoclonal IgM with rheumatoid factor (RF) activity that characteristically may precipitate at low temperatures (*Fazi et al., 2010*).

Besides B-cell activation (non-antigen-specific and antigen-specific), HCV seems to infect B lymphocytes directly (*Bokle and Sepp, 2010*).

Correction of anemia and maintenance of stable hemoglobin levels using erythropoiesis stimulating agents (ESA) is an important aspect of ESRD management (*Kalantar-Zadeh and Aronoff, 2009*).

Epo therapy leads to improved humoral immune response, either directly or via T-cells help (*Prutchi-Sagiv et al., 2005*).

Epo treatment was associated with enhanced lymphocyte activity of both T- and B-cells (*Lifshitz et al., 2010*).

Erythropoietin-receptor (EPO-R) presence on all populations of immune cells implies that EPO/rhEPO can influence lymphocytes, monocyte and granulocytes directly and somehow modulate their immunological responses (*Liwoska et al., 2011 a*).

High doses of rHu EPO enhanced in vitro immunoglobulin production of various plasma cell lines, as well as human plasma cell generated in vitro (*Prutchi-Sagiv et al., 2005*).

The uremic patient on regular hemodialysis (HD) is subjected to a wide range of immune modulators including the uremic state per se, multiple transfusions and exposure to bio incompatible materials and endotoxins. Erythropoietin (EPO) therapy may raise concern about its potential influence on this complex Scenario (*William et al., 1998*).

Aim of work

Is to determine the effect of erythropoietin on IgM level in ESRD patients infected with HCV on regular hemodialysis as IgM is one of the markers of cryoglobulinemia.

Patients and methods

This study was conducted at hemodialysis unit of Shubra Munieipal Hospital between august 2010 to February 2011. it was conducted on 30 ESRD hepatitis C positive patients on regular hemodialysis with bicarbonate dialysate and polysulfone membrane dialyser, three times per week. All patients had chronic hepatitis C infection for less than 10 years with liver enzymes less than two fold increase above normal (specially ALT) and last blood transfusion more than 30 days ago.

These patients were divided into 2 groups

First group: Includes 15 ESRD hepatitis C positive patients on regular HD and on erythropoietin therapy. Patients of this group were administered erythropoietin dose of 4000 IU/week.

Second group: Includes 15 ESRD hepatitis C positive patients on regular HD and not on erythropoietin therapy.

We excluded from the study patients with history of DM, autoimmune and allergic diseases.

Patients with hepatitis βvirus confection, dialysis vascular access infection, history of paraproteinemia, systemic vasculitis, acute hepatitis liver cell failure or chronic infections other than HCV and chronic inflammatory diseases were excluded from the study.

All patients were subjected to full history and complete physical examination, complete blood count, blood urea nitrogen, serum creatinine, liver enzymes (AST and ALT), serum albumin, and serum IgM by ELISA (quantitative assay).

Methods

1- Creatinine

This assay is a kinetic method (*Yatzidis, 1974*).

Assay principle

Creatinine in alkaline solutions react with picrate to form a colored complex. The rate of complex formation is measured photometrically at 492 nm.

Calculations

A2-A2: A(specimen)-A (standard)

* Concentration of creatinine in serum or

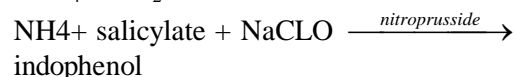
$$\text{plasma (mg/dL)} = \frac{A_{\text{Specimen}}}{A_{\text{standard}}} \times 2$$

2- Urea

This procedure is enzymatic-spectrophotometric (*Tabacco et al., 1979*).

Assay principle

Urea in the sample originates by means of the coupled reactions described below, a colored complex that can be measured by spectrophotometry : urea + H₂O $\xrightarrow{\text{Urea}}$ 2NH₄ + CO₂



Calculations

The urea calculation in the sample is calculated using the following general formula:

$$\text{Urea in sample} = \frac{A_{\text{Sample}}}{A_{\text{Standard}}} \times \text{Standard}$$

X sample dilution factor
Where c= concentration

3- Albumin (BCG):

This assay is colorimetric method (Doumas et al. , 1971).

Assay principle

In a buffered solution bromo-cresol green forms with albumin, a green colour complex whose intensity is proportional to the amount of albumin present in the specimen calculations:

$$\text{Albumin Concentration (g/dL)} = \frac{A_{\text{Specimen}}}{A_{\text{Standard}}} \times 4$$

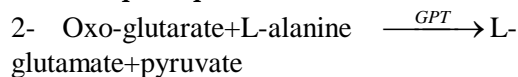
4- ALT (SGPT)

Liqui-UV test (*Schumann and Klauke, 2003*).

Assay principle

Kinetic method for the determination of ALT activity according to the recommendations of the expert panel of the IFCC (International federation of clinical chemistry) without pyridoxal-phosphate activation.

Reaction principle:



5- AST (SGOT)

Liqui-UV test (*Schumann and Klauke, 2003*).

Assay principle

Kinetic method for the determination of AST activity according to the recommendation of the expert panel of the IFCC (international federation of clinical chemistry) without pyridoxal-phosphate activation reaction principle: 2-Oxo-glutarate+L-aspartate $\xrightarrow{\text{GOT}}$ L-glutamate=oxaloacetate.



6- Serum IgM by Elisa (Diagnostic Automatic Inc., 2009).

Intended use: to quantitate total human immunoglobulin M (IgM).

Principle of procedure

Solid phase capture sandwich ELISA assay using a Microwell format.

Patients and standard dilutions dilute each serum or plasma specimen to be tested initially 1: 1000 in phosphate buffered saline (PBS) e.g 10 µl of specimen into 990 ml of PBS, then subdilute 1:10 with the IgM specimen diluent provided for a final dilution of 1:10.000. Prepare serial two fold dilutions of the human IgM standard: neat, 1:2, 1:4, 1:8 etc., with the specimen diluent provided, use the specimen diluent alone as the blank control well.

Assay procedure

Allow each reagent to reach room temperature before use

- 1- Add 100 µL of diluted specimen or standard to each Microwell.
- 2- Incubate at room temperature for 60 minutes.
- 3- Decant and wash each Microwell four times with wash buffer (dilute buffer 1:15 with reagent grade water).
- 4- Add 100 µL of HRP conjugated goat anti-human IgM to each well.
- 5- Incubate at room temperature for 60 minutes.
- 6- Decant and wash as in step 3.
- 7- Add 100 µL of TMB/peroxidase substrate and incubate at room temperature for 30 minutes.
- 8- Terminate the reaction with 100 µL of 0.5 N sulfuric acid.
- 9- Zero the MICrowell reader at 450 nm using the specimen diluent zero control well.
- 10- Determine the optical density (O.D.) of the remaining wells.
- 11- Construct a standard curve using the O.D. values obtained for each of the standards.

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12- Interpolate the unknowns from the standard curve.
Dynamic range = 0.031 µg/ml. 2.0 µg/ml

* P value > 0.1 is considered non-significant

Statistical analysis

Statistical presentation and analysis of the present study was conducted, using the mean, standard error, student t-test, chi-square and linear correlation coefficient by SPSS V17. We also used Analysis of variance (ANOVA) test to compare different items in the same group in quantitative data.

P-value ≤ 0.05 is considered significant

* P value = 0.05 to < 0.1 is considered borderline significance

* P value ≤ 0.01 is considered highly significant

Results

On comparing first and second group as regards age, there was no statistically significant difference between 1st group (mean±SD = 50.533±8.766 years) and 2nd group (mean±SD = 51.133± 6.632 years) using unpaired student t-test (p-value = 0.834).

We didn't find a statistically significant difference between 1st and 2nd group as regards sex distribution, (p-value = 0.140) using chi-square test, while females constituted 40% of 1st group and 66.67% of 2nd group and the total number of females included in the study constituted 53.33% of all participants in the study. Males constituted 60% of 1st group and 33.33% of 2nd group with a total of 46.67% of all participants in the study

Table (1): Comparison of first group and second group as regards serum creatinine

	S. creatinine (mg/dL)		T-test*	
	Range	Mean±SD	t	p-value
First group	6.100-12.500	10220±2.066	0.290	0.774
Second group	7.000-15.900	9.993±2.207		

* Unpaired student t-test

Table (2): Comparison of first group and second group as regards blood urea nitrogen

	Blood urea nitrogen (mg/dL)		T-test*	
	Range	Mean±SD	t	p-value
First group	112.000-184.000	148.400±21.596	0.164	0.871
Second group	107.000-200.000	146.800±31.122		

* Unpaired student t-test

Table (3): Comparison of first group and second group as regards AST level in serum

	AST Iu/L		T-test*	
	Range	Mean±SD	t	p-value
First group	8.000-22.000	13.600±3.851	-2.282	0.030
Second group	10.000-21.000	16.467±2.973		

* Unpaired student t-test

Table (4): Comparison of first group and second group as regards ALT level in serum

	ALT Iu/L		T-test*	
	Range	Mean±SD	t	p-value
First group	5.000-15.000	8.533±2.800	-1.707	0.099
Second group	6.000-15.000	10.200±2.541		

* Unpaired student t-test

Table (5): Comparison of first group and second group as regards serum albumin

	S. albumin (g/L)		T-test*	
	Range	Mean±SD	t	p-value
First group	3.100-4.200	3.573±0.371	-0.141	0.889
Second group	3.000-4.300	3.593±0.404		

* Unpaired student t-test

Table (6): Comparison of first group and second group as regards hemoglobin (Hgb) level

	Hgb (g/dL)		T-test*	
	Range	Mean±SD	t	p-value
First group	7.800-14.500	10.113±2.144	1.992	0.056
Second group	5.700-12.400	8.600±2.015		

* Unpaired student t-test

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Table (7): Comparison of first group and second group as regards hematocrit (Hct) level

	Hct (%)		T-test*	
	Range	Mean±SD	t	p-value
First group	20.800-41.700	31.080±6.487	1.990	0.056
Second group	17.300-37.000	26.687±5.568		

* Unpaired student t-test

Table (8): Comparison of first group and second group as regards white blood cells

	WBC (x 10 ⁹ /L)		T-test*	
	Range	Mean±SD	t	p-value
First group	3000.000-87.00	5373.333±1810.472	0.430	0.670
Second group	2100.000-8000.00	5073.333±2005.516		

* Unpaired student t-test

Table (9): Comparison of first group and second group as regards serum IgM level

	IgM (ug/mL)		T-test*	
	Range	Mean±SD	t	p-value
First group	40.300-213.700	119.467±61.781	0.238	0.814
Second group	43.000-214.500	114.647±48.536		

* Unpaired student t-test

Table (10): Comparison of IgM level in serum in males and females in both first and second groups together

	IgM (ug/mL)		T-test*	
	Range	Mean±SD	t	p-value
Female	40.300-213.700	125.025±52.345	0.850	0.403
Male	43.000-214.500	107.950±57.722		

* Unpaired student t-test

Table (11): Correlation of serum level of IgM and different parameters of the study in first group

First group	IgM	
	R*	P-value
Age	-0.119	0.674
S. creatinine	-0.374	0.170
BUN	-0.470	0.077
SGOT (AST)	-0.467	0.079
SGPT	-0.247	0.375
S. albumin	0.035	0.901
EPO dose	0.242	0.384
Hgb	0.005	0.987
Hct	0.122	0.665
WBC	-0.104	0.712

* Linear correlation coefficient (r)

Table (12): Correlation of serum level of IgM and different parameters of the study in second group

Second group	IgM	
	R*	P-value
Age	-0.147	0.600
S. creatinine	-0.064	0.821
BUN	0.300	0.277
SGOT (AST)	0.338	0.218
SGPT (ALT)	0.467	0.079
S. albumin	-0.085	0.762
Hgb	-0.305	0.270
Hct	-0.223	0.425
WBC	0.172	0.540

* Linear correlation coefficient (r)

Table (13): Correlation of serum level of IgM and different parameters of the study in both first and second group

1 st & 2 nd group	IgM	
	R*	P-value
Age	-0.131	0.491
S. creatinine	-0.227	0.229
BUN	-0.057	0.766
SGOT (AST)	-0.167	0.377
SGPT (ALT)	0.034	0.858
S. albumin	-0.021	0.912
EPO dose	0.242	0.384
Hgb	-0.102	0.591
Hct	0.000	1.000
WBC	0.028	0.885

* Linear correlation coefficient (r)

Discussion

The depression of the immune response in the uremic patient is global and concerns both humoral and cellular sectors (*Foley and Collins, 2007*).

Disorders of both innate and adaptive immune systems and functional abnormalities of monocytes, neutrophils and dendritic cells, are directly linked with infection risk in this patient population (*Lim et al., 2007*).

Death from sepsis is 50 times higher in hemodialysis patients than in the general population even after accounting for other comorbidities. One of the most difficult causes to treat is the development of an acquired immune dysfunction associated with chronic kidney disease (CKD) and dialysis therapy (*Geara et al., 2010*).

Hepatitis C virus (HCV) is commonly associated with autoimmune disease as extra-hepatic manifestations (EHM).

The most important auto-immune diseases associated with HCV are mixed essential cryoglobulinemia (MEC) and Sjogren syndrome (SS) (*Awad et al., 2011*). Increasing evidence suggests that HCV can interfere with innate immune activation at multiple levels (*Jang and Chung, 2010*).

HCV itself seems to be able to stimulate B cells through different pathways and mechanisms (*Bokle and Sepp, 2010*).

The persistent of stimulation of B cells by viral antigen could be responsible for leading to polyclonal and later to monoclonal expansion of B cells (*Ito et al., 2011*).

The highest level of B-lymphocyte stimulator have been found in chronic HCV-infected subjects with clinical and laboratory features of autoimmunity (*Bokle and Sepp, 2010*).

EPO structure presents elements of cytokines composition and that is why, it is considered that this hormone, a part from its influence on red blood cells system, can regulate immunological responses (*Liwoska et al., 2011b*).

Studies over the last 12 years demonstrated that erythropoietin is probably able to modulate or amplify some signaling pathways important for human lymphocytes and monocyte

functions. There are also many studies demonstrating the role of rHu EPO in improving immune responses in CRF patients and at the same time suggesting that rHu EPO may act as an immunomodulating cytokine in the human organism (*Liwoska et al., 2011a*).

In our study, there was no statistical significant difference as regards age ($P = 0.834$) and sex distribution ($p = 0.140$) between the first group with EPO therapy and the second group without EPO therapy. Serum immunoglobulin concentrations tend to increase with age (*Gonzalez et al., 2008*).

On comparing first and second groups, there was no statistical significant difference as regards serum creatinine levels ($p = 0.774$) and blood urea nitrogen levels ($p = 0.871$), which means that HCV infection together with concomitant EPO therapy didn't influence these two parameters in ESRD patients on regular HD.

In our study, second group showed higher serum AST levels than first group on EPO therapy ($p = 0.03$). Also, second group showed a borderline significantly higher serum ALT levels than first group ($p = 0.099$). In our study, EPO seems to have an anti-inflammatory response influencing our markers of hepatic inflammation or may be it may have a liver supporting effect. Further studies are needed to elucidate this role using liver biopsy findings.

Compared to non-uremic HCV patients, ESRD patients with chronic hepatitis C have milder hepatic necroinflammation and fibrosis (*Trevizoli et al., 2008*).

Patients with ESRD and HCV infection displayed normal ALT levels. Indeed ALT levels in these patients were significantly lower than those found in patients infected with HCV without renal damage but with similar grades and stages of liver alterations. It has been proposed that the increase in hepatocytes of HCV-infected patients with ESRD who are on chronic dialysis produces a hepatoprotective effect (*Contreras et al., 2007*).

Causes of reduction in ALT activity in these patients are only partially known, such as a reduction in pyridoxal – 5' – phosphate, vitamin B₁₂, coenzymes of

ALT, suppression of AST and ALT synthesis in hepatocytes and an inhibition of AST and ALT released from hepatocytes into the blood stream, as well as the possibility of liver protection by the hepatocyte growth factor, which is higher in patients with chronic renal failure (*Lin et al., 2008*).

Among HD patients, serum ALT levels are elevated in 4-67% patients with positive anti-HCV antibodies, 12-31% of patients with positive HCV-RNA and one third of patients with biopsy proven hepatitis (*Perira and Levey, 1997*).

Shin et al., (2006) reported on two cases after accidental ten times overdose administration of recombinant human erythropoietin (rHu EPO) up to 318,000 units a day in acute myocardial infarction, that the only side effects they found were elevated liver enzymes and hemoglobin levels. These patients were followed up as out patients and elevated enzymes soon normalized.

In *Berglund and Ekblom study (1991)*, that aimed to evaluate the effect of treatment with subcutaneous recombinant human erythropoietin (rHuEPO), 20-40 Iu/kg body weight, 3 times a week, Serum aspartate aminotransferase (AST) and serum alanine aminotransferase (ALT) were unchanged after rHu EPO treatment. We didn't find a statistically significant difference in serum albumin between first and second group ($p = 0.889$), which means that EPO had no effect on serum albumin level.

Rhee and *Erickson (2012)* reported that protein energy malnutrition (PEM) diminishes immunoglobulin (IgA, IgM and IgG) concentrations and cytokine production.

This is not the case in our study as serum albumin is within normal range.

In our study, hemoglobin (Hgb) levels were borderline higher in first group than second group ($p = 0.056$). Also hematocrit (Hct) levels were borderline higher in first group than second group ($p = 0.056$) and this was expected due to administration of EPO in first group.

Khurana et al. (2008) hypothesized that the chronic inflammation as a result of HCV infection or the increased production from the regenerating liver cells causes increased

circulating EPO causing improved Hct in these patients. Also, he reported that hepatitis C patients tend to have higher baseline hemoglobin and decreased need for EPO therapy on dialysis.

Recently, some studies and case reports indicated attenuated anemia in HD patients with HCV infection, and they previously considered this to be related to increased erythropoietin production after hepatic stimulation by chronic infection with hepatitis virus (*Alasran et al., 2009*).

In *Lin et al., study (2008)*, there were increased Hb levels in chronic HCV infected patients with ESRD.

In contrast, *Abdalla et al. (2000)*, reported a higher EPO requirement in HCV positive versus HCV negative patients that was a result of altered iron metabolism induced by chronic infection.

In our study, there was no statistically significant difference in white blood cells in blood ($p = 0.670$) between the two groups, which means that erythropoietin didn't increase white blood cells count above normal, but it only normalized it.

Different circumstances such as chronic renal failure, hemodialysis process, chronic hepatitis C virus infection and various dietary restrictions that we practice with those patients influenced immune system and immunoglobulin production. We didn't find a statistically significant difference in IgM level in serum between first group and second group, which means that EPO didn't influence much IgM level production by stimulated B-lymphocytes by either EPO or HCV infection.

Little is known about the effect of ESRD on B-cell sub-populations (*Pahl et al., 2010*).

The increase in PMNL counts in CKD has been suggested to be a sign of pre-activation. The number of PMNL increasers in relation to the GFR decrease ($P < 0.0001$) PMNL decreases with increasing serum C-reactive protein and IL-6 and decreased albumin, all associated with declining GFR (*Sela et al., 2005*).

In our study this was not the case, as we had lymphopenia but no undernourishment. *Sardenberg et al. (2006)* findings suggest that uremic toxicity plays an essential role in PMN apoptosis and that dialysis may correct or normalize apoptotic rates.

ESRD and especially HD, is associated with B-cell lymphopenia (*Kato et al., 2008*).

HCV infection is associated with leucopenia in HD patients, is as common as in non-HD patients with liver cirrhosis (*Ng et al., 2008*). Chronic hepatitis C virus (HCV) infection is associated with B cell activation, although underlying mechanisms are unclear (*Sugalski et al., 2010*).

This is evidenced by an elevation in serum immunoglobulin isotypes; IgG and its subclasses IgG₁, and IgG₂ and IgM. Mean serum IgM was increased in patients with HCV infection compared with healthy controls (*Lotfy et al., 2006*).

However, it has been documented that Ig levels, serum IgG isotypes and both IgM and IgA production are normal in dialysis patients (*Hauser et al., 2008*).

Starzyk et al. (1993) in their study on 10 patients with chronic renal failure treated with hemodialysis (HD) T and B cell populations were determined in peripheral blood, together with immunoglobulin concentration. There was no significant change in the concentration of IgA and IgM.

We didn't find in our study a significant difference in IgM level between males and females. This was not the case in *Gonzalez et al. (2008)* study who reported that IgM levels are higher in females than in males. Sex differences in immunoglobulins concentrations specifically high IgM levels in females, have been attributed to hormonal effects on B lymphocytes.

IgM didn't show in our study, any significant correlation to any of the measured parameters of the study including erythropoietin dose. To our knowledge, we are the first to study the effect of EPO on IgM level in HCV positive patients on regular hemodialysis.

In a previous study by *Debska-Slizien et al. (2003)*, in order to find the influence of erythropoietin on immunological system of patients with chronic renal failure, it was found that treatment with EPO did not alter plasma immunoglobulin (IgG, IgM and IgA), as well as total count of lymphocytes. In a previous study, by *Costa et al. (2008)*, 50 HD patients, 25 responders and 25 non responders to rHuEPO, were compared to each other and to 25 healthy controls. No statistically significant differences were found between the three groups of

individuals concerning immunoglobulin serum levels (IgG, IgM and IgA).

In a previous study by *Schaefer et al. (1992)*, who studied whether erythropoietin interferes with B cell function and the mechanisms of this effect, IgM production, which appeared to be normal in uremia, remained unchanged.

A retrospective study was done to determine whether rHu EPO treatment modulates the humoral arm of the immune system in MM patients. There was a significant increase in the levels of normal Ig (IgG, IgA or IgM) in response to rHu EPO, during the 3-9 months from treatment initiation *Gadassi et al. (2007)* and *Prutchi-Sagiv et al. (2006)*.

Data indicate a direct stimulant effect of erythropoietin on B- lymphocytes in end-stage renal failure. Production of IgM was enhanced (*Kimata et al., 1991*).

These findings also show that the pharmacologic response to rHuEPO is a function of the dose.

Moreover, these effects were seen in concentrations much higher than that used in our study.

Conclusion:

ESRD with all its restrictions seems to protect patients from increased level of serum IgM due to HCV infection and erythropoietin therapy and subsequent cryoglobulinemia. Further studies at molecular level of B-cell functions are still needed to elucidate the causes of this protection.

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References

- Abdalla A, Owda A, Fedail H et al. (2000):** Influence of hepatitis C virus infection upon parenteral iron and erythropoietin responsiveness in regular hemodialysis patients. *Ren Fail*; 31(5): 349-354.
- Alasran K, Sabry A, Algareeb A, et al. (2009):** Effect of hepatitis C virus on hemoglobin and hematocrit level sin Saudi hemodialysis patients. *Ren Fail*; 31(5): 349-354.
- Awad A, St'uve O, Mayo M, et al. (2011):** Antigliutamic acid decarboxylase antibody-

associated ataxia as extrahepatic autoimmune manifestation of hepatitis C infection: A case report. *Case reports in neurological medicine*, Article ID 975152, 4 pages, doi: 10.1155/2011/975152.

Berglund B and Ekblom B (1991): Effect of recombinant human erythropoietin treatment on blood pressure and some haematological parameters in healthy men. *J Intern Med* 229(2): 125-130.

Bokle BC and Sepp NT (2010): Hepatitis C virus and autoimmunity. *Autoimmune Highlights* 1: 23-35.

Contreras A, Ruiz L, Cruz G, et al. (2007): End stage renal disease and hepatitis C infection: comparison of alanine aminotransferase level and liver histology in patients with and without renal damage. *Annals of hepatology*; 6(1): 48-54.

Costa E, Lima M, Alves JM, et al. (2008): Inflammation, T cell phenotype, and inflammatory cytokines in chronic kidney disease patients under hemodialysis and its relationship to resistance to recombinant human erythropoietin therapy. *J Clin Immunol*; 28: 268-275.

Debska-Slizien A, Rutkowski B, Manitus J, et al. (2003): Influence of erythropoietin on immunological system of patients with chronic renal failure. *Pol Merkur Lekarski* 15(88): 326-7; discussion 327-329.

Diagnostic Automation, Inc (2009): Total human IgM assay. Diagnostic automation; available from <http://www.rapidtest.com/index.php?i=total-human-IgM-ELISA-Kit&id=51&cat=11>. downloaded in 1/1/2011.

Doumas B, Watson W and Biggs M (1971): Albumin standards and the measurement of serum albumin with bromocresyl green. *Clin Chim Acta* 31: 87-96

Fabrizi F, Lunghi G, Ganeshan S, et al. (2007): Hepatitis C virus infection and the dialysis patient. *Seminars in Dialysis*, 20(5): 416-422.

Fazi C, Dagklis A, Cottini F, et al. (2010): Monoclonal B cell lymphocytosis in hepatitis C virus infected individuals. *Cytometry Part B*; 78 B (Suppl 1): 561-568.

Foley RN and Collins AJ (2007): End stage renal disease in the United States: An update from the United States Renal Data System. *J Am Soc Nephrol* 18: 2644-2648.

Gadassi N, Prutchi Sagiv S, Oster HS et al. (2007): Elevated normal immunoglobulin and stable M0protein levels in multiple myeloma patients on erythropoietin treatment. American society of hematology 49th annual meeting December 8-11, Atlanta; GA tiny URL: <http://www.mindcull.com/abstract:e74c2d6a7053dd6822ed3e431628a70d>.

Geara AS, Castellanos MR, Bassil C et al. (2010): Effects of parathyroid hormone on immune function. *Clinical and Developmental Immunology*, Volume 2010. Article ID 418695, 10 pages, doi: 10.1155/2010/418695.

Gonzalez A, Alende G, Gude F, et al. (2008): Serum levels of immunoglobulins (IgG, IgA, IgM) in a general adult population and their relationship with alcohol consumption, smoking and common metabolic abnormalities. *Clinical and experimental immunology*, 151: 42-50.

Haag-Weber M and Morl WH (1996): Dysfunction of polymorphonuclear leucocytes in uremia. *Semin Nephrol*, 16(3): 192-201.

hauser AB, STinghen AE, Kato S, et al. (2008): Characteristics and causes of immune dysfunction related to uremia and dialysis. *Peritoneal Dialysis International* 28(3): S1832-S187.

Ito M, Kusunoki M, Mochida K, et al. (2011): HCV infection and B-cell lymphomagenesis. *Advances in hematology*, Article ID 835314, 8 pages.

Jang JY and Chung RT (2010): New treatments for chronic hepatitis C. *Korean J Hepatol* 16: 263-277.

Kalantar-Zadeh K and Aronoff G (2009): Hemoglobin variability in anemia of chronic kidney diseases. *J Am Soc Nephrol*; 20: 479-487.

Kato S, Chmielewski M, Honda H, et al. (2008): Aspects of immune dysfunction in end stage renal disease. *Clin J Am Soc Nephrol*; 3: 1526-1533.

Khurana A, Nickel Ae, Narayanan M, et al. (2008): Effect of hepatitis C infection on anemia in hemodialysis patients. *Hemodialysis international*; 12: 94-99.

Kiechl S, Lorenz E, Reindl M, et al. (2002): Toll-like receptor 4 polymorphisms and atherogenesis. *N Engl J Med* 347: 185-192.

Kimata H, Yoshida A, Ishioka C, et al. (1991): Human recombinant erythropoietin directly stimulates B cell immunoglobulin production and proliferation in serum free medium. *Clinical and experimental immunology*; 85: 151-156.

Lifshitz L, Tabak G, Gassman M, et al. (2010): macrophages as novel target cells for erythropoietin. *Haematologica* 2010; 95(11): 1823-1831.

Lim WH Kireta S, Leedham E et al. (2007): Uremia impairs monocyte and monocyte-derived dendritic cell function in hemodialysis patients. *Kidney Int.*, 42: 1138-1148.

Lin Y, Lin C, Lee C, et al. (2008): Chronic hepatitis ameliorates anemia in hemodialysis patients. *Nephrology*; 13: 289-293.

Liwoska KA, Bryl E, and Witkowski JM (2011a): Erythropoietin receptor is detectable on peripheral blood lymphocytes and its expression increases in activated T lymphocytes. *Haematologica* 201196(03): e13-e13.

- Liwoška KA, Jaswilewicz A, Birył E et al. (2011b):** Erythropoietin as an immunomodulating agent. *Nephro-Urol Mon* 3(4): 247-251.
- Lofly M, El-Kady IM, Nasif WA, et al. (2006):** Distinct serum immunoglobulins pattern in Egyptian patients with chronic HCV infection analyzed by nephelometry. *Journal of Immunoassay and immunochemistry*, 37(1): 03-114.
- Ng YY, Lin CC, Wu SC, Hwang SJ, Mo CH, Yang WC, and Lee SD (2002):** Leukopenia and thrombocytopenia in hemodialysis patients with hepatitis B or C virus infection and non hemodialysis patients with hepatic cirrhosis. *Clin Nephrol*; 57(4): 289-95.
- Pahl MV, Sastry, Sepassi L et al. (2010):** Effect of end stage renal disease on B-lymphocyte subpopulations, IL-7, BAFF and BAFF receptor expression *Nephrol Dial Transplant* 25: 205-212.
- Perira BJ and Levey AS (1997):** hepatitis C virus infection in dialysis and renal transplantation. *Kidney Int*; 51: 981-999.
- Prutchi-Sagiv S, Grolishevsky N, Oster HS, et al. (2006):** Erythropoietin treatment in advanced multiple myeloma is associated with improved immunological functions: Could it be beneficial in early disease? *British Journal of Haematology*; 135: 660-672.
- Prutchi-Sagiv S, Neumann D and Mittleman M (2005):** Erythropoietin as an immunotherapeutic agent: new uses for an old drug? *The Journal for Innovative Ideas in Biomedical Research*; 2: 587-596.
- Rhee J. and Erickson T. , (2012):** Erythropoietin stimulant and other blood doping methods, Ch 17, *Medical Toxicology of drug abuse: synthesized chemical and psychoactive plants* , by Donald G. Barceloux. John & Wiley sons , published 2012.
- Sardenberg C., Suassina P. , Andreoli MMC., et al. (2006):** Effects of uremia and dialysis modality on polymorphonuclear cell apoptosis and function. *Nephrol Dial Transplant*; 21(1): 160-165.
- Schaefer RM, Paezek L, Berthold G, et al. (1992):** Improved immunoglobulin production in dialysis patients treated with recombinant erythropoietin. *Int J Artif Organs*; 15(4): 204-8.
- Schumann G and Klauke R (2003):** New IFCC reference procedures for the determination of catalytic activity concentrations of five enzymes in serum: preliminary upper reference limits obtained in hospitalized subjects. *Clin Chim Acta*, 327: 69-79.
- Shin DM, Kwon YI, Choi SI, et al. (2006):** Accidental ten times overdose administration of recombinant human erythropoietin (rHu Epo) up to 318,000 units a day in acute myocardial infarction: report of two cases. *Basic Clin Pharmacol. Toxicol*; 98(2): 222-224.
- Sela S, Shurtz-Surirski R, Cohen-Mazor M, et al. (2005):** Primed peripheral polymorphonuclear leukocytes: a culprit underlying chronic low grade inflammation and systemic oxidative stress in chronic kidney disease. *J Am Soc Nephrol*; 16(8): 2431-2438.
- Starzyk J, Sarnecka S, Bartelik S, et al. (1993):** Subpopulations of T and B lymphocytes, rosette tests, levels of immunoglobulins in peripheral blood and NBT test in the initial period of hemodialysis conducted with cuprophan or cellulose acetate dialysers in patients with chronic renal failure. *Wiad Lek* 46(21-22): 817-823.
- Sugalski JM, Rodriguez B, Mori S, et al. (2010):** Peripheral blood B cell subset skewing is associated with altered cell cycling and intrinsic resistance to apoptosis and reflects a state of immune activation in chronic hepatitis C virus infection. *J Immunol* 185: 3019-3027.
- Tabacco A, Meattini F, Moda E, et al. (1979):** Simplified enzymic/colorimetric serum urea nitrogen determination. *Clin Chem* 25: 336-7.
- Trevizoli JE, de Paula MR, Ribeiro LF, et al. (2008):** Hepatitis C is less aggressive in hemodialysis patients than in nonuremic patients. *Clin J Am Soc Nephrol* 3: 1385-1390.
- William J, Saad N, Salib M et al. (1998):** The acute effect of intravenously administered recombinant human erythropoietin on the immune response of uremic patients maintained on regular hemodialysis. *Artif Organs* 22(3): 192-196.
- Yatzidis H (1974):** New method for direct determination of true creatinine. *Clinical Chemistry*, 20(9): 1131-1134.

دراسة تأثير الإريثروبوتين على مستويات الأجسام المضادة غلوبولين المناعي إم فى المرضى المصابين بعدوى فيروس الكبدى سى المزمن والمعاشين على الديال الدموى

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كل من مرض البولينا والديال الدموى يسبب إحباط للمناعة فى المرضى المصابين بالفشل الكلوى المزمن والمعاشين على الديال الدموى .

فقد كانت هناك دراسات تدل على زيارة الأجسام المضادة الإيمونوجلوبولين المناعى G و M ومستوياتهم فى دم المرضى المصابين بفيروس الكبدى سى مقارنة بالمجموعات الضابطة من الأصحاء

هناك أدلة تؤكد أن هناك تأثير مباشر لعقار الإريثروبوتين على خلايا بي المناعية . كميات كبيرة من عقار الإريثروبوتين تسببت فى زيادة إنتاج الإيمونوجلوبولين المناعى فى الأبحاث المعملية كما تسببت فى تكاثر كل أنواع البلازما وكذلك خلايا البلازما البشرية المنتجة معملياً والتي تنتمى إلى مجموعة بي من الخلايا المناعية والتي تنتج الأجسام المضادة .

• المرضى والوسائل :

هذه الدراسة تم إجراؤها فى وحدة الغسيل الكلوى بمستشفى شبرا العام فى الفترة ما بين أغسطس 2010 إلى فبراير 2011 . تم عمل الدراسة على 30 من مرضى الديال الدموى المنتظم والمصابين بفيروس الكبدى سى . تم استخدام سائل البيكربونات فى الغسيل الكلوى لهؤلاء المرضى وكذلك مرشحات البولى سالفون ، لمدة 4 ساعات ثلاث مرات أسبوعياً .

• تم تقسيم المرضى إلى مجموعتين :

■ المجموعة الأولى : 15 مريض يستخدمون عقار الإريثروبوتين فى حدود جرعة 4000 وحدة دولية فى الأسبوع .

■ والمجموعة الثانية : 15 مريض لا يستخدمون عقار الإريثروبوتين لكل المرضى تم عمل الآتى : فحص طبي كامل ، صورة دم كاملة ، بولينا فى الدم ، كرياتينين فى الدم ، إنزيمات كبدية فى الدم AST و ALT ، نسبة الألبومين فى الدم ومستوى إيمونوجلوبولين المناعى إم باستخدام ELISA (إختبار كمى) .

• النتائج :

لم يكن هناك فرق واضح إحصائياً بين مجموعتى الدراسة فيما يتعلق بالآتى : السن ، توزيع جنس المرضى ، عدد كرات الدم البيضاء ، ALT و AST ، كرياتينين فى الدم بولينا فى الدم ومستوى إيمونوجلوبولين المناعى إم فى الدم . المجموعة الأولى أظهرت فرقاً شبه أعلى عن المجموعة الثانية بخصوص نسبة الهيموجلوبين ونسبة الهيماتوكريت فى الدم .

• الإناث لم يكن لديهم فى دراستنا مستوى من إيمونوجلوبولين إم المناعى فى الدم أعلى من الذكور كل العلاقات بين الإيمونوجلوبولين المناعى إم والقياسات الأخرى بالدراسة لم تسفر عن علاقات إيجابية مباشرة أو سلبية لها أهمية إحصائية .

• الإستنتاج :

فيما يبدو أن ارتفاع نسبة البولينا فى الدم تقوم بحماية مرضى الفشل الكلوى المزمن والمعاشين على الديال الدموى من التعقيدات التى تصاحب إلتهاب الكبدى سى وكذلك من تأثير الإريثروبوتين على الأجسام المناعية (إيمونوجلوبولين) المناعى فى الدم ، وكلا المؤثرين يؤديان إلى زيادة إيمونوجلوبولين المناعى أم فى الدم .